

Dov Prusky



**Department of Postharvest Science of
Fresh Produce
The Volcani Center, Agricultural
Research Organization**

1st Lecture Fruit Pathology

Subjects of Study

1. Development of postharvest disease after harvest and the factors modulating their development (1.5h)
2. Different mechanism for disease resistance of fruits to postharvest diseases (2-3 h)
3. Mechanism of pathogen attack after harvest, microbiomes (3h).
4. Mycotoxins in fruits (1-2h)
5. Physical and biological factors affecting postharvest diseases (2h)
6. Disease control (4h)
7. Resistance to Fungicides (1h)
8. Biological control (1-1.5)

Objectives of Storage

- To keep the freshness of the produce
- To prevent senescence processes
- To prevent water loss
- To inhibit ripening
- To inhibit pathogen development

However as we increase the length of the storage
there is a decrease in quality and increase of
pathogen development

Postharvest “life” of harvested produce

The harvest indicate the end of the period of growth and the start of the period of senescence

- Start of biochemical changes
- Effect on storage length



Initiation of the pathogen colonization

What are the factors affecting the losses of harvested produce by pathogens?

Before Harvest

- Weather conditions
- Level of maturity
- Location of the fruit on the tree
- Sensitivity to wounding's
- Harvesting conditions

After harvest

- Conditions of the packing house
- Treatment at the packinghouse
- Humidity during storage
- Transport conditions

Quality decline of harvested produce: Where does it take place?

1. In the orchard: weather conditions, hot weather, ripening conditions
2. During harvest: way of harvest, time to transfer to the packing house, transport to the packing house
3. In the packing house and postharvest handling: quality of the packinghouse, cold storage, humidity
4. Transport to international ports: cold storage, level of oxygen, level of CO₂.

Very complicated system !!!

What are the factors affecting the losses of harvested produce?

Before Harvest

- Weather conditions
- Level of maturity
- Location of the fruit on the tree
- Sensitivity to wounding's
- Harvesting conditions

After harvest

- Conditions of the packing house
- Treatment at the packinghouse
- Humidity during storage
- Transport conditions
- Temperature of storage

Too many factors are affecting the produce quality and disease development!!!

**The objective !!!
Is to improve of
quality by prevention
of decay
development during
storage**

What is the source of the inoculum for host infection?

Before harvest:

1. Air (Botrytis, Penicillium)
2. Water, Humidity (Colletotrichum)
3. Soil remnants (Phytophthora)
4. Agricultural tools

Direct penetration and quiescence: Botrytis



Direct penetration +
quiescent symptoms

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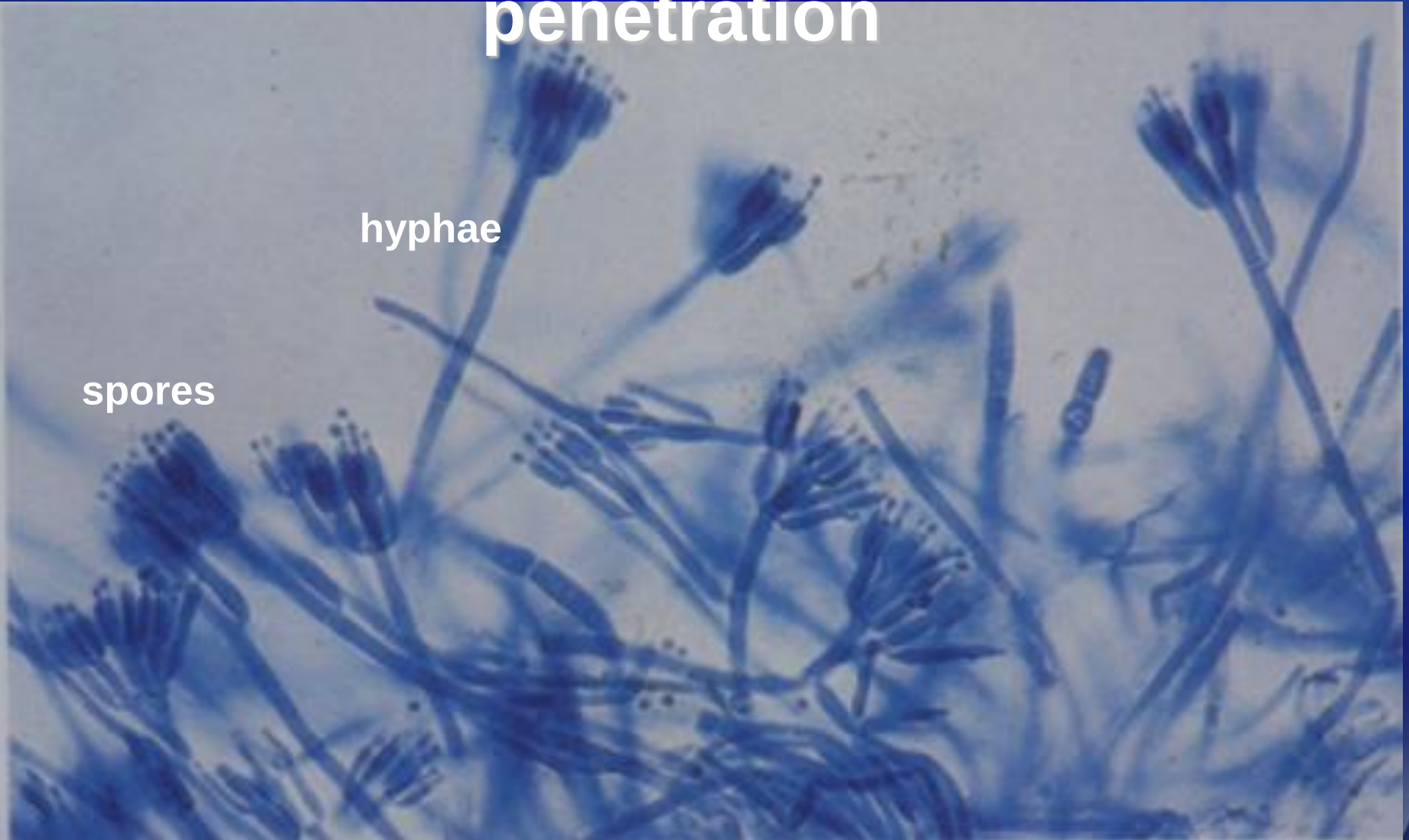
Green mold rot of citurs cause by *Penicillium digitatum* by wound penetration



Penicillium side peel wound penetration

hyphae

spores

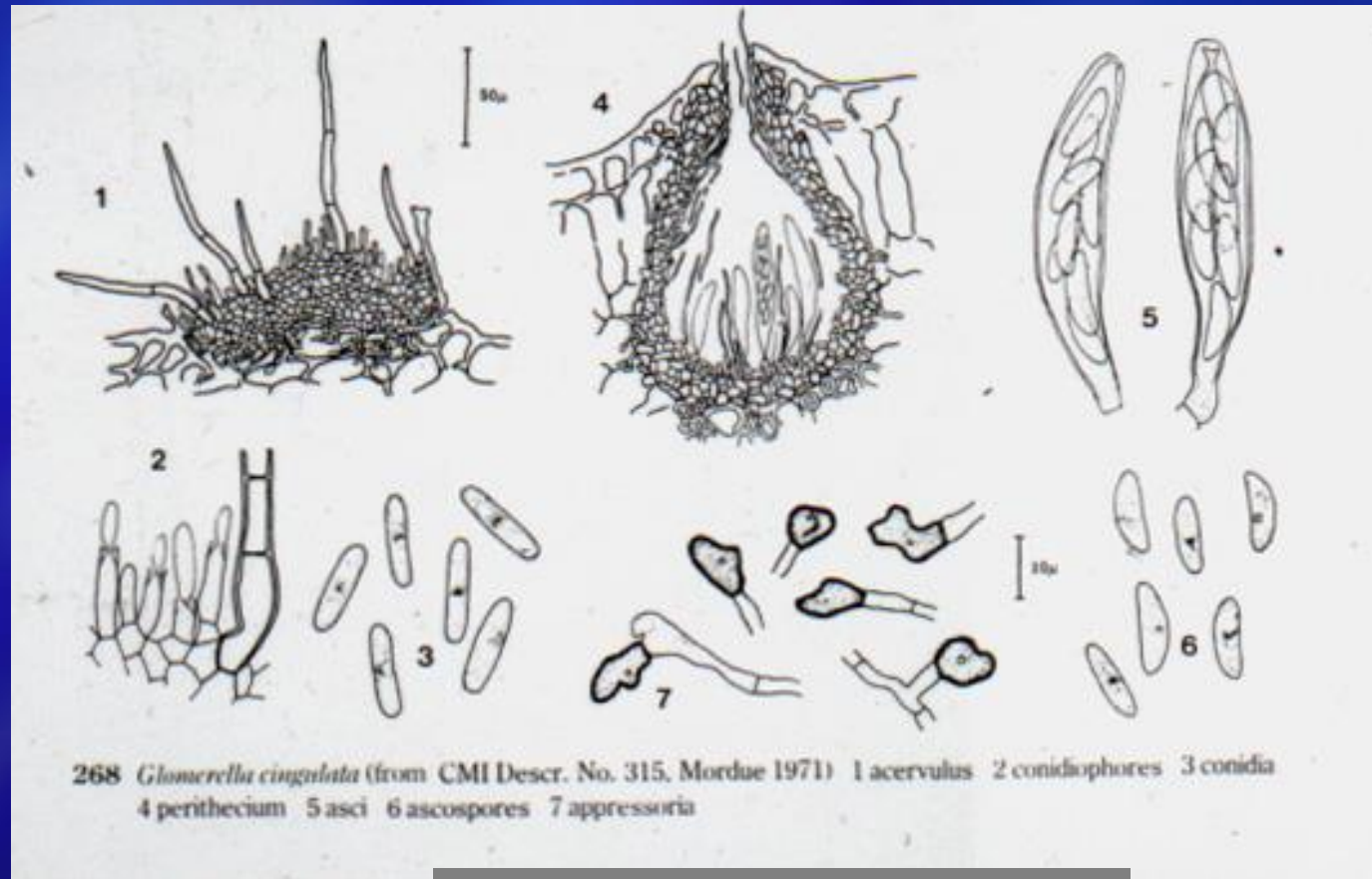


What is the source of the inoculum for host infection?

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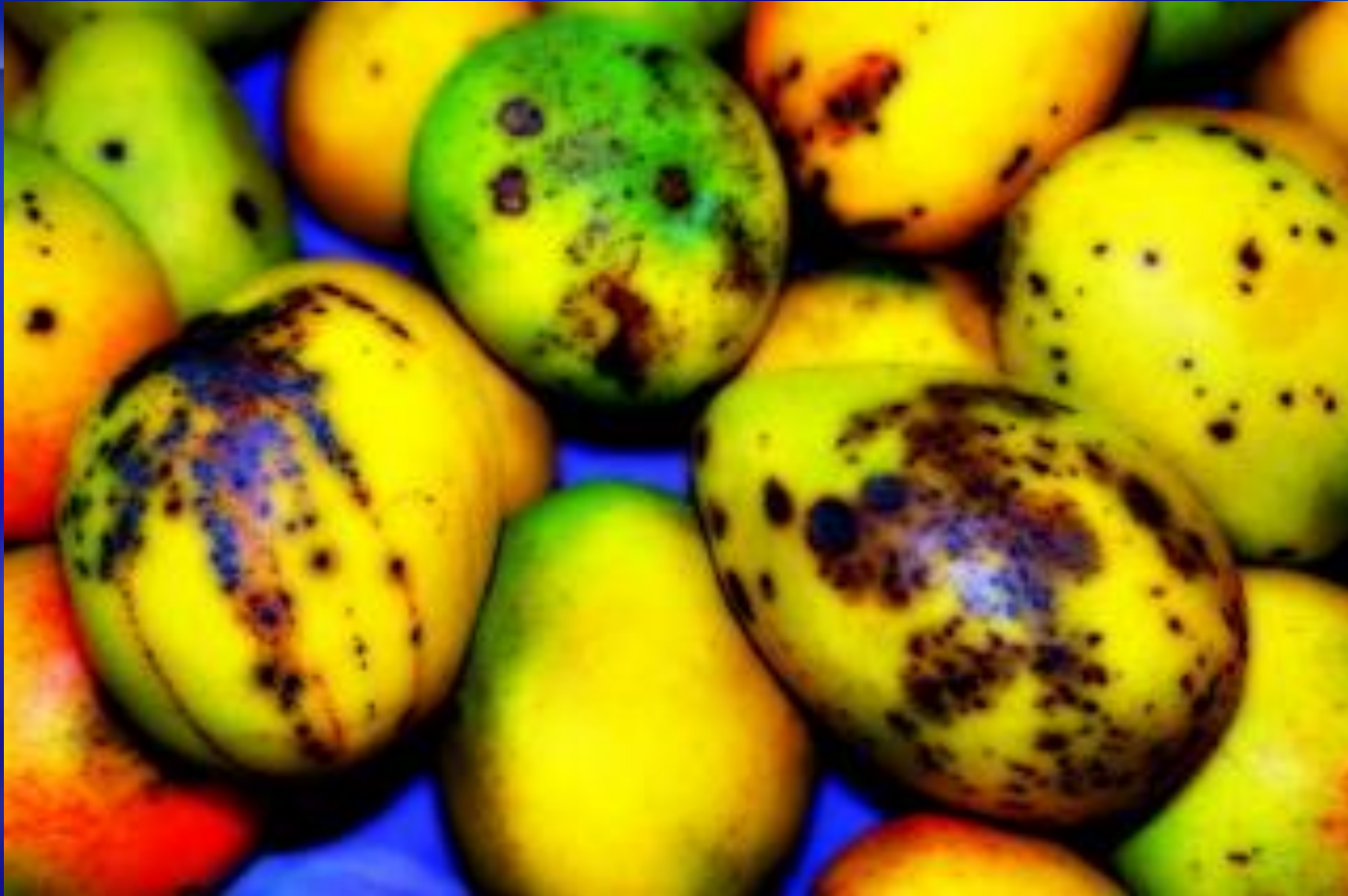
1. Air (Botrytis, Penicillium)
2. Water, Humidity (Colletotrichum)
3. Soil remnants (Phytophthora)
4. Agricultural tools

Colletotrichum the pathogen the cause Anthracnose



Direct penetration + no
symptoms

Humidity run off and active infections



**Colletotrichum (Anthracnose)
in Mango fruits**

What is the source of the inoculum for host infection?

Before harvest:

1. Air (Botrytis, Penicillium)
2. Water, Humidity (Colletotrichum)
3. Soil remnants (Phytophthora)
4. Agricultural tools

Phytophthora infestans



Direct penetration + small
symptoms

What is the source of the inoculum for host infection?

Before harvest:

1. Air (Botrytis, Penicillium)
2. Water, Humidity (Colletotrichum)
3. Soil remnants (Phytophthora)
4. Agricultural tools

Specificity of the host: how important?

Specificity of the colonization is dependent in several factors:

- 1. Capability to breach the cuticle**
- 2. Direct penetration or wound penetration**
- 3. Nutritional needs**
- 4. Enzyme production**
- 5. Toxin accumulation**
- 6. Genetic background**

Specificity of the pathogen to attack the host:

- Monophage- *Penicillium*
- Polyphage- *Alternaria*, *Botrytis*,
Rhizopus

Penicillium side peel wound penetration



Specificity of the pathogen to attack the host:

- Monophage- *Penicillium*
- Polyphage- *Alternaria*, *Botrytis*, *Rhizopus*

Direct penetration and quiescence: Botrytis

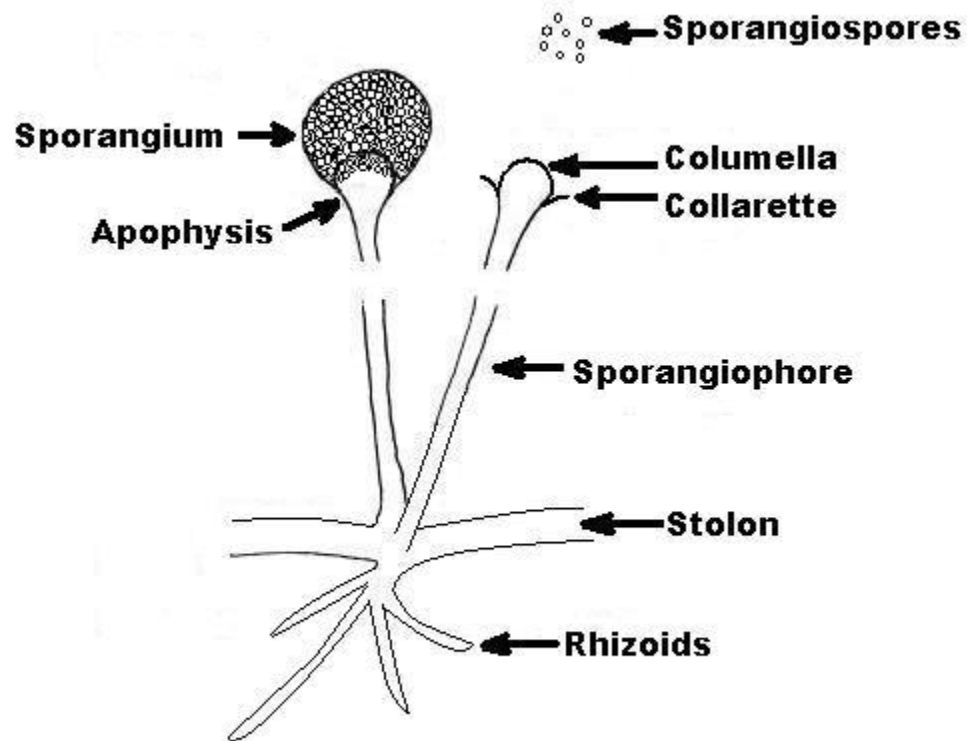


Direct penetration +
quiescent symptoms



Alternaria alternata

Rhizopus



Specificity of the pathogen to attack the host:

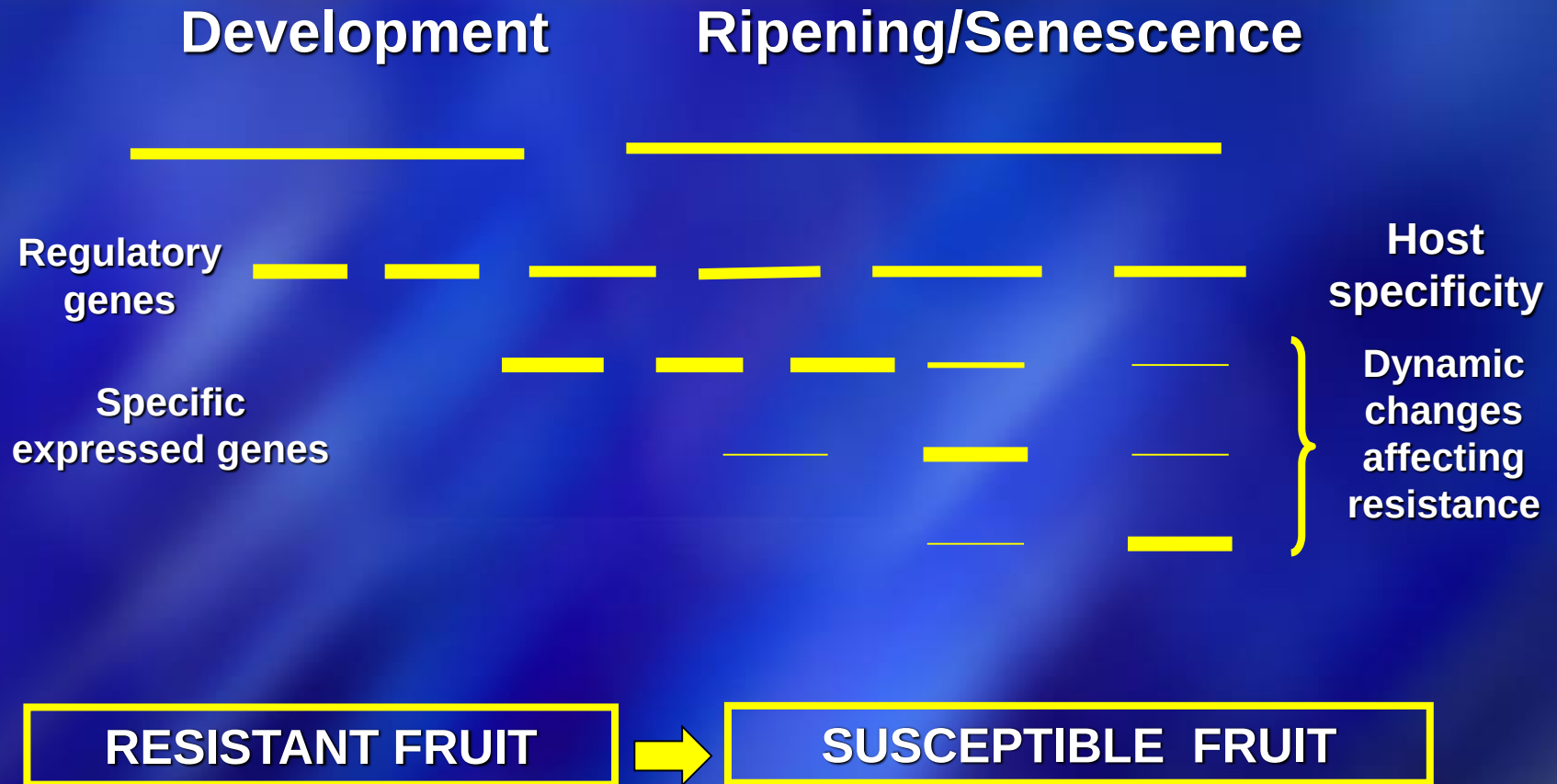
- Monophage- *Penicillium*
- Polyphage- *Alternaria, Botrytis, Rhizopus*

Defense or pathogenicity in fruit pathogen interactions

1. Basic defense lines

**2. Hierarchical response line
during colonization**

Mechanisms of host responses in fruit pathogen interactions



FRUIT AND PATHOGEN INTERACTION

FRUIT
RESISTANCE

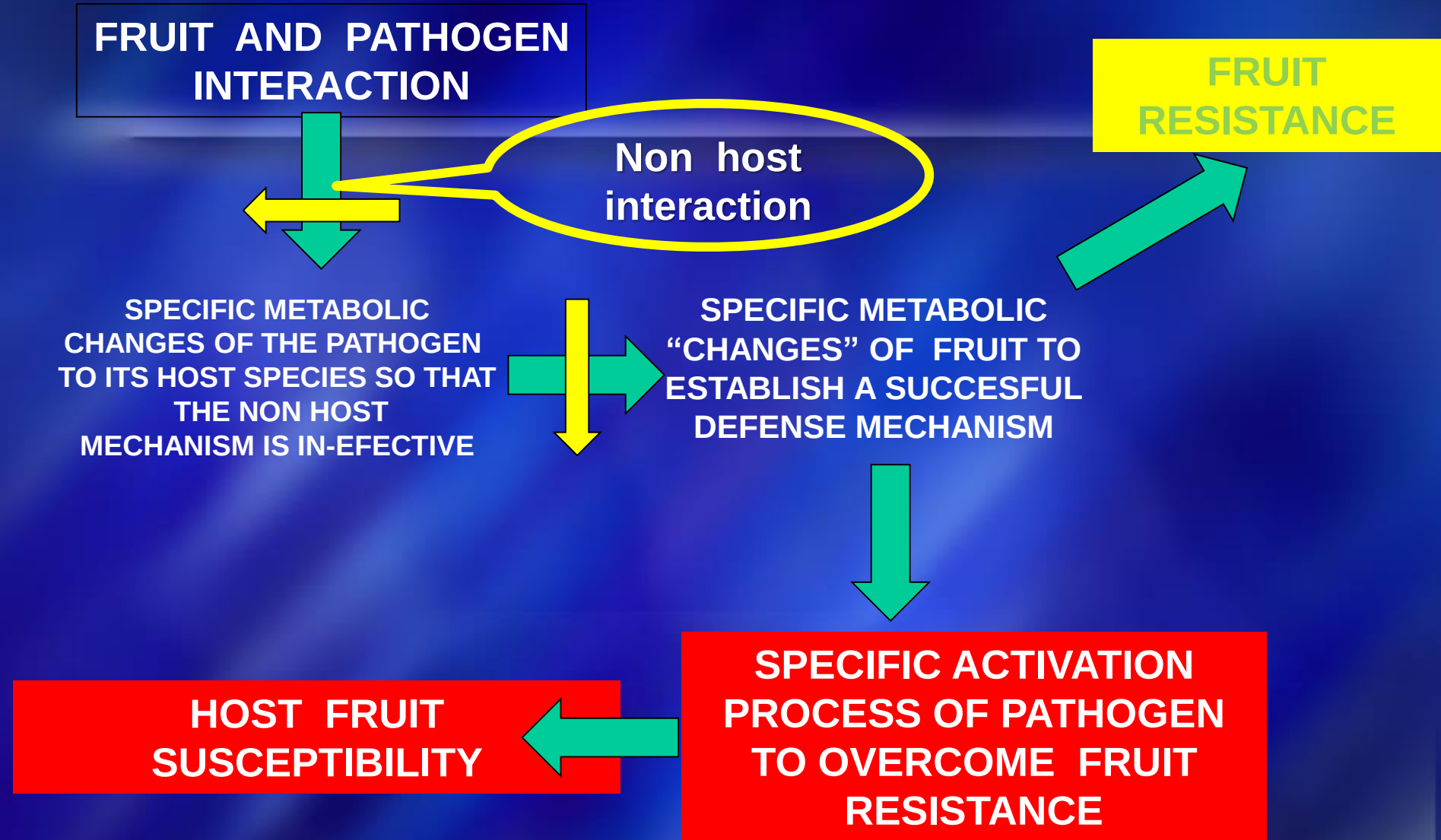
Non host
interaction

SPECIFIC METABOLIC
CHANGES OF THE PATHOGEN
TO ITS HOST SPECIES SO THAT
THE NON HOST
MECHANISM IS IN-EFFECTIVE

SPECIFIC METABOLIC
“CHANGES” OF FRUIT TO
ESTABLISH A SUCCESSFUL
DEFENSE MECHANISM

HOST FRUIT
SUSCEPTIBILITY

SPECIFIC ACTIVATION
PROCESS OF PATHOGEN
TO OVERCOME FRUIT
RESISTANCE



Fungal specificity

PATHOGEN	DISEASE	PRIMARY HOST	
<i>Penicillium italicum</i>	Blue mold	Citrus	Mono
<i>Phytophthora infestans</i>	Late Blight	Potato, tomato	Mono
<i>Penicillium spp</i>	Blue mold	Tomato, cucumber, melon	Poly
<i>Phomopsis</i>	Stem end	Citrus, mango, avocado	Poly
<i>Rhizopus stolonifer</i>	Soft Rot	Stone and pome fruit, grape, avocado, papaya, strawberry, raspberry, cherry, tomato, pepper, Eggplant, carrot, melon, pea, bean sweet potato	Poly

Conditions for fungal germination, penetration and colonization on horticultural crops:

- Optimal temperature 20-25 C
- Optimal relative humidity 90-95%
- Concentrations of O₂ higher than 1%
- Concentration of CO₂ lower than 15-20%
- pH range between 4-6
- Presence of nutritional factors (glucose, sucrose, fructose in the wound)
- Adhesion

Nutritional factors in sweet potato colonized by *Rhizopus*



Rhizopus oryzae germination and growth after supplementation with Sweet Potato Juice active Fraction (SPAF) and the whole Extract (SPJE)

Sweet potato juice extract
SPJE(mg/ml).

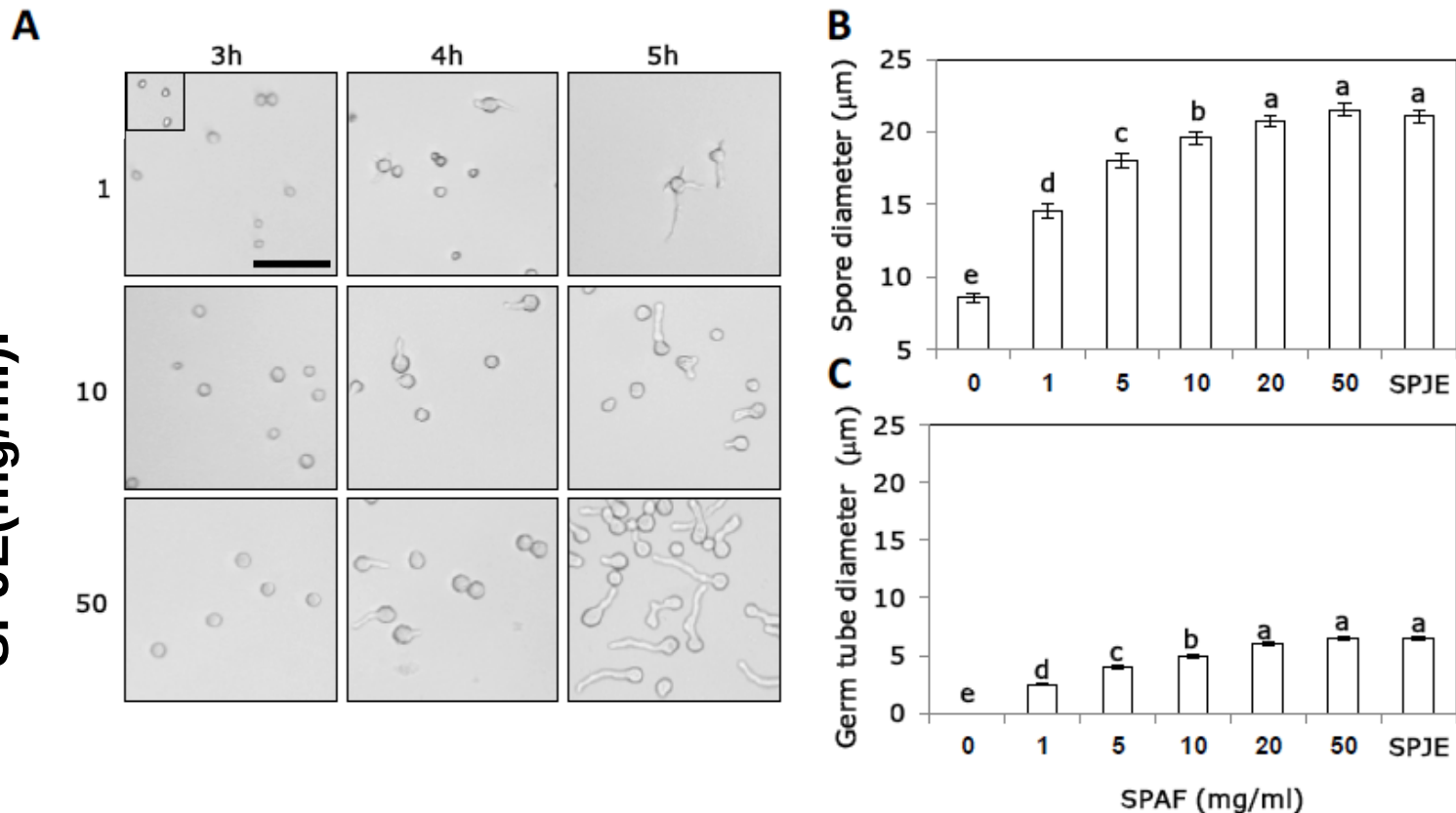
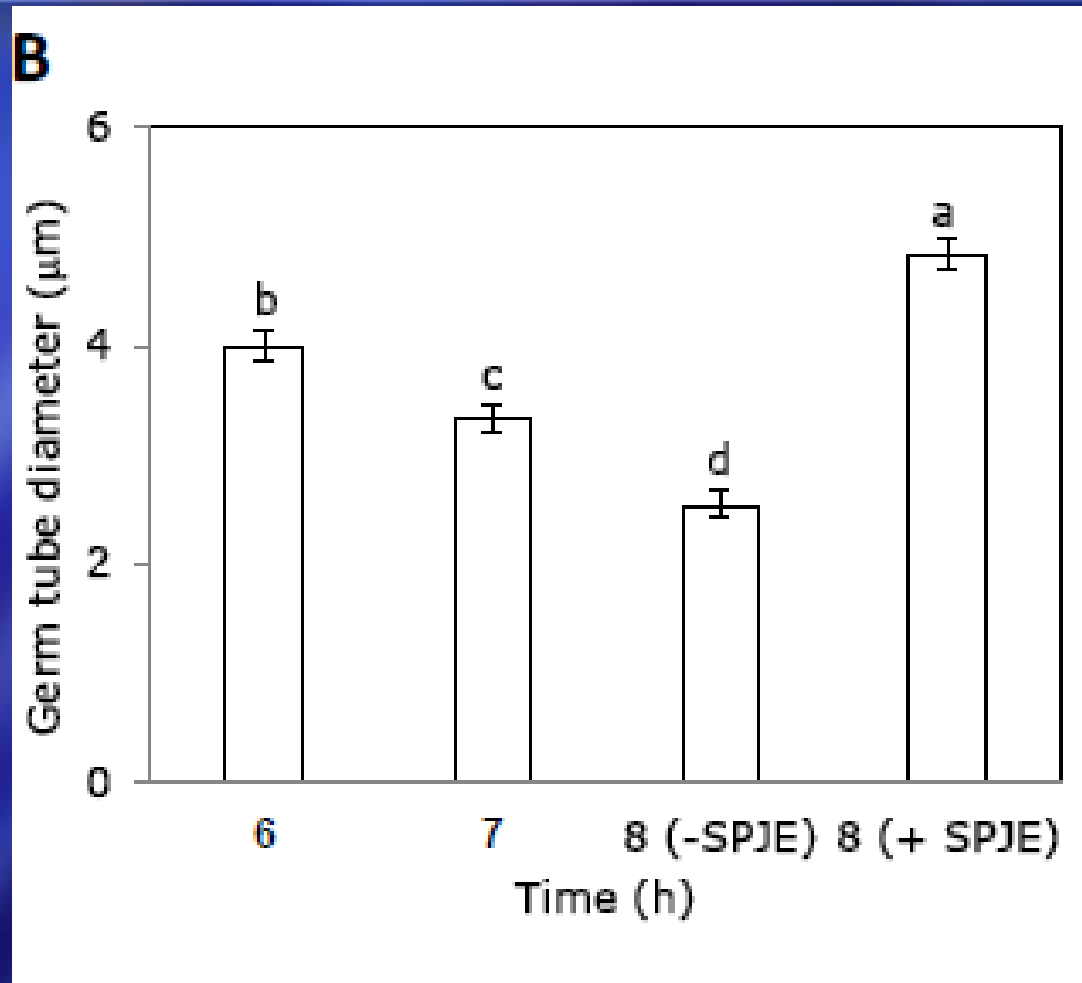


Fig. 2. Dose response of *R. oryzae* spores to sweetpotato active fraction (SPAF) as compared to sweetpotato juice extract (SPJE). A, Micrographs of spores after different incubation intervals (inset shows water-incubated spores, bar = 100 μm). B, Spore diameter after 5 h of incubation. C, Germ-tube diameter after 8 h incubation. Values are means $\pm$ SE (n = 100). Different lowercase letters above the bars denote significant differences ($P < 0.05$).

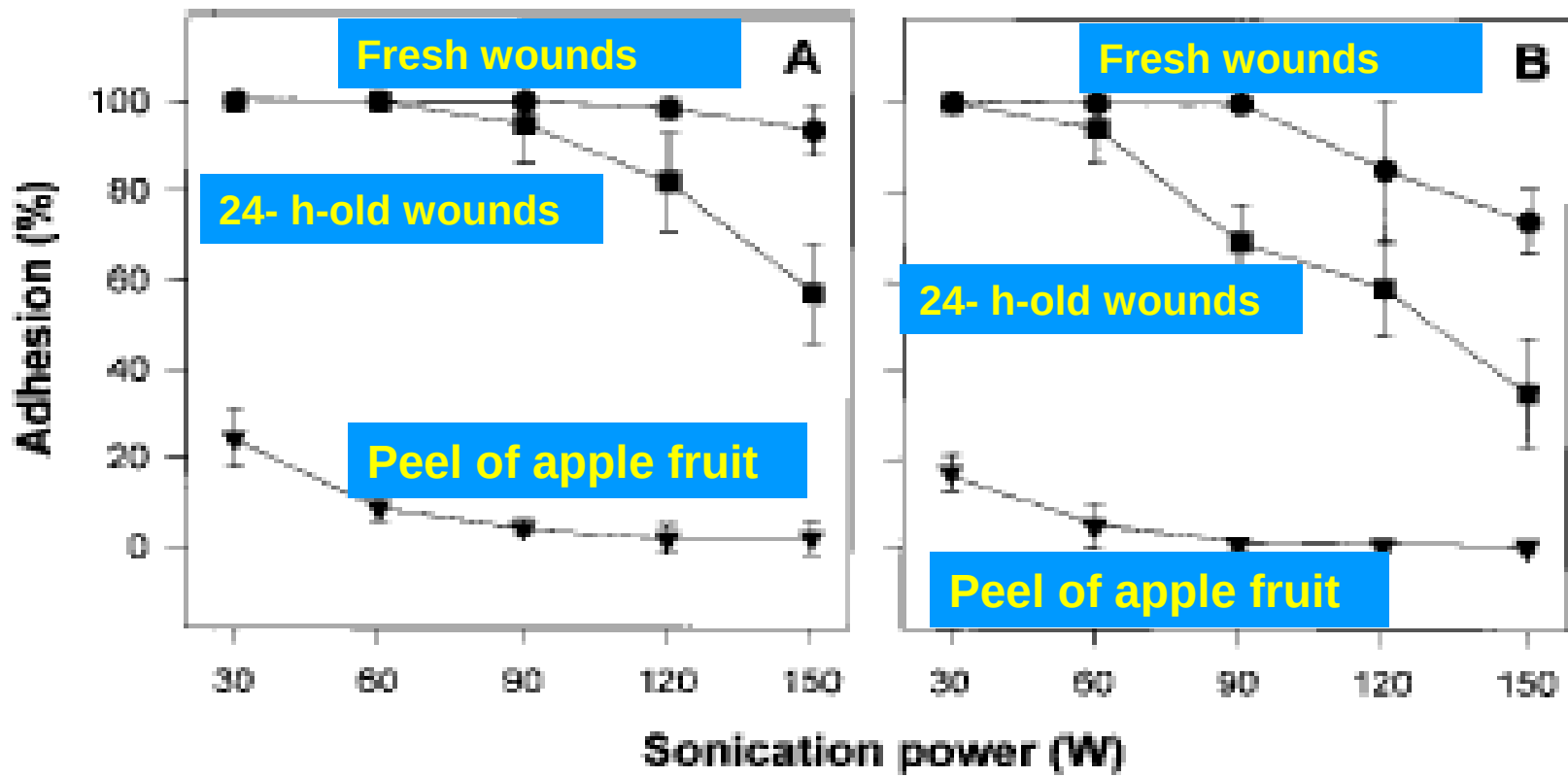
***Rhizopus oryzae* germ-tube shrinkage as a result of starvation and recovery after supplementation with sweetpotato juice extract (SPJE).**



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- **Adhesion**

Percent adhesion of (A) *Botrytis* and (B) *Penicillium* conidia on apple fruit:



Botrytis cinerea

Penicillium expansum

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- **Adhesion**
- **Fruit volatiles**

Effect of volatiles

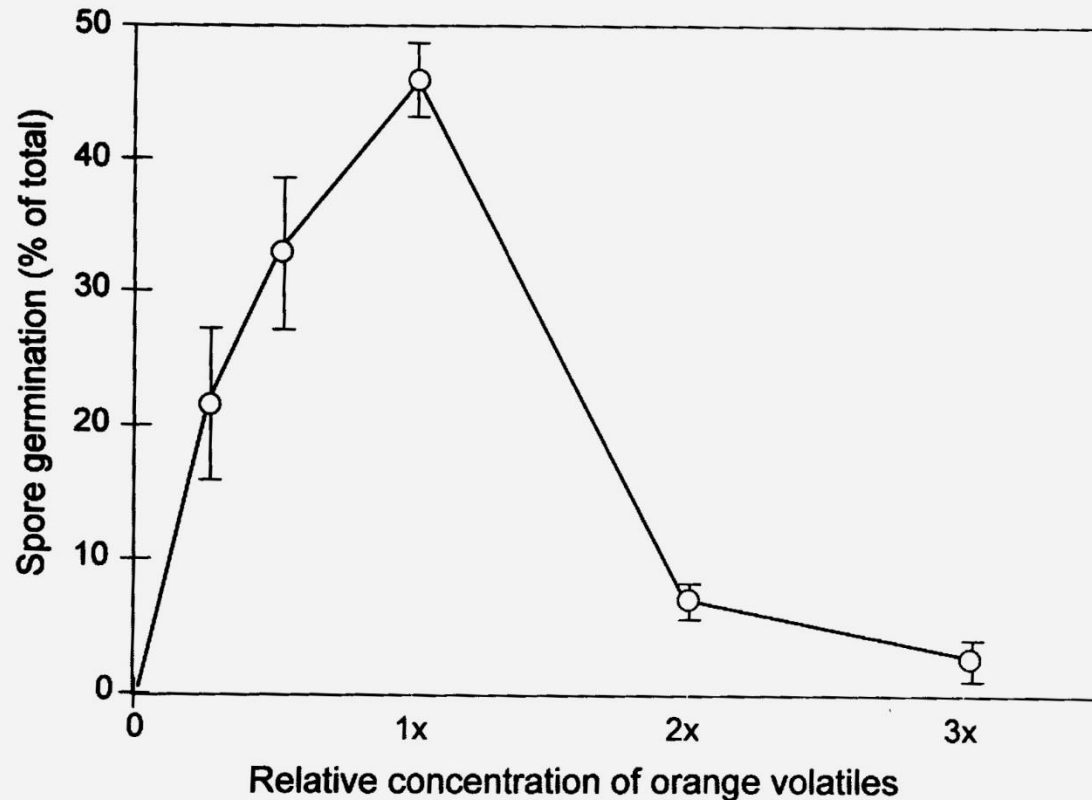
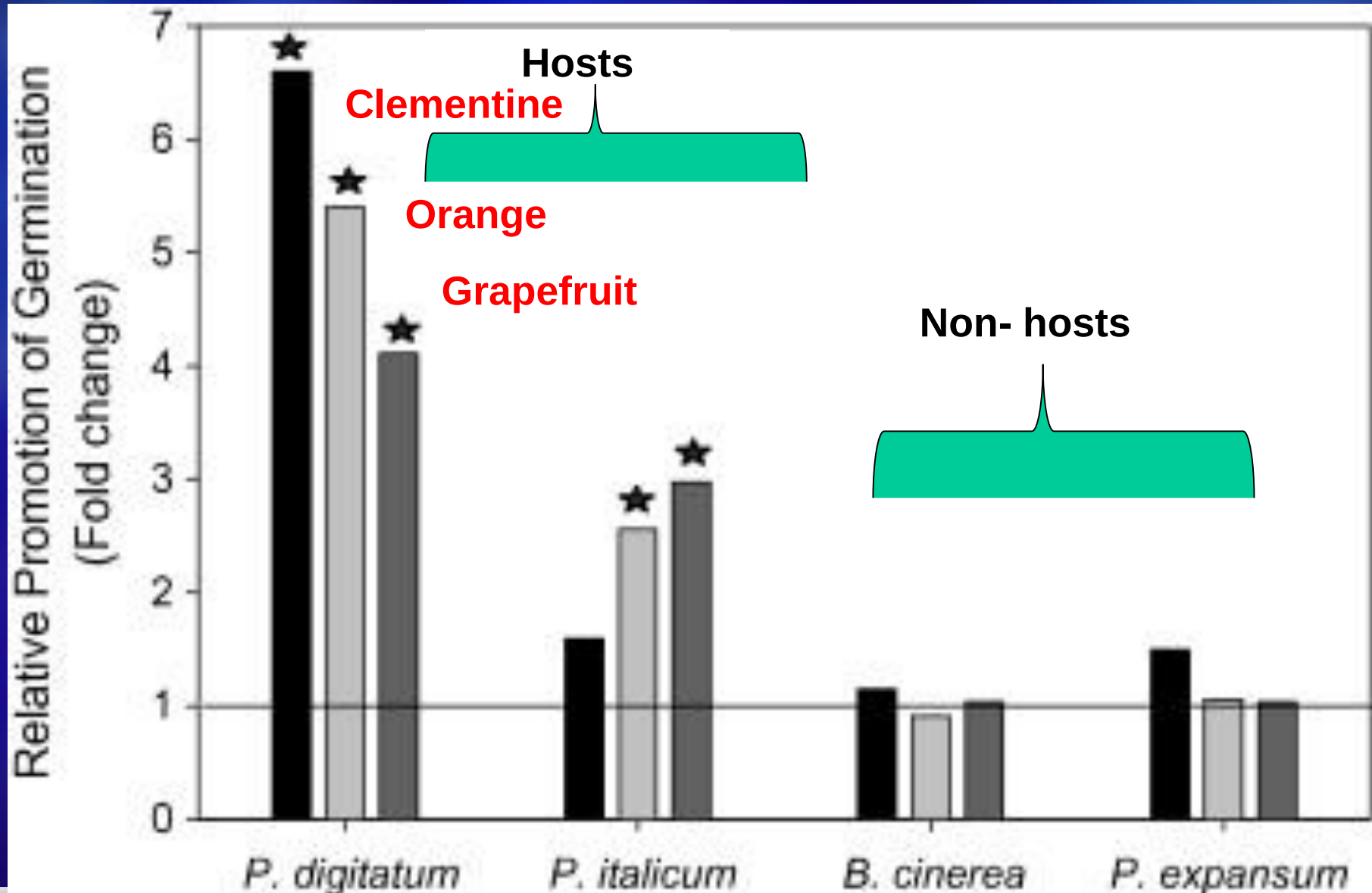
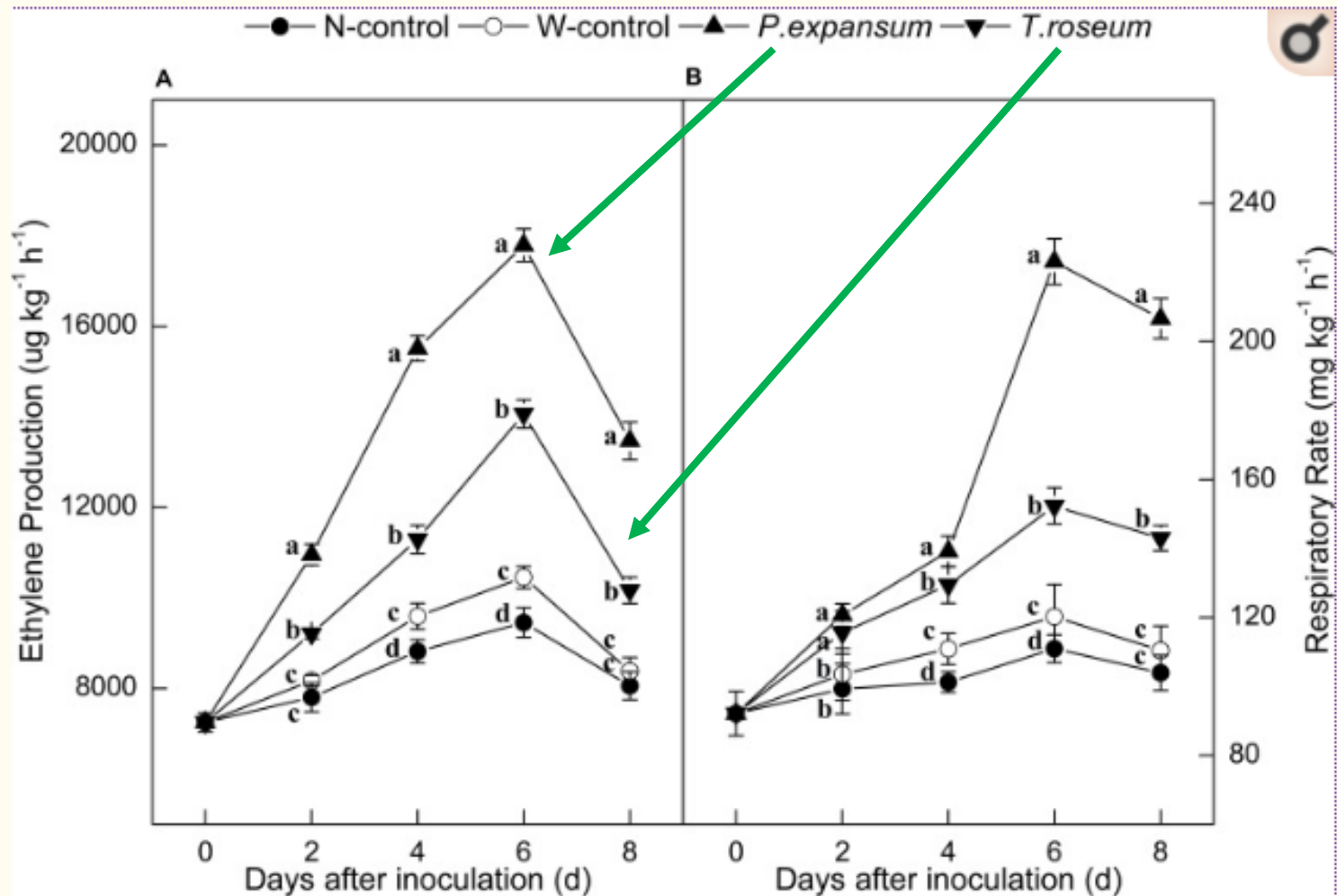


Fig. 3. Effect of concentration of the synthetic mixture of volatile compounds on the germination of *Penicillium digitatum* spores. 1x is the concentration typical of the natural mixture measured surrounding wounded oranges. Bars indicate standard errors. (Reproduced from Eckert and Ratnayake, 1994 with permission of the American Phytopathological Society).

The effect of volatiles released from peel discs of clementine, orange and grapefruit germination of *P. digitatum*, *P. italicum*, *B. cinerea* and *P. expansum*



Effect of *Penicillium expansum* and *Trichothecium roseum* on ethylene production and respiration of apple fruit storage



Conditions for fungal germination, penetration and colonization on horticultural crops:

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- **Adhesion**
- **Fruit volatiles**

Penetration of postharvest pathogens

Spread of spores

- Air movement
 - Water, water splash, humidity run off
 - Mechanical wounds
 - Direct contact
-
- **Direct penetration**
 - **Penetration through wounds**

Penetration of postharvest pathogens

Direct penetration :

Showing no Symptoms- Quiescent infections by producing appressoria for penetration

- *Botrytis cinerea*
- *Colletotrichum gloeosporioides*

Showing small initial symptoms:

- *Phytophthora infestans* in potato
- *Phytophthora citrophthora* in citrus

Direct penetration and quiescence: Botrytis



Direct penetration +
quiescent symptoms

Direct penetration of *Botrytis* penetrating plum and nectarine fruits

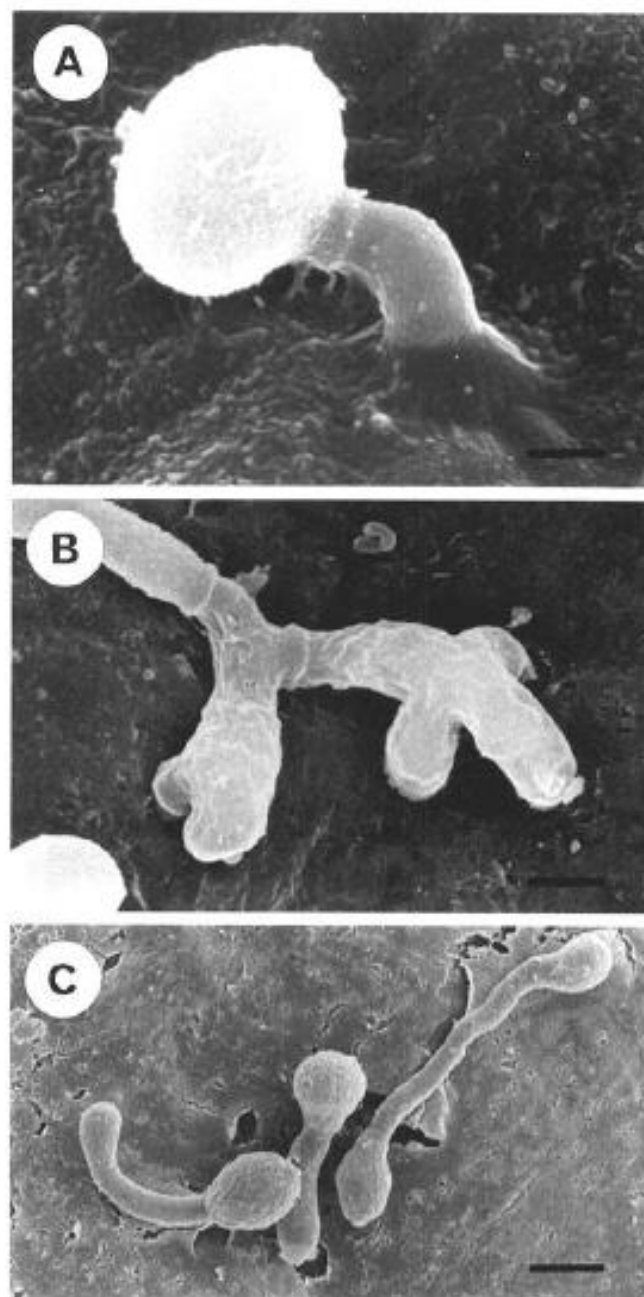


Fig. 1. Scanning electron microscope micrograph of appressoria formed by *Botrytis cinerea* on plum and nectarine fruit. **A**, Protoappressorium on plum 9 h after inoculation. Scale bar = 3.3 μm . **B**, Lobate appressoria on nectarine 48 h after inoculation. Scale bar = 4 μm . **C**, Simple, hyaline appressoria on nectarine 24 h after inoculation. Scale bar = 9 μm .

**Quiescent infection defined by Prusky (1996),
is :**

**A latent, or dormant parasitic relationship in
which the pathogen spends long periods
during the host's life in a quiescent stage
until, under specific circumstances, it
becomes active.**

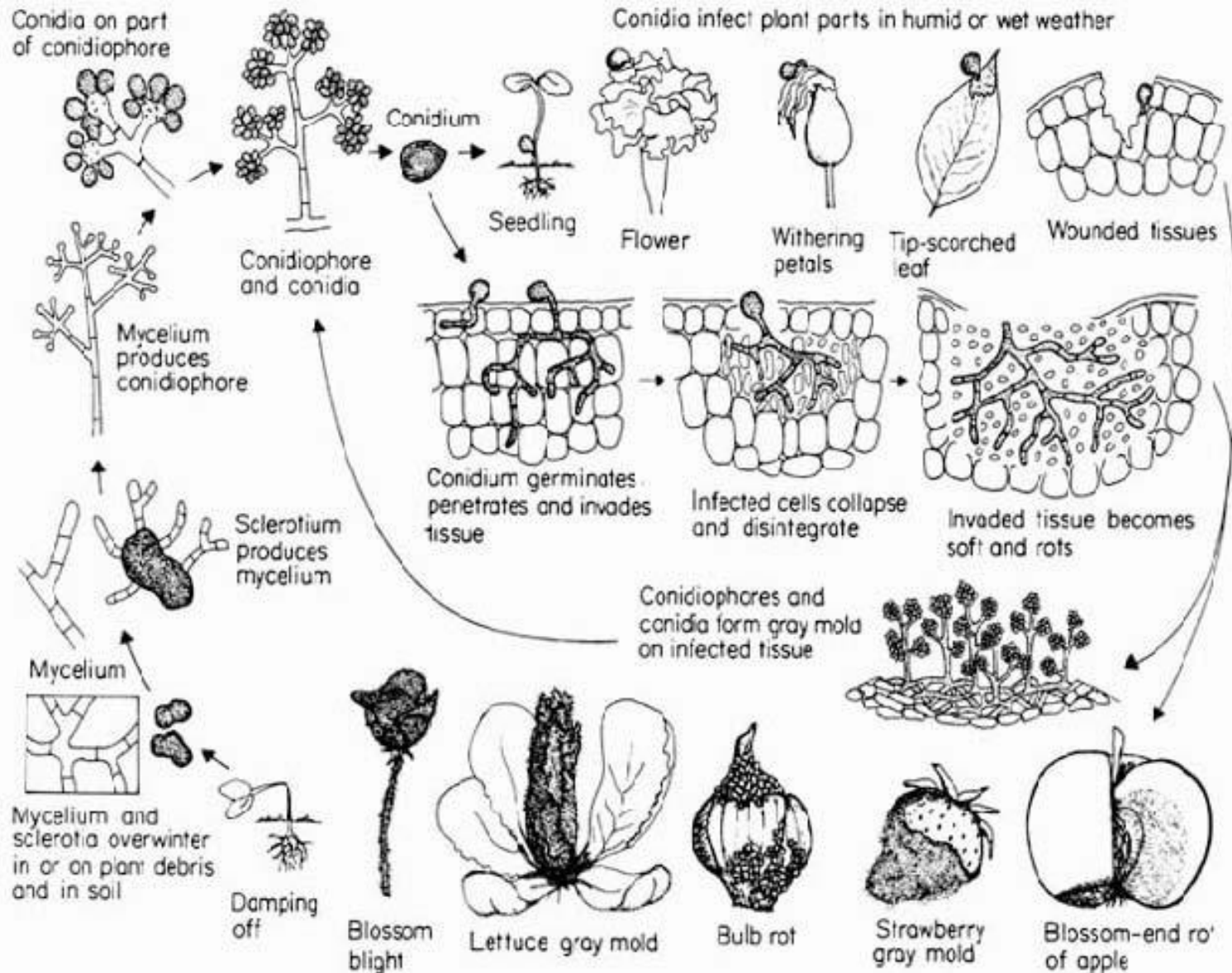
**Swinburne (1995) proposed the term “period
of quiescence” to differentiate the quiescent
parasitic relationship from the latent period.**

Air distribution and direct infection



Botrytis quiescent (latent infections)

Botrytis life cycle



Botrytis



Quiescent infections of Botrytis in grapes





Direct penetration and colonization:

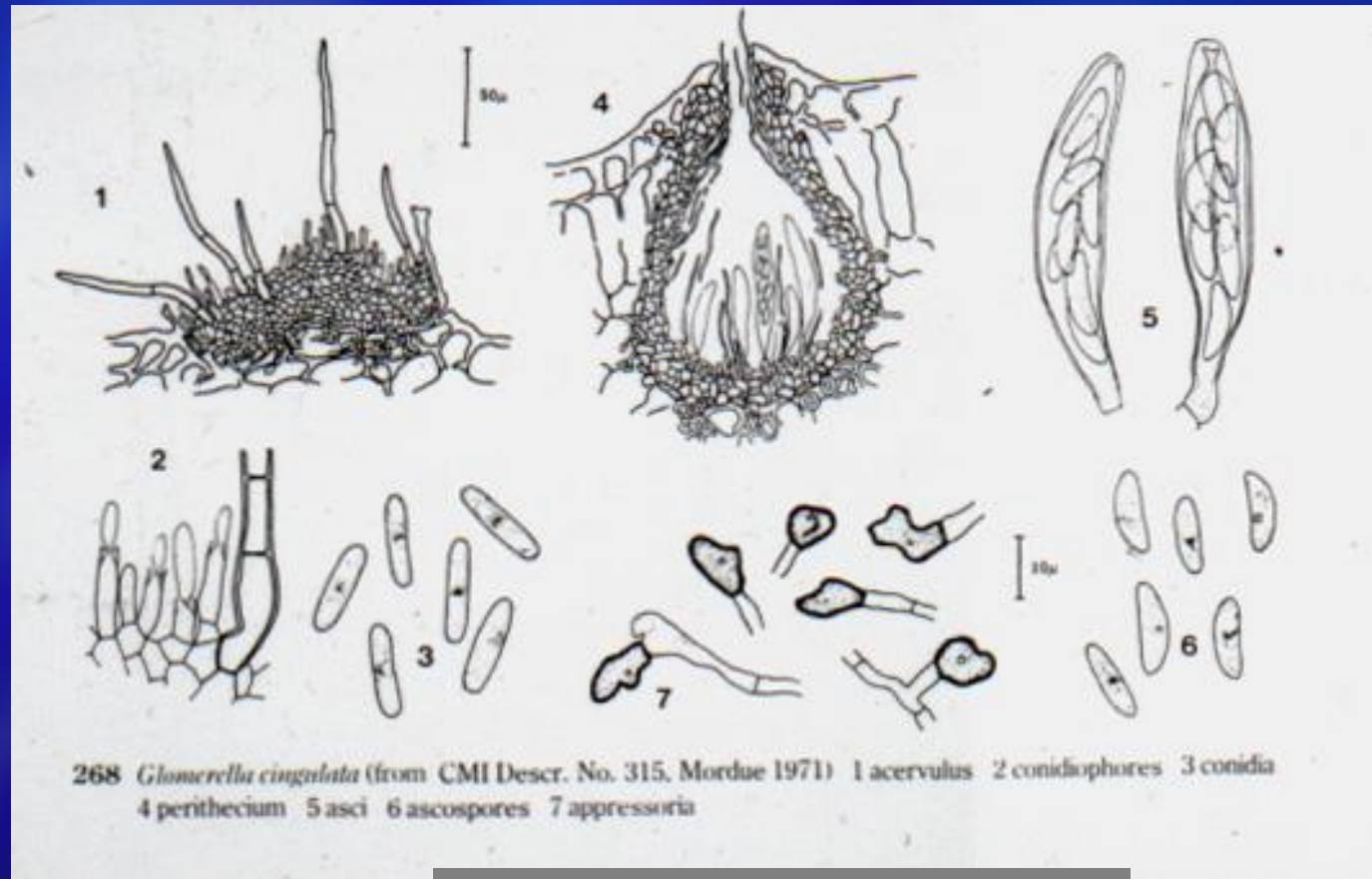
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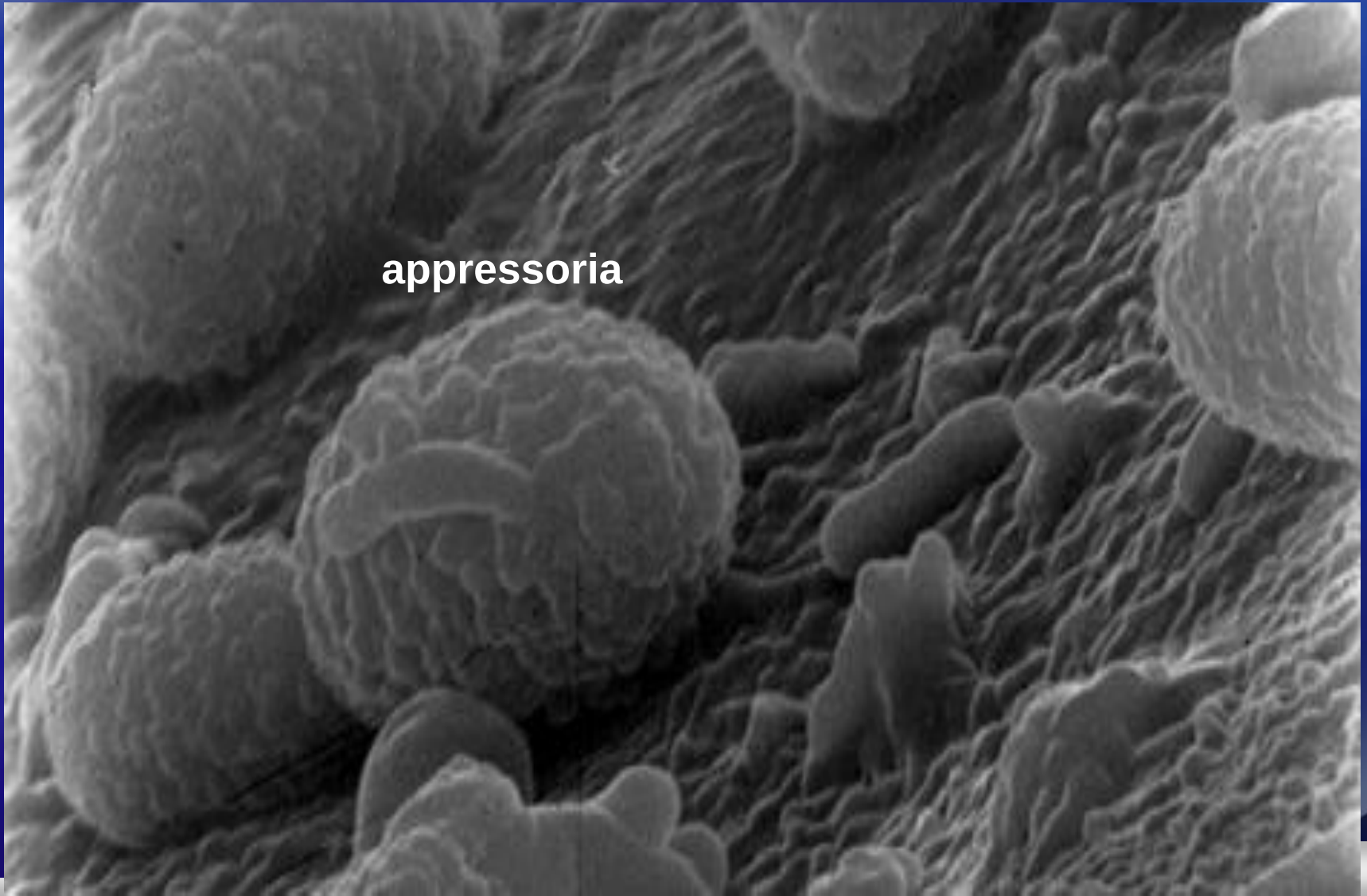
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Colletotrichum the pathogen the cause Anthracnose

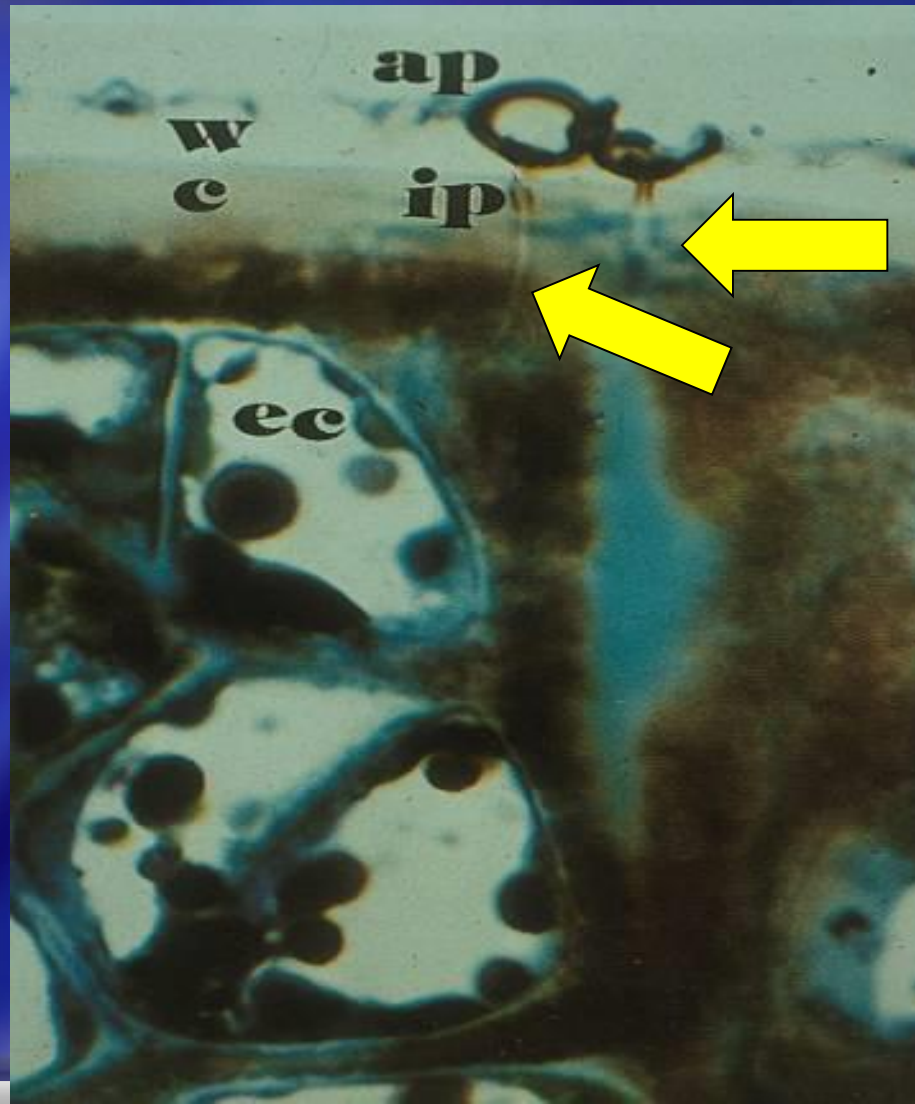


Direct penetration + no
symptoms

Colletotrichum gloeosporioides germination



Appressoria penetration and quiescence by *Colletotrichum*

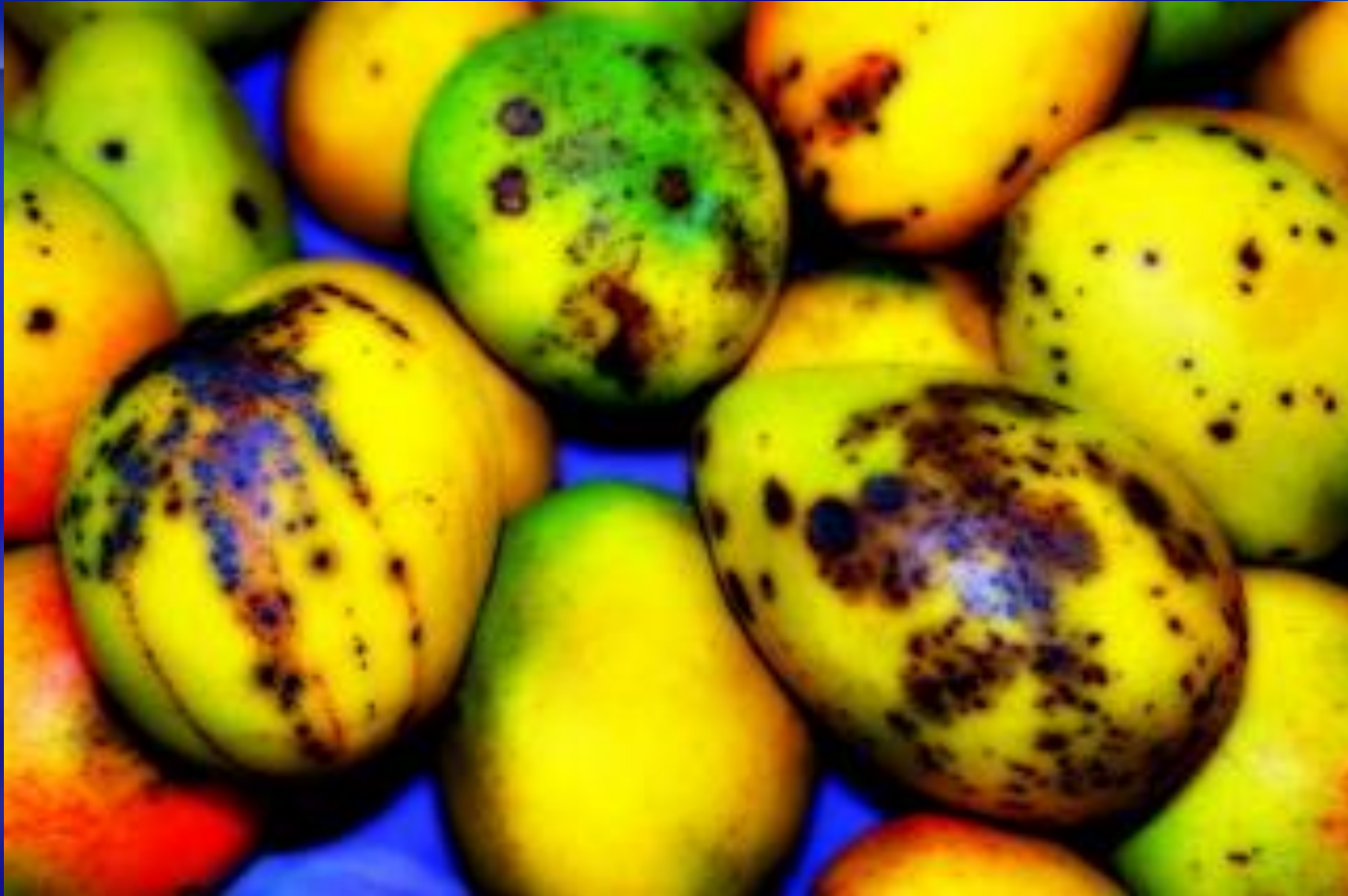


Humidity run off and quiescent infections



Colletotrichum (Anthracnose) direct penetration and quiescent infections in Mango fruits

Humidity run off and active infections



**Colletotrichum (Anthracnose)
in Mango fruits**



Colletotrichum in Mango fruits

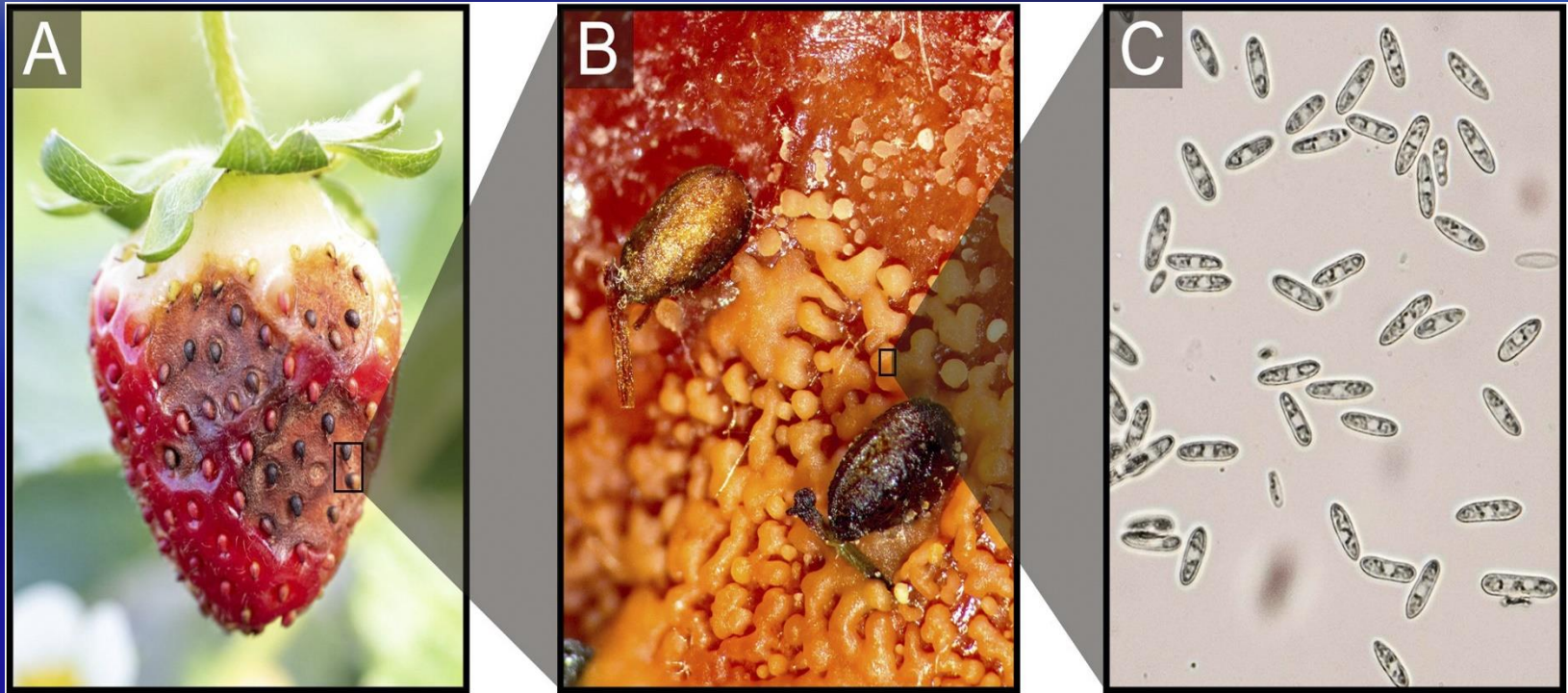
Symptoms of diseases caused by *Colletotrichum* spp. on representative temperate fruits: peach, apple, grape, and strawberry.



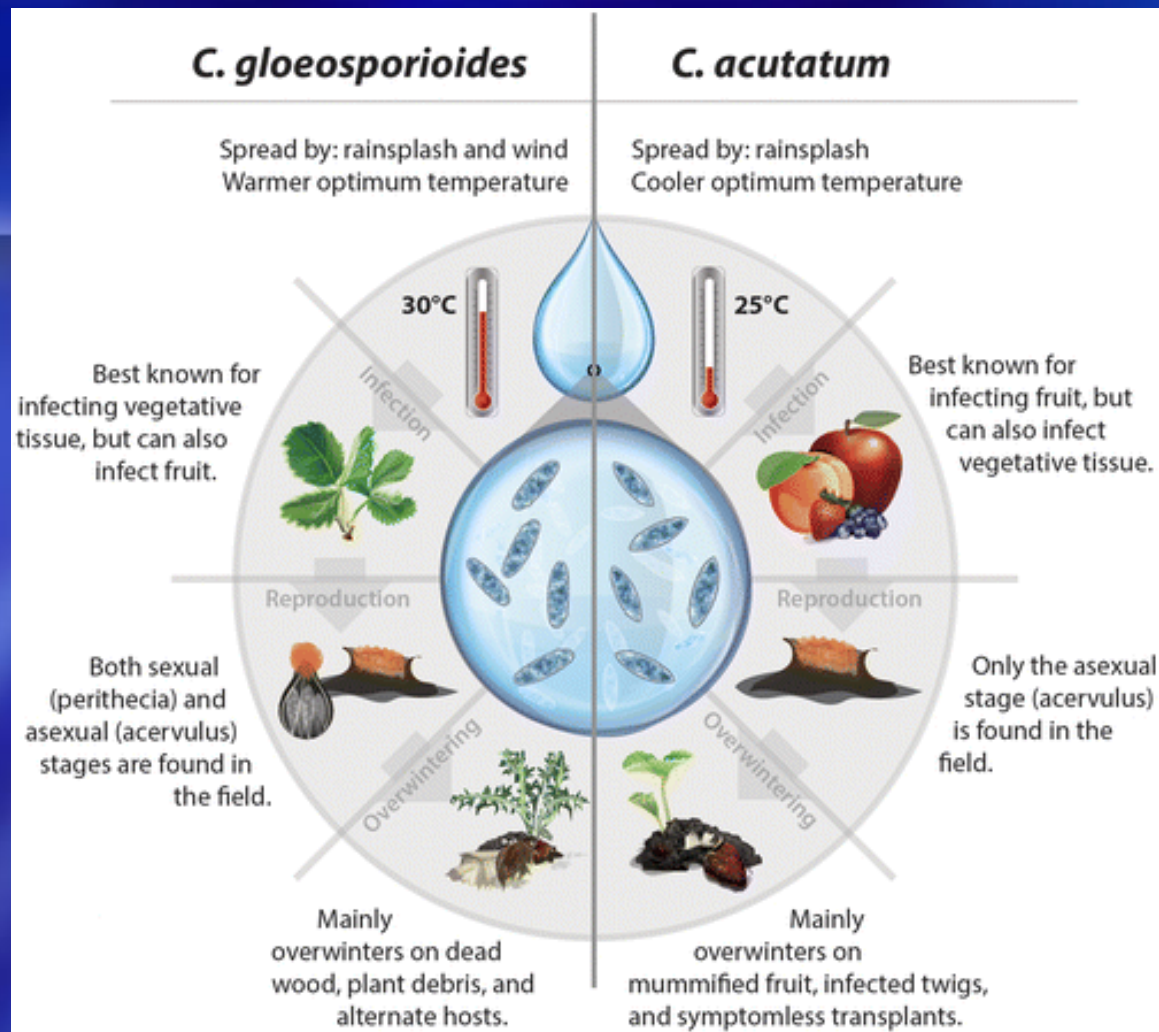
Published in: Madeline Dowling; Natalia Peres; Sara Villani; Guido Schnabel; *Plant Disease* 2020, 104, 2301-2316.

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Symptoms and signs of *Colletotrichum* spp. causing anthracnose fruit rot on strawberry: **A**, ripening strawberry infected by *C. acutatum*; **B**, microscopic image of *C. acutatum* sticky spore masses on anthracnose lesion; **C**, microscopic image of *C. acutatum* spores.



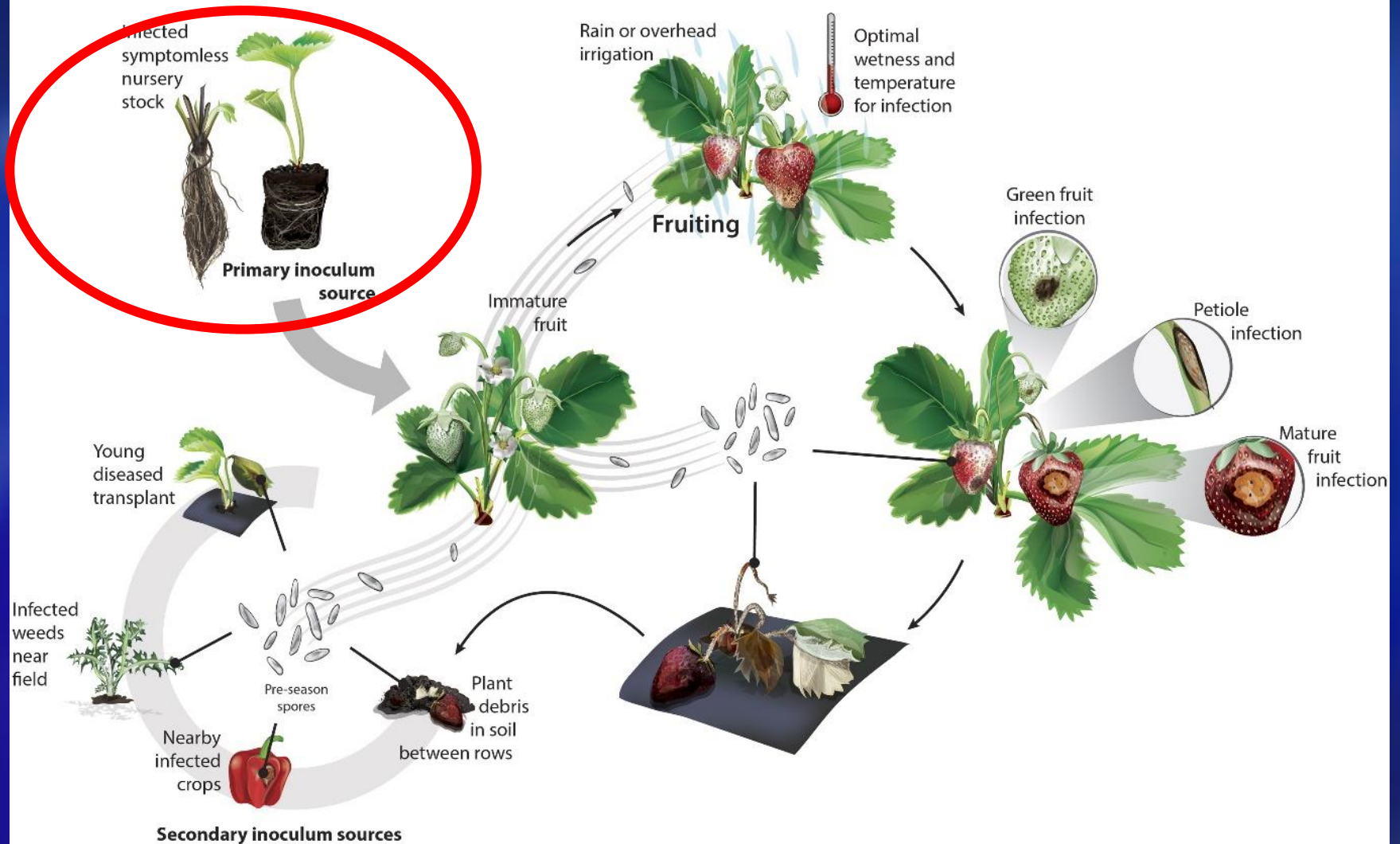
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A comparison of *Colletotrichum acutatum* and *C. gloeosporioides* on temperate fruit crops.

Anthracnose on Strawberry

(*Colletotrichum acutatum*)



• Representative *Colletotrichum* disease cycle: disease cycle of anthracnose on strawberry caused by *C. acutatum*.

COMMON NAME

Diseases caused by *Colletotrichum* spp. can have different common names on different hosts.



BITTER ROT



ANTHRACNOSE



RIPE ROT

GENUS

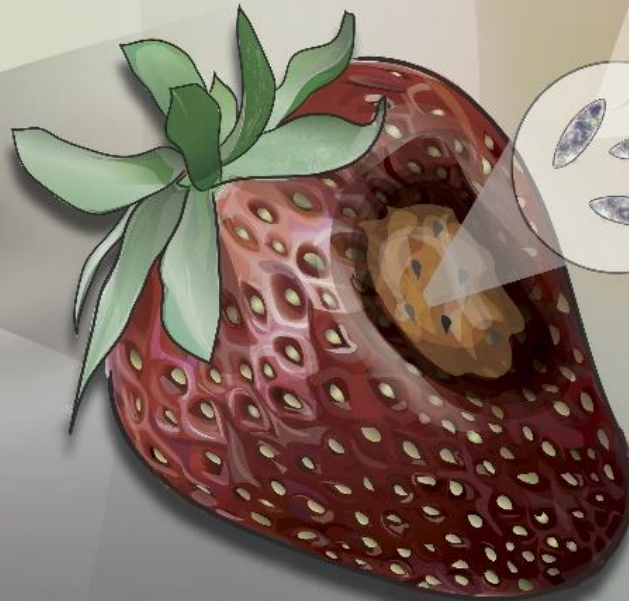
Diseases caused by fungi in the genus *Colletotrichum* can be recognized by orange spore masses and sunken lesions on symptomatic tissue.

COMPLEX

Multiple *Colletotrichum* complexes can cause disease on the same host. Identification to the complex level requires sequencing at least one gene.

SPECIES

Colletotrichum complexes contain multiple species. Identification to the species level often requires sequencing at least three genes.



Future genetic testing may reveal even more diversity within currently defined *Colletotrichum* spp.

Illustrated by Madeline Dowling
www.phytographics.com

Levels of complexity in distinguishing *Colletotrichum* spp.

LIFECYCLE OF ANTHRACNOSE OF AVOCADOS

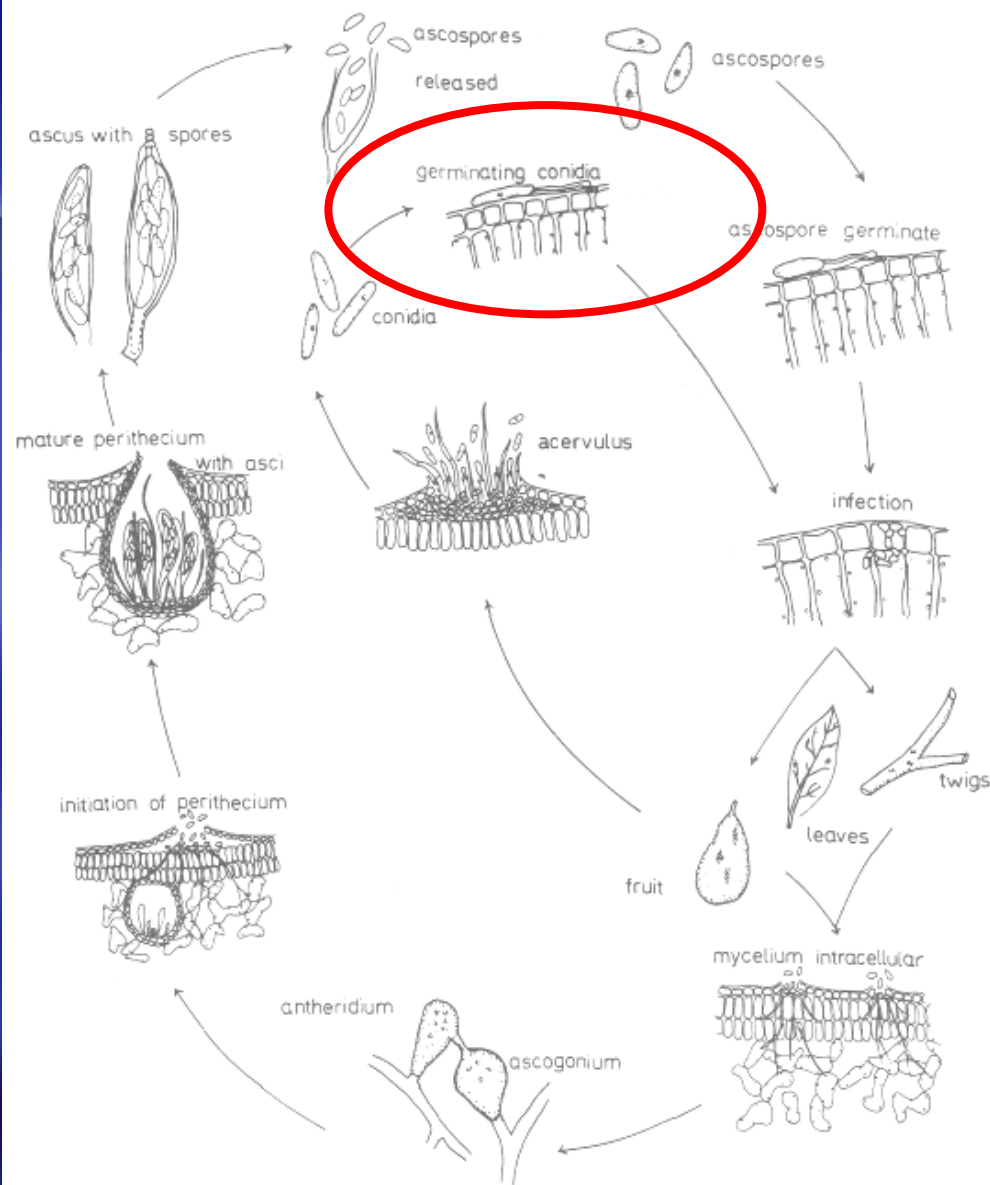
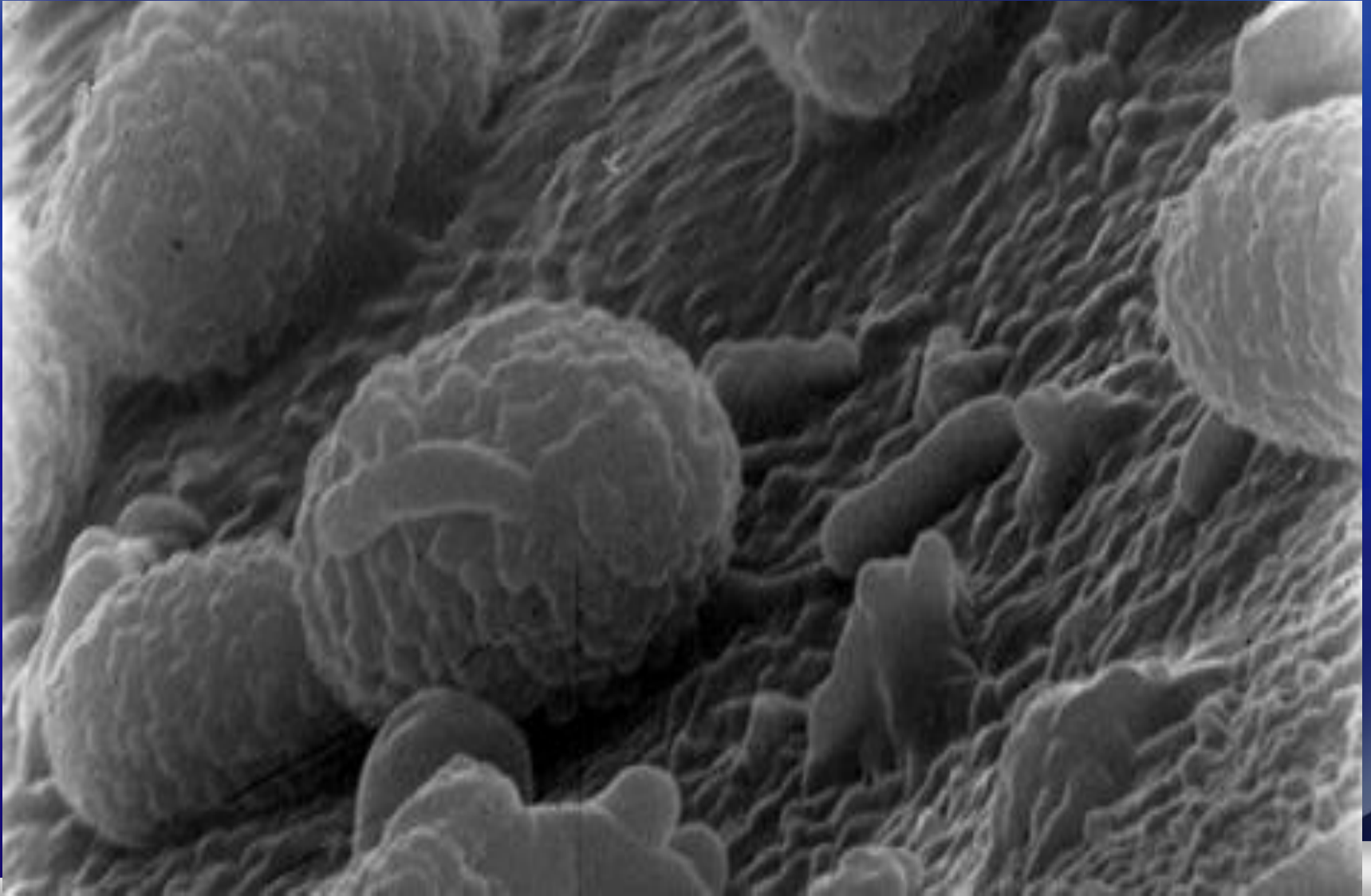
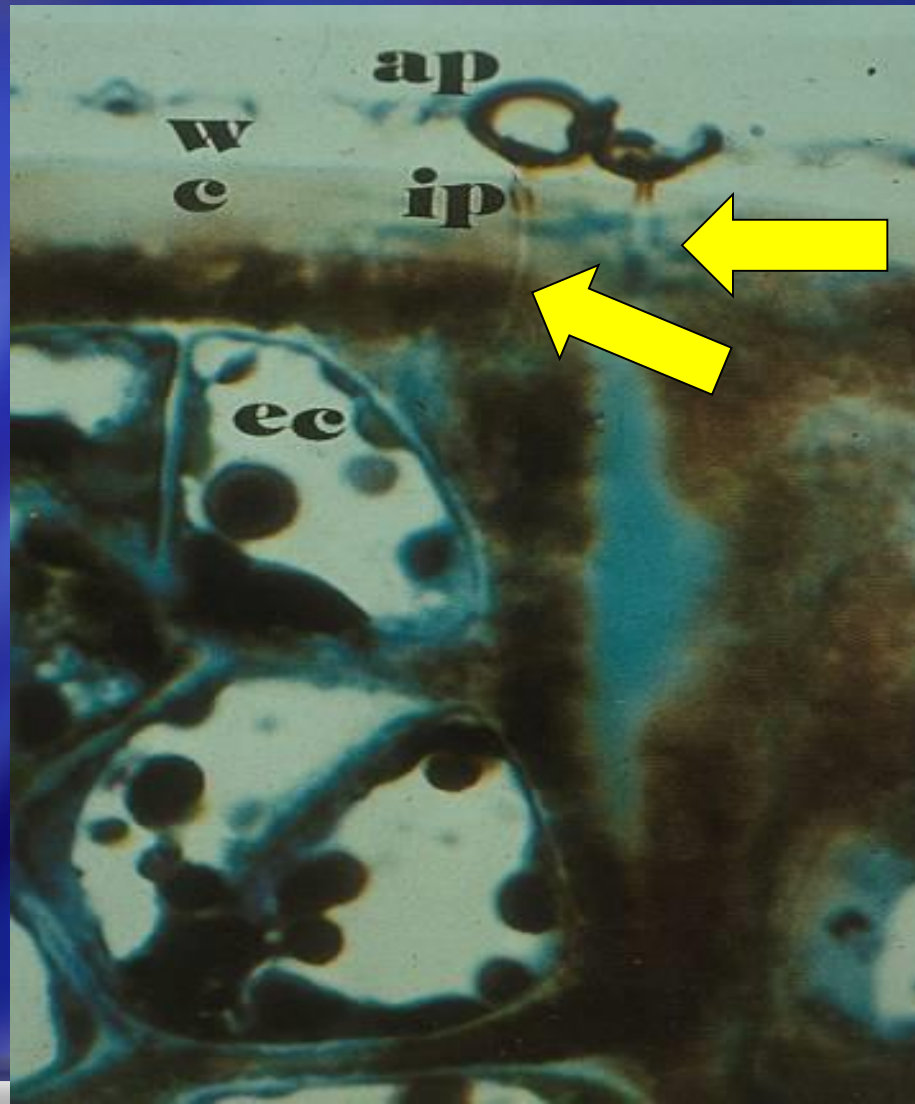


FIG. 3: Schematic presentation of the life cycle of *Colletotrichum gloeosporioides* on avocados

***Colletotrichum gloeosporioides* germination**



Appressoria penetration and quiescence by *Colletotrichum*



Colletotrichum (Anthracnose) in ripe avocado



Colletotrichum (Anthracnose) in ripe avocado



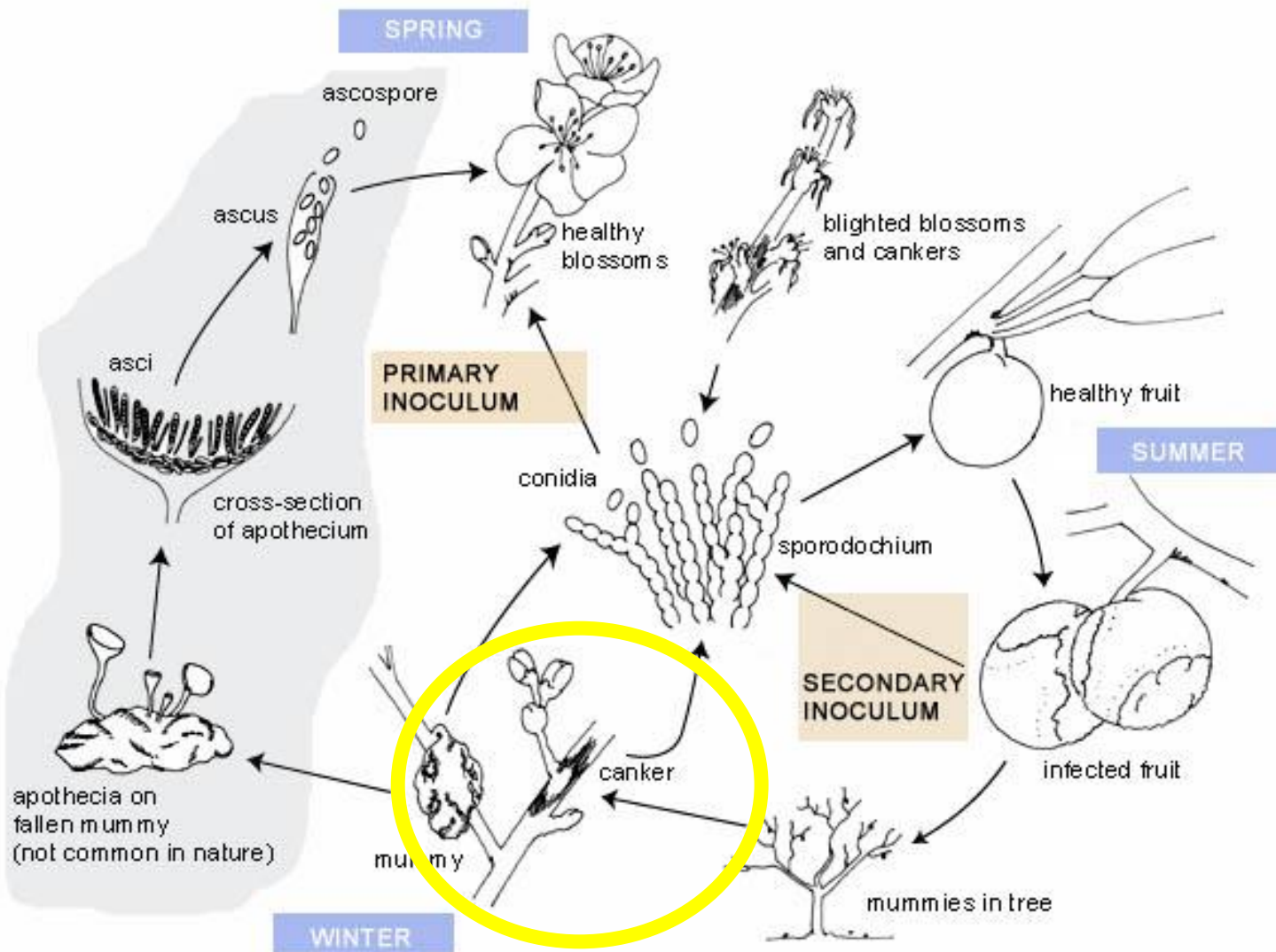
Anthracnose in Chilli



Anthracnose in banana



Monilia fructicola



Drawing courtesy Vickie Brewster

Wound penetration as a result of direct penetration and wound caused by wind insects insects



**Mummy from
last year fruit**

Monilinia sp sporulating on mummified fruit left from last year infects new cherry flowers resulting in blossom blight.

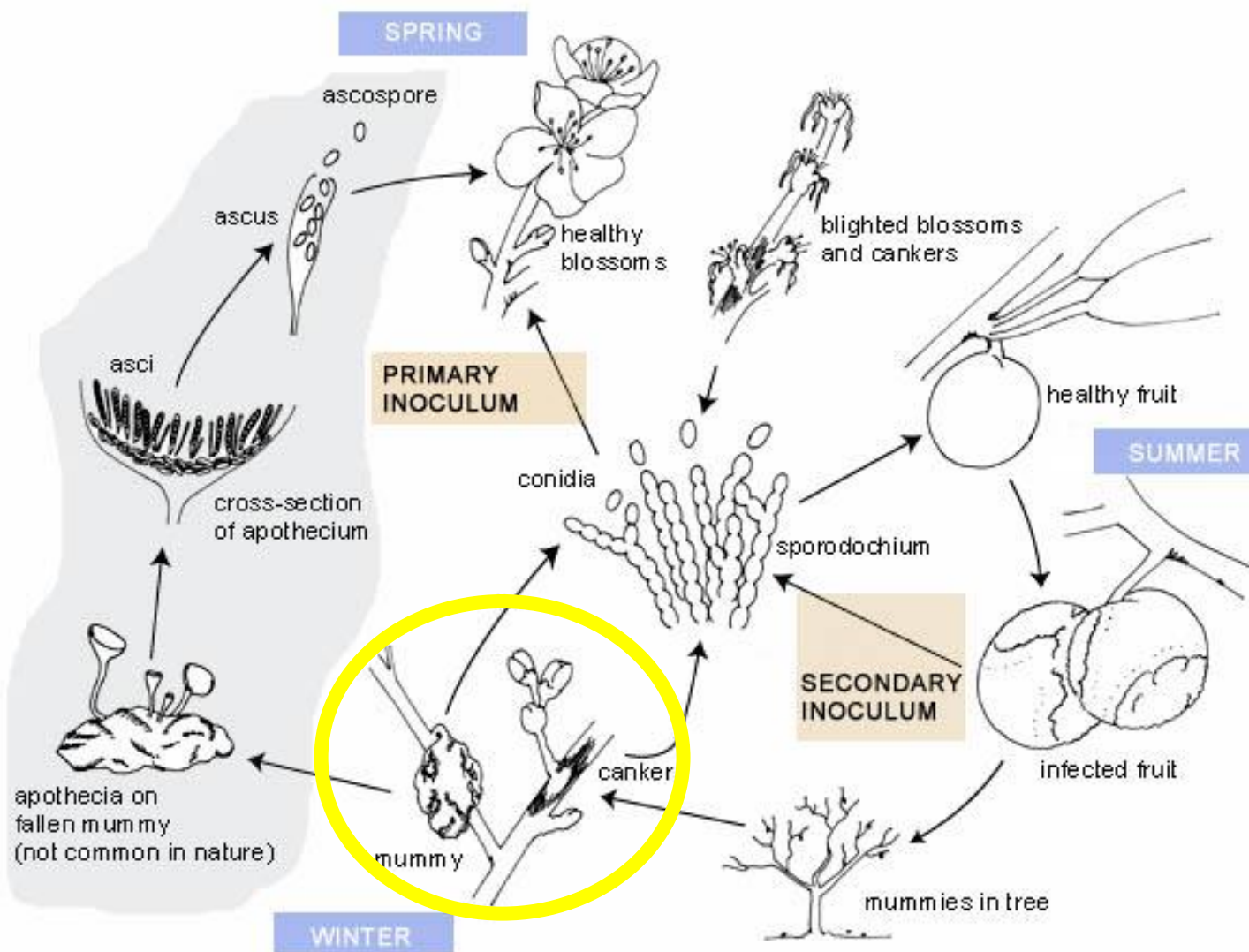


Monilinia fructicola



Direct penetration + no
symptoms

Monilia fructicola



Drawing courtesy Vickie Brewster

Direct penetration and colonization:

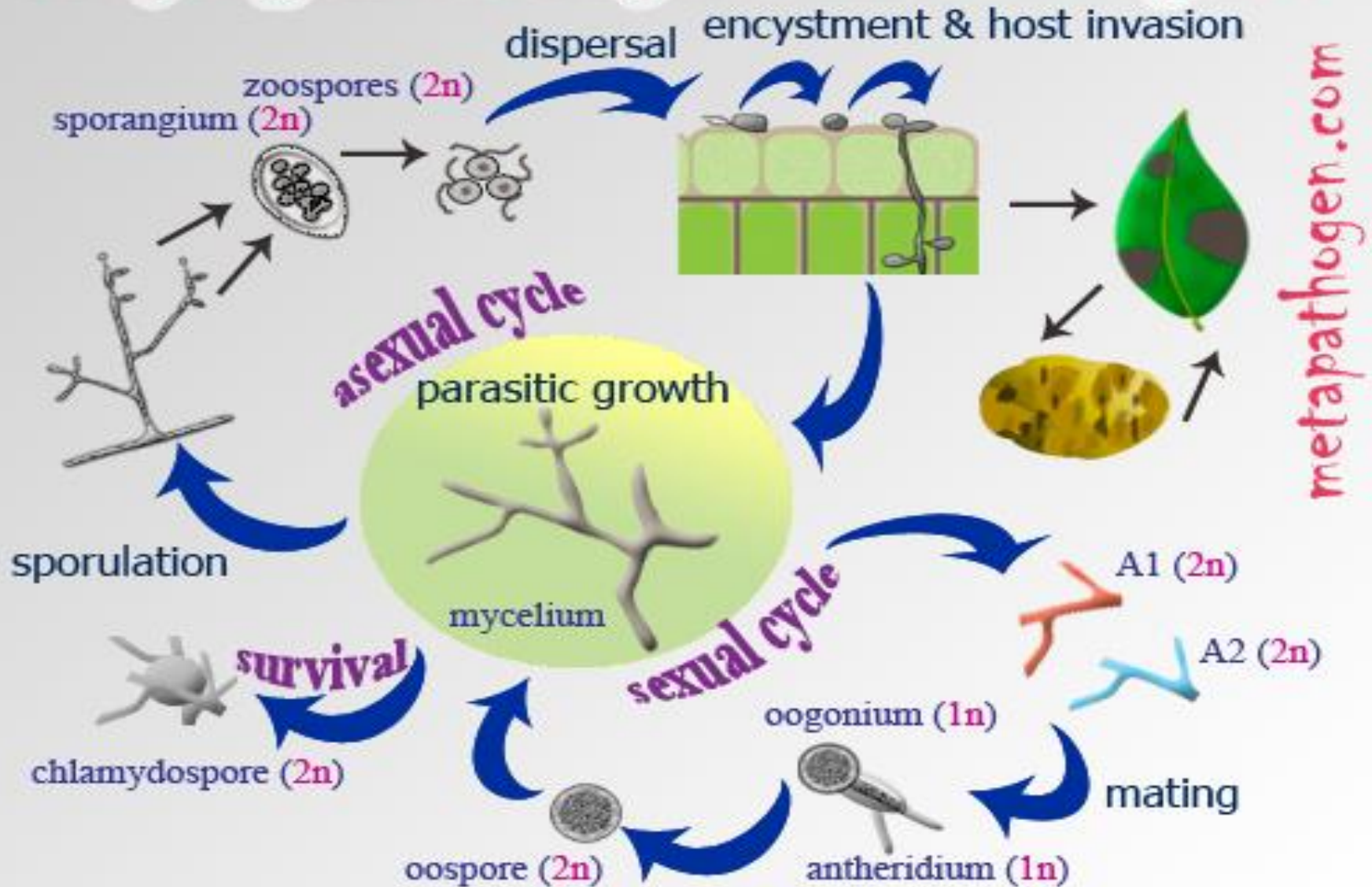
Showing no Symptoms- Quiescent infections by producing appressoria for penetration

- *Botrytis cinerea*
- *Colletotrichum gloeosporioides*
- *Monilinia fructicola*

Showing small initial symptoms:

- *Phytophthora infestans* in potato
- *Phytophthora citrophthora* in citrus

Phytophthora infestans life cycle



Phytophthora infestans



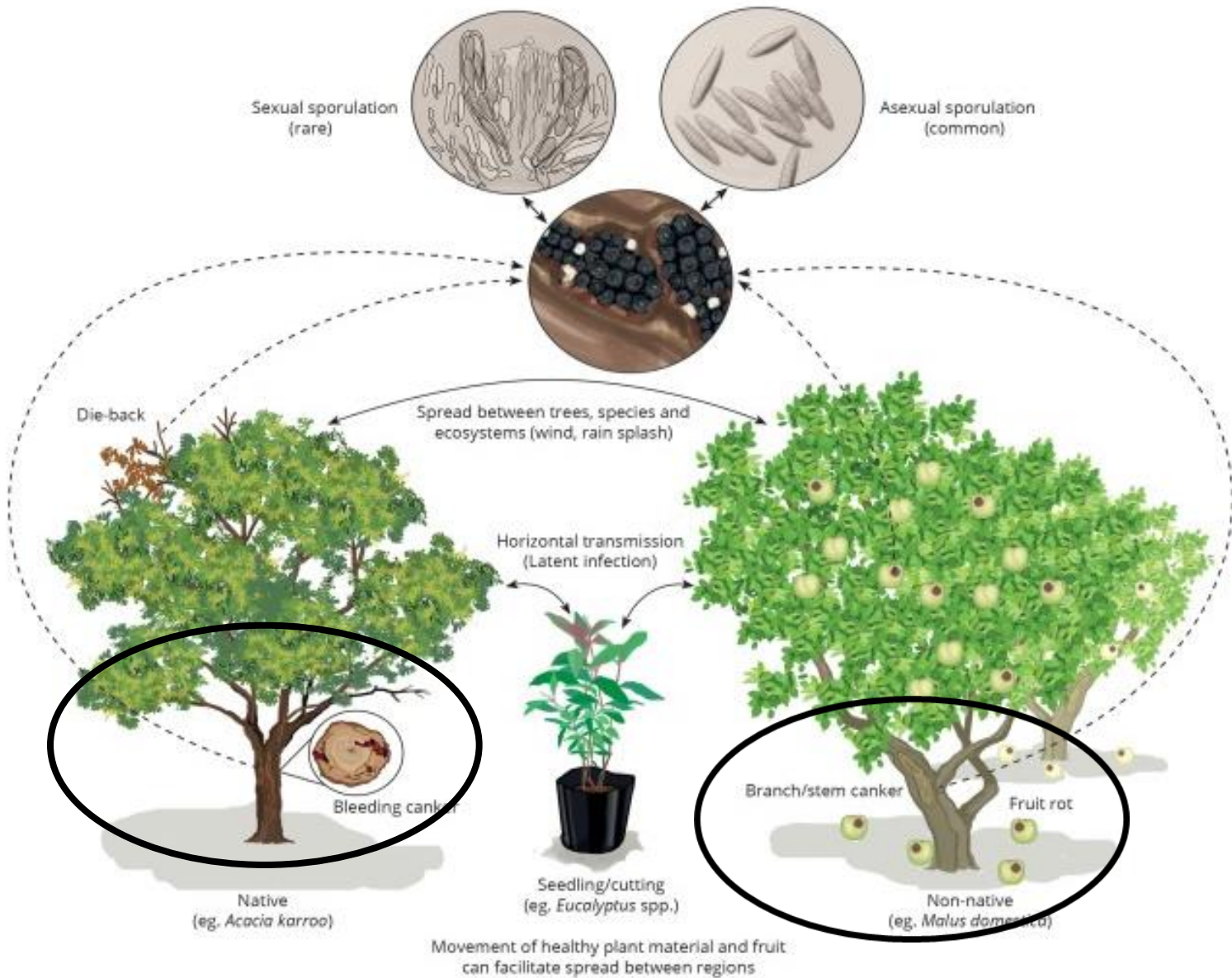
Direct penetration + small
symptoms

Phytophthora infestans



Infected tubers

An example of the changes in disease incidence: a field disease that become and postharvest disease: *Botryosphaeria dothidea*.



The illustration represents infection of the fungus on three hosts, as observed in South Africa, including native (*Acacia karroo*) and non-native (*Eucalyptus* and *Malus domestica*) hosts.

Typical disease symptoms include dieback, cankers and fruit rot. Fruiting structures containing sexual and/or asexual spores are often found associated with the disease symptoms and these are dispersed through wind or rain splash.

Movement of latent or endophytically infected plant material to new regions poses a threat to quarantine systems. Given the broad host range and geographical distribution of *B. dothidea*, all three hosts could be replaced by various others, as could the geographical setting.

Penetration through wounds

- **Small wounds**
- **Lenticels**
- **Stem wounds**
- **Wounds caused by hands**
- **Wounds caused by pruning**
- **Wounds caused by nails**
- **Wounds caused by birds**
- **Wounds caused by insects**
- **Wound caused by storage conditions**

Penetrations by wounds

- *Penicillium sp.*
- *Colletotrichum*
- *Lasiodiplodia theobromae*
- *Rhizopus stolonifer*
- *Alternaria alternata*
- *Botrytis cinerea*

Classification of wound penetration according to period of penetration of the fungus

- 1. Before harvest**
- 2. During harvest**
- 3. After harvest**

Classification of way of disease penetration according to period of penetration of the fungus

Before harvest

Phytophthora infestans

Phytophthora citrophthora

Botrytis cinerea

Alternaria alternata

During harvest

Penicillium italicum

Penicillium expansum

Lasiodiplodia theobromae

Colletotrichum

After harvest

Monilinia

Rhizopus stolonifer

Botrytis cinerea

Classification of way of disease penetration according to period of penetration of the fungus

Before harvest

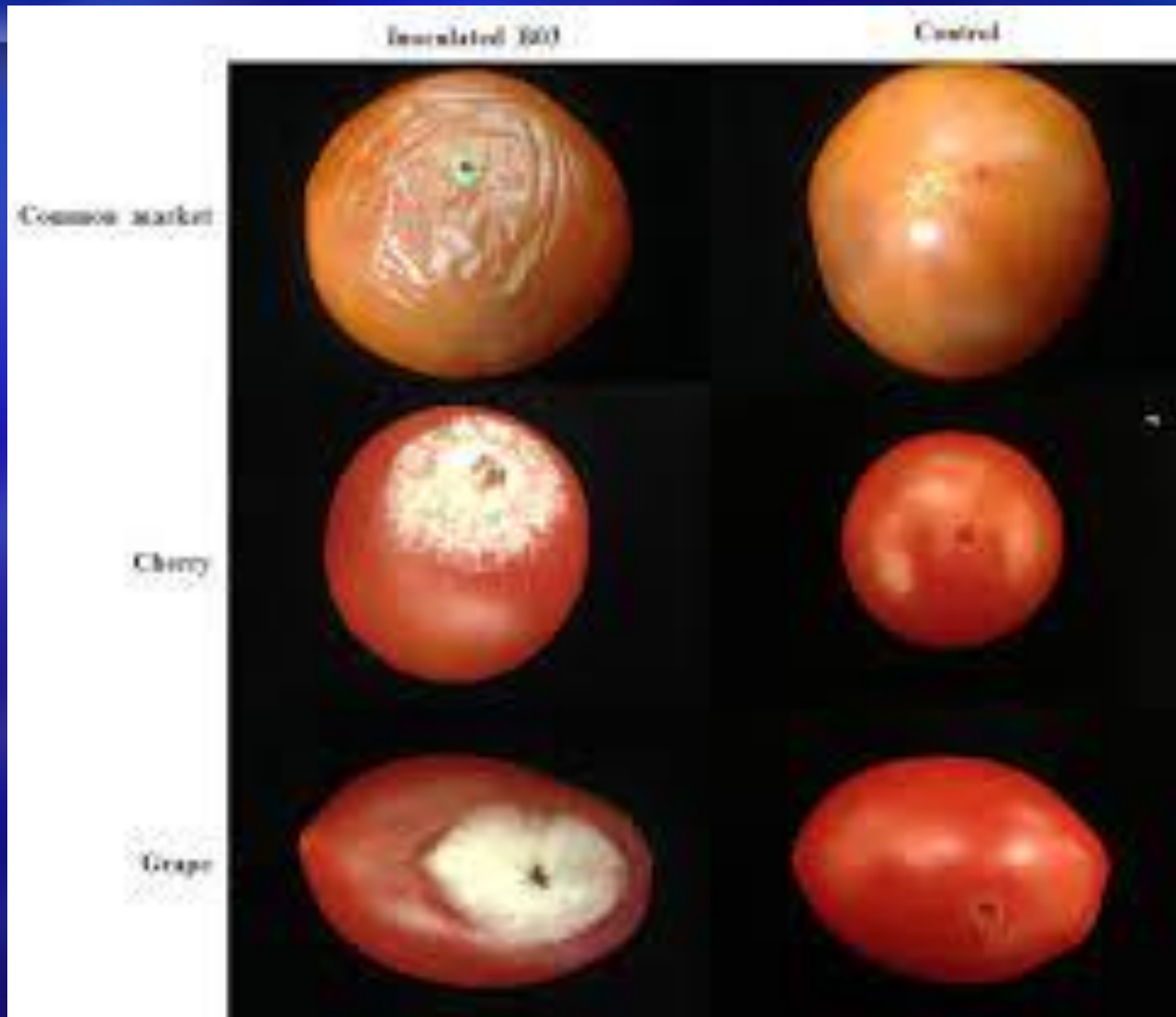
Phytophthora infestans

Phytophthora citrophthora

Botrytis cinerea

Alternaria alternata

Botrytis



Cracks as a result of growth



Classification of way of disease penetration according to period of penetration of the fungus

Before harvest

Phytophthora infestans

Phytophthora citrophthora

Botrytis cinerea

Alternaria alternata



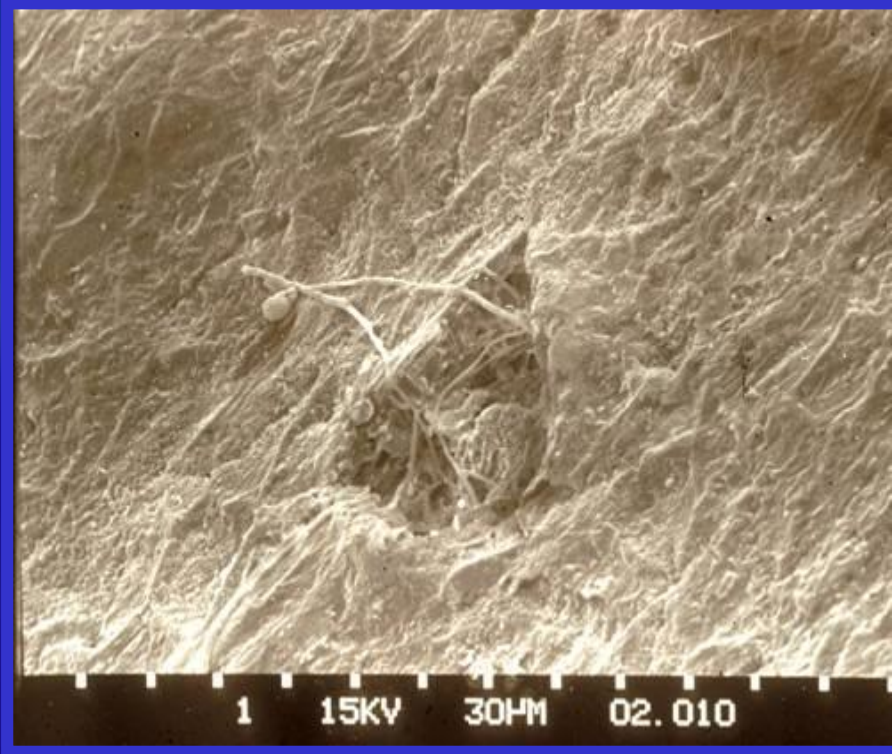
Alternaria alternata

**Alternaria rot of tomatoes, peppers and eggplants
caused by *Alternaria alternata* wound penetration**



Mode of infection of Alternaria in mango fruit: natural openings

a. Lenticel penetration

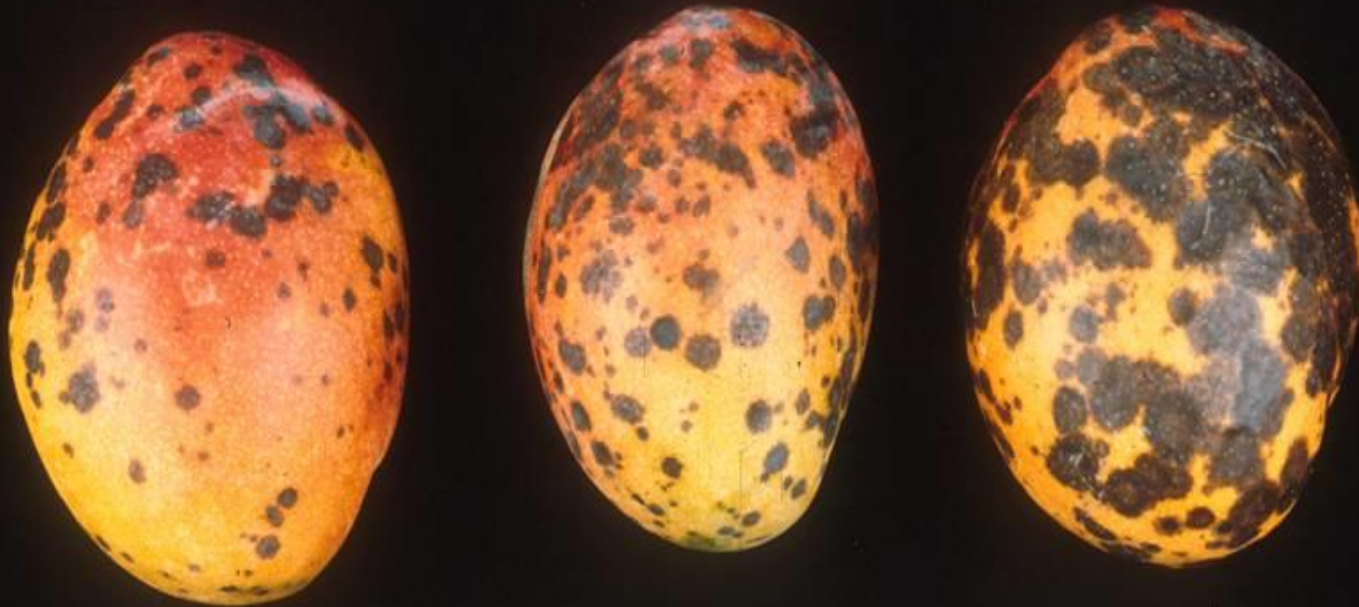


b. Initial symptoms



Alternaria penetration through
lenticels in Mango fruits

***Alternaria alternata* symptoms**



MATURE HADEN

2W

AT 5

Classification of way of disease penetration according to period of penetration of the fungus

Before harvest

Phytophthora infestans

Phytophthora citrophthora

Botrytis cinerea

Alternaria alternata

During harvest

Penicillium italicum

Penicillium expansum

Lasiodiplodia theobromae

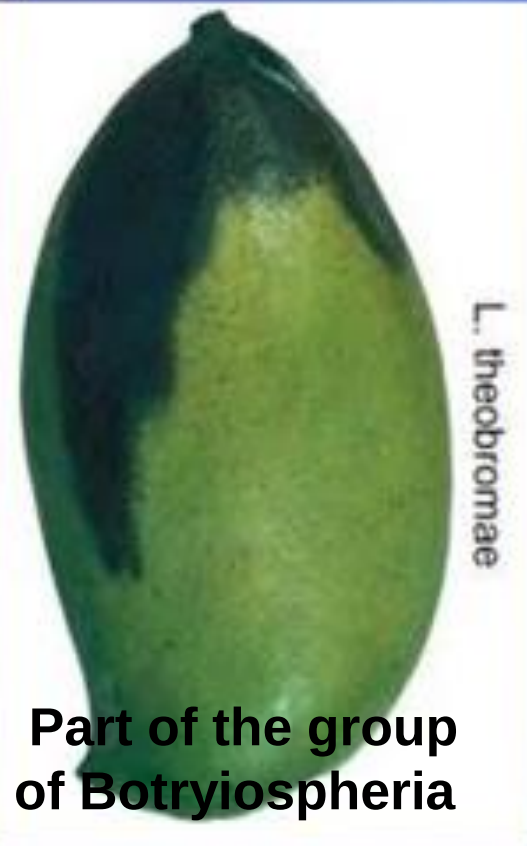
Colletotrichum

Lasiodiplodia theobromae stem end penetration



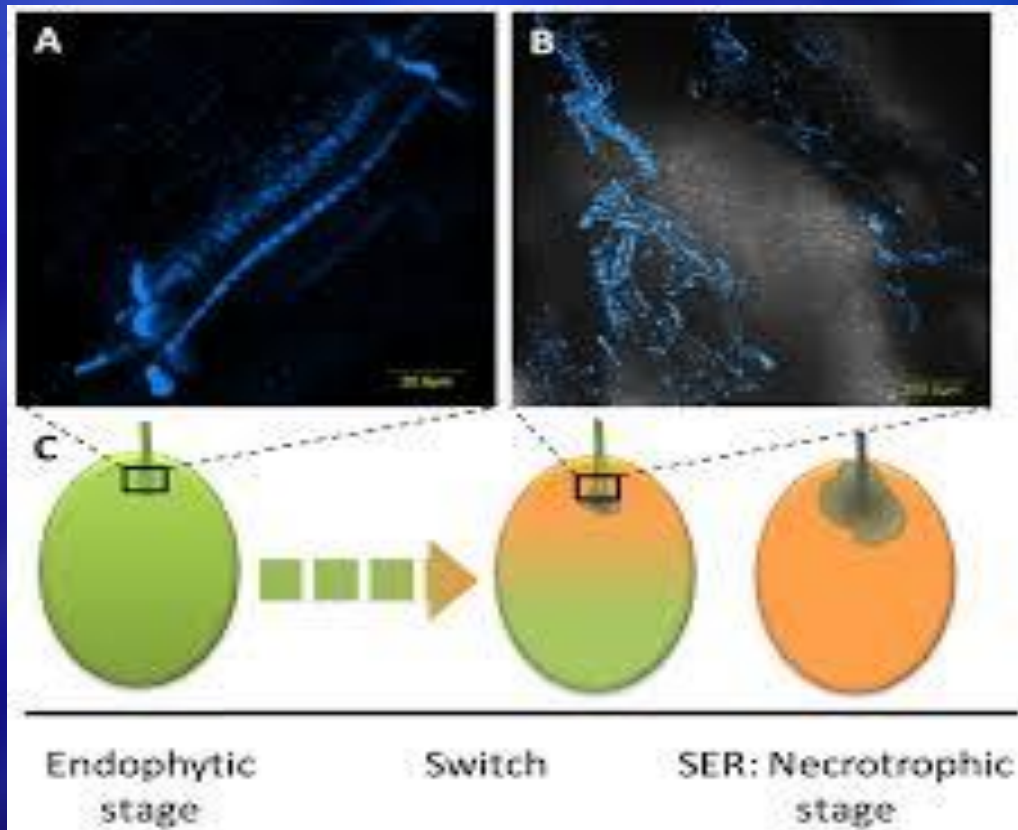
Test.

Inoc.



Part of the group
of Botryosphaeria

Stem end infected by penetrating pathogens



Alkan et al.
2015

Lasioidiplodia, penetration through the stem end



Lasioidiplodia, penetration through the stem end



Colletotrichum stem end penetration of avocado



Classification of way of disease penetration according to period of penetration of the fungus

Before harvest

Phytophthora infestans

Phytophthora citrophthora

Botrytis cinerea

Alternaria alternata

During harvest

Penicillium digitatum and *italicum*

Penicillium expansum

Lasiodiplodia theobromae

Colletotrichum gloesporioides

Green mold rot of citurs cause by *Penicillium digitatum* by wound penetration

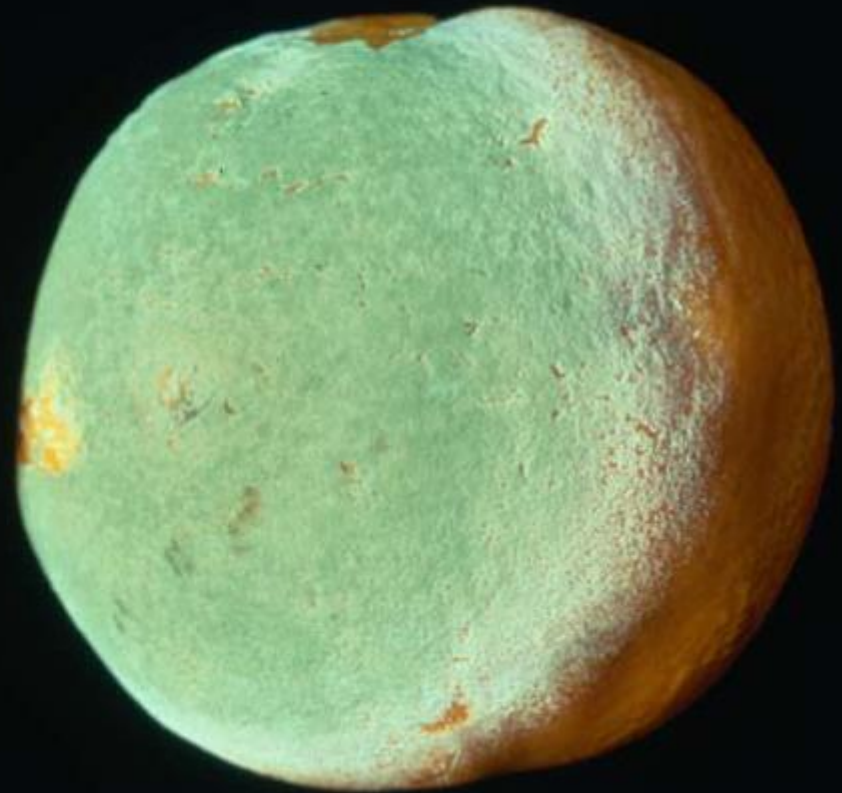


Blue mold rot of citrus cause by *Penicillium italicum*
wound penetration

Penicillium italicum



Penicillium digitatum



Penicillium expansum wound penetration



Classification of way of disease penetration according to period of penetration of the fungus

Before harvest

Phytophthora infestans

Phytophthora citrophthora

Botrytis cinerea

Alternaria alternata

During harvest

Penicillium italicum

Penicillium expansum

Lasiodiplodia theobromae

Colletotrichum

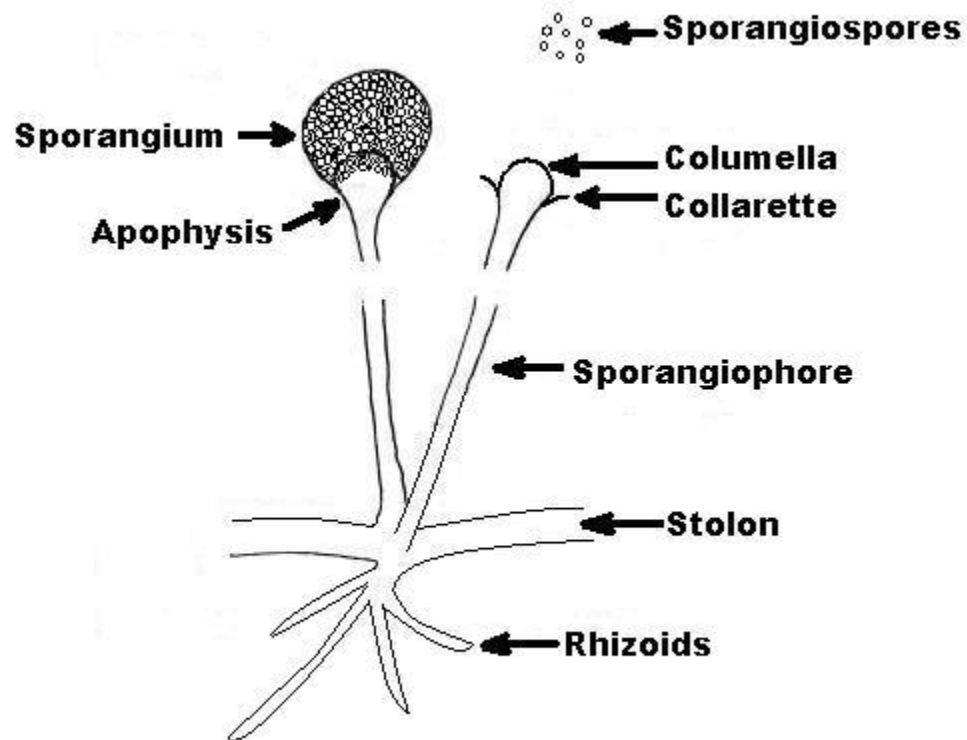
After harvest

Monilinia

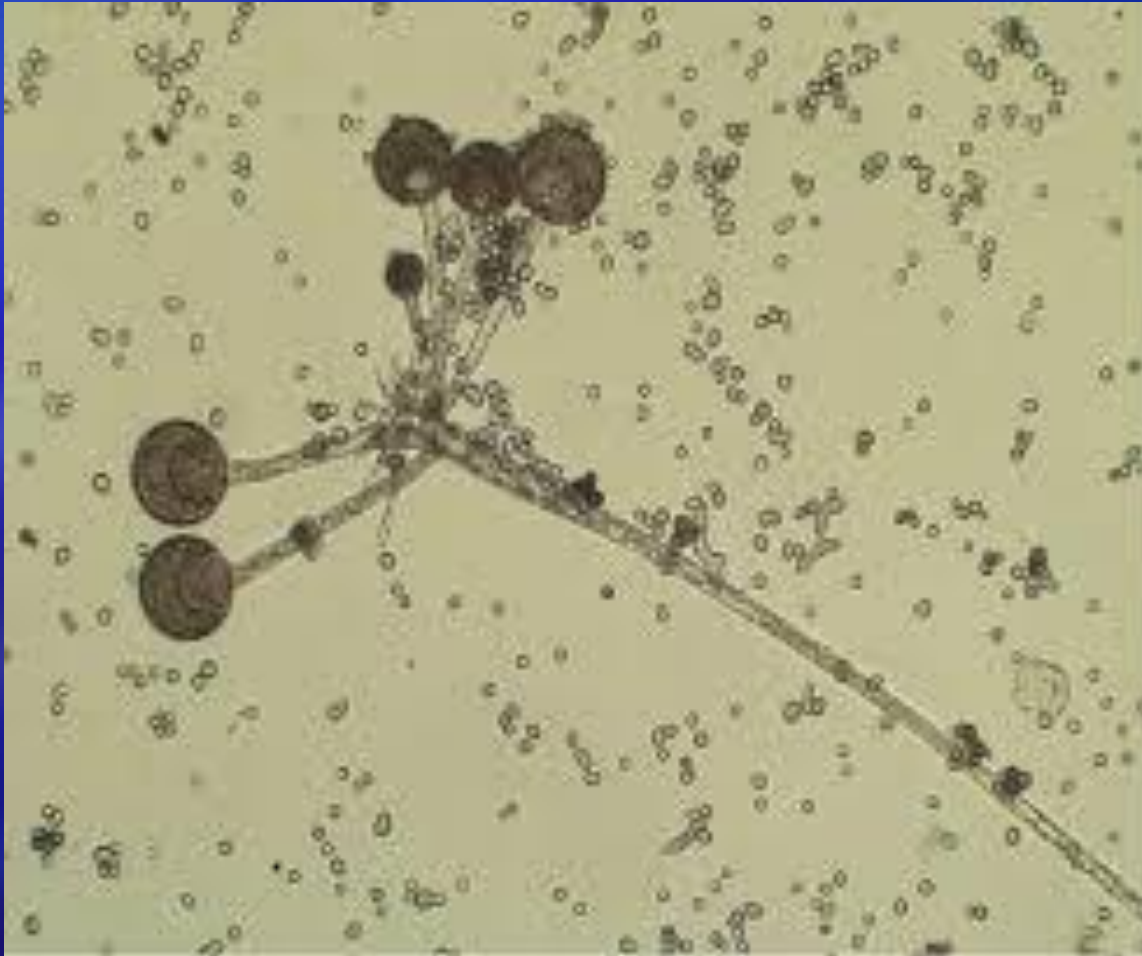
Rhizopus stolonifer

Botrytis cinerea

Rhizopus



Spores of Rhizopus in water solutions of decayed fruits



Rhizopus wound penetration in stored fruit





Summary

Penetration of postharvest pathogens

Spread of spores

- Air movement
 - Water, water splash, humidity run off
 - Mechanical wounds
 - Direct contact
-
- **Direct penetration**
 - **Penetration through wounds**

Classification of way of disease penetration according to period of penetration of the fungus

Before harvest

Phytophthora infestans

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