

4th Lesson, Pathology

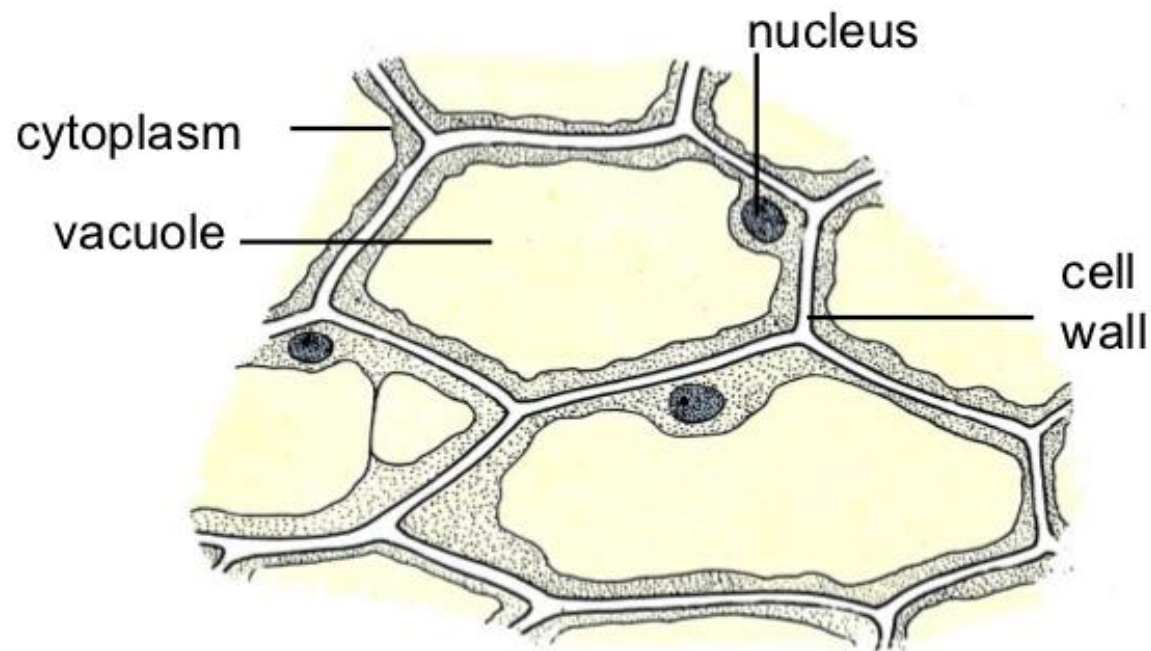
**The fungal mechanism contributing to
fungal host colonization**

**Activation of fungal
pathogenicity factors
during ripening fruits**

Fruit decay include fruit maceration

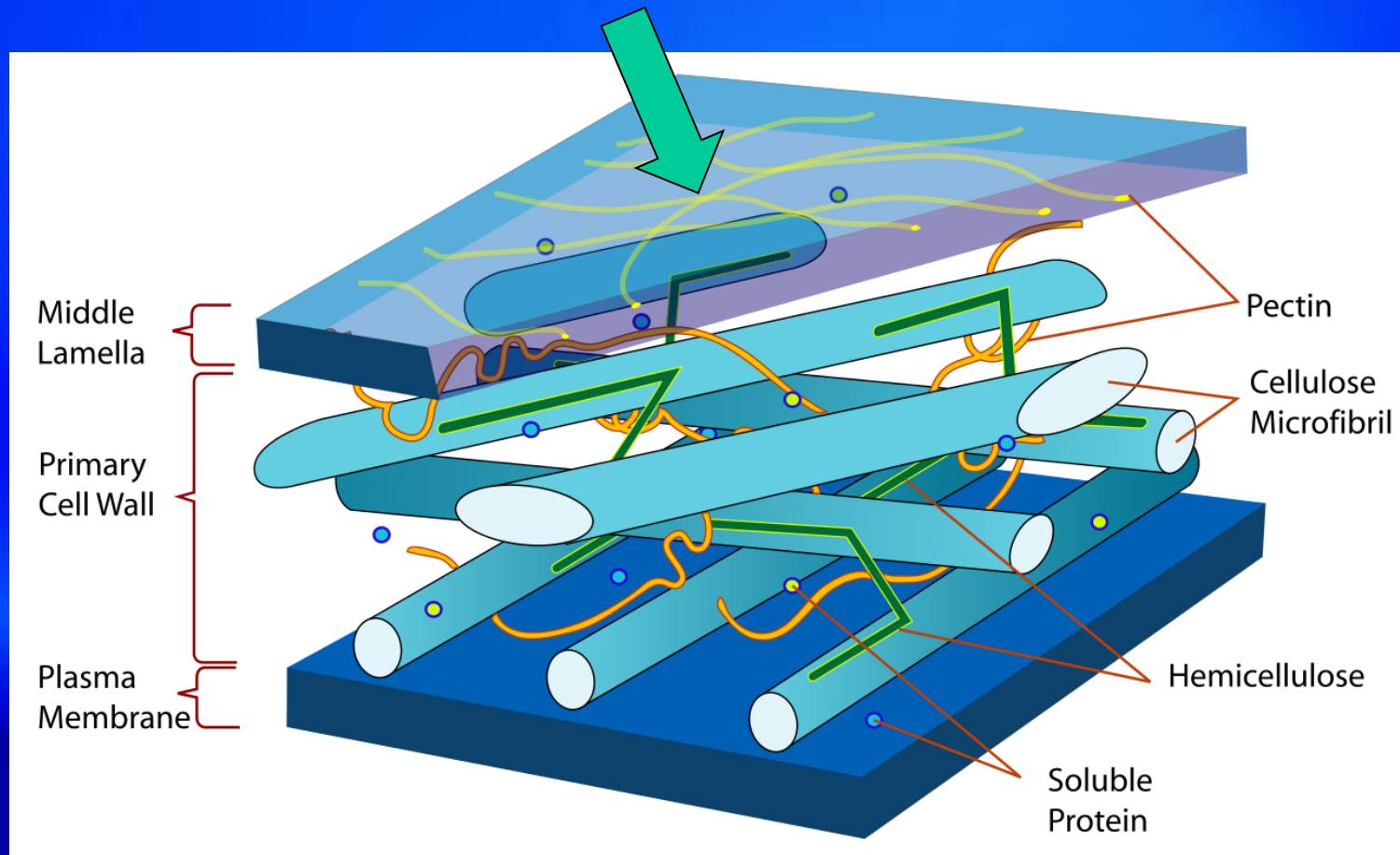


Plant cells

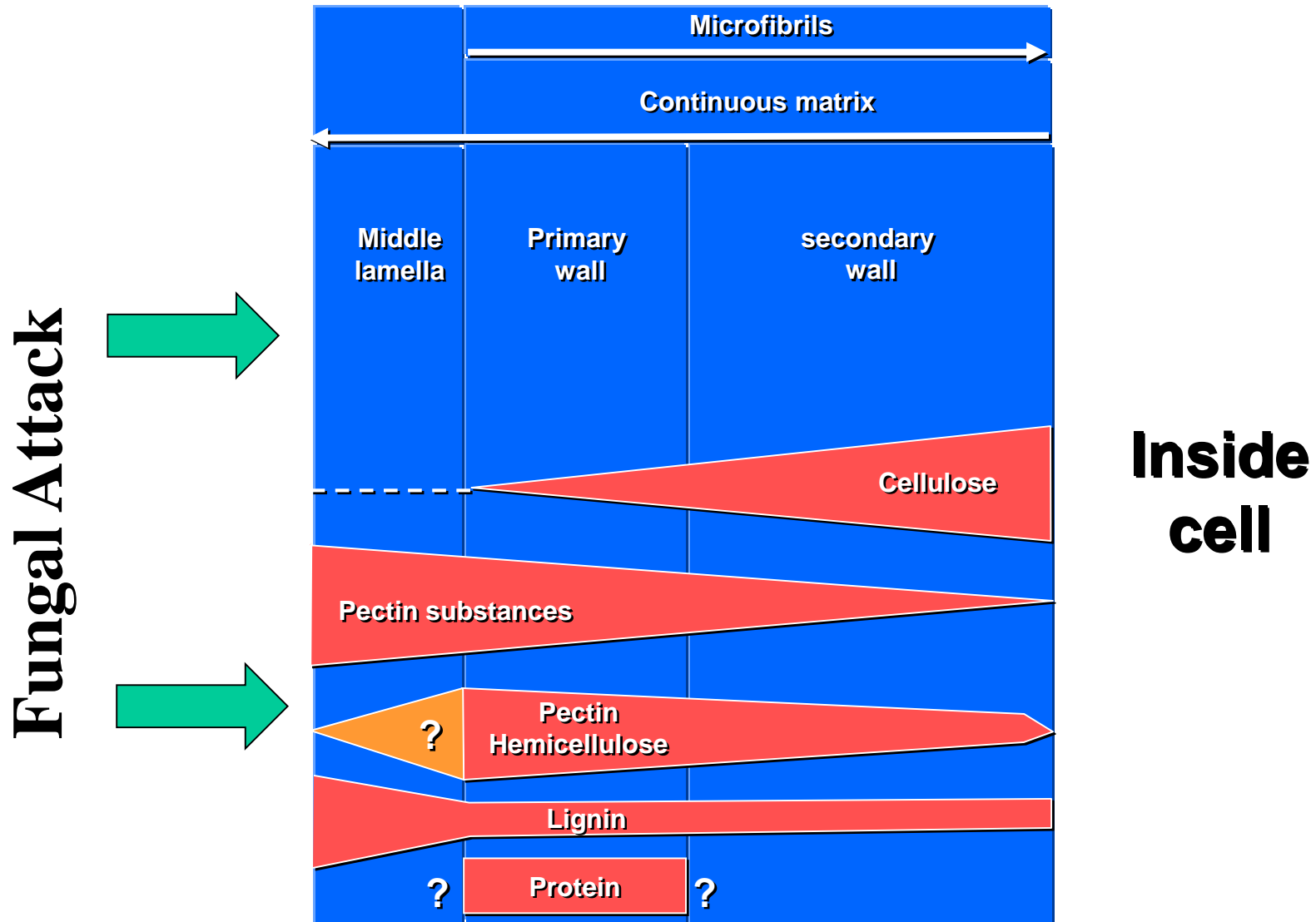


Plant cells differ from animal cells in having a cell wall outside the cell membrane, and a large, fluid-filled vacuole

Cell wall of the plant



General Cell wall structure



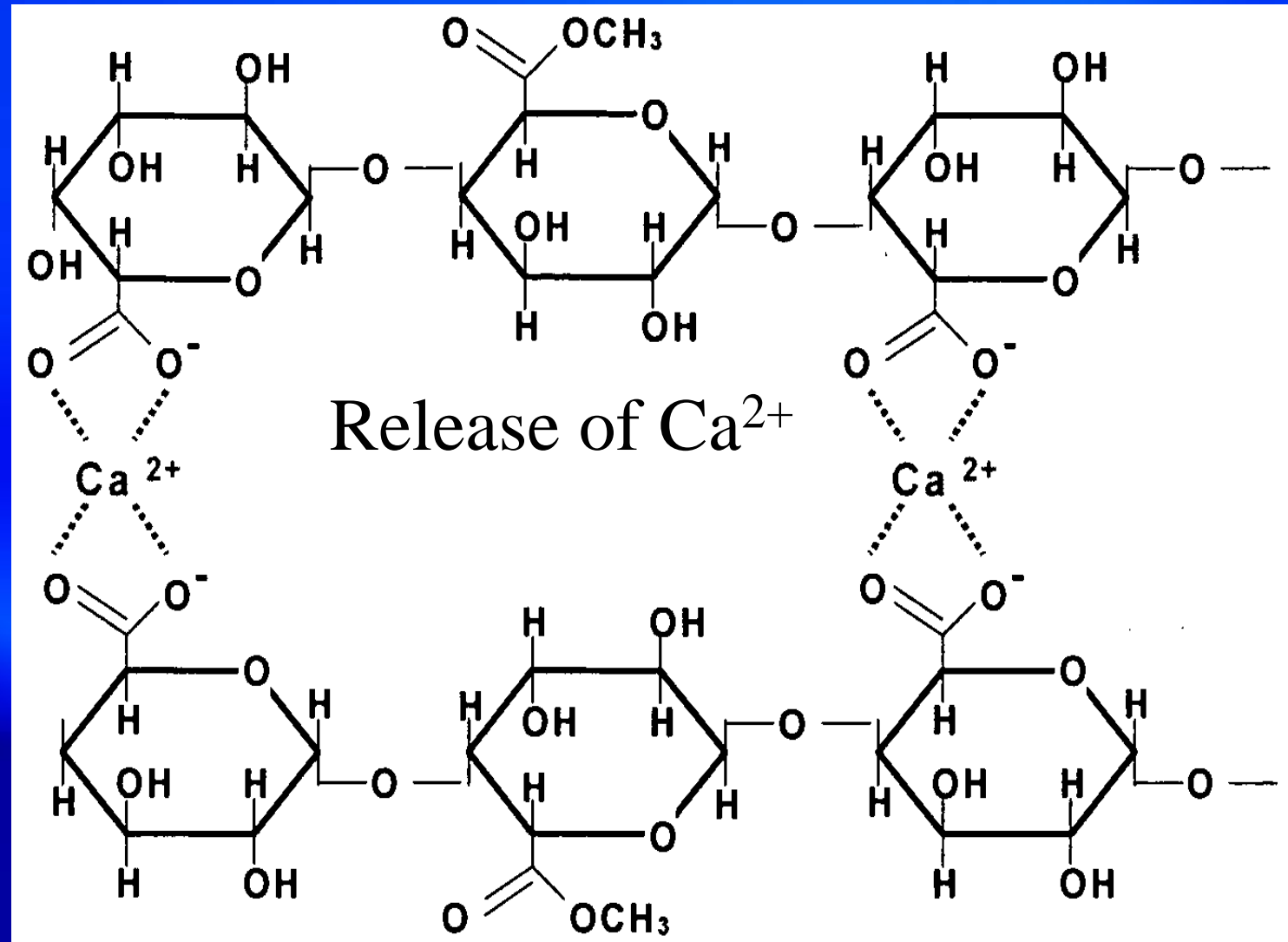
Pectic materials in the cell walls

- Pectic material is the main component of the middle lamella and primary wall.
- Pectic material contains α -1,4- galacturonic acid chains with rhamnose α -1,2 linkage
- 16% rammnogalacturonam
- 9% galactan, galactose in β -1,4 linkage
- 9% arabinan, arabinose in α -1,5 linkage
- Cellulose and hemi-cellulose is the main component of the secondary wall
- Glucose in β -1,4 linkage

Pectolytic enzymes

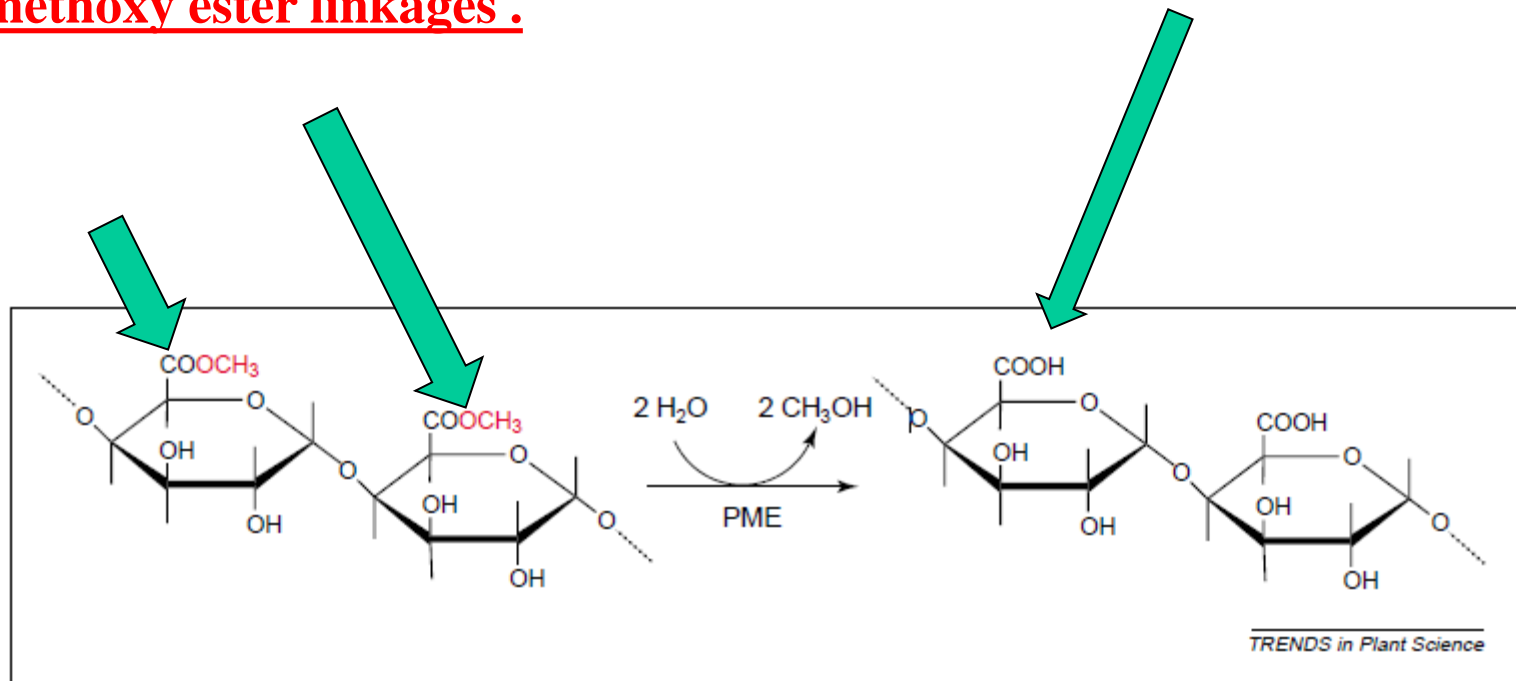
- Pectin methyl esterases (PME)
- Polygalacturonase (PG)
- Pectate lyase (PL)

PME activity first step: The release of Ca^{2+} between the glacturonic acid chains



Pectin Methylesterases second step:

Pectin methylesterase catalyzes the de-esterification through transferring the C6 carboxyl groups to water molecules altering the degree and pattern of methyl esterification and resulting in the formation of high molecular weight pectins with new **non-methoxy ester linkages**.



Pectolytic enzymes

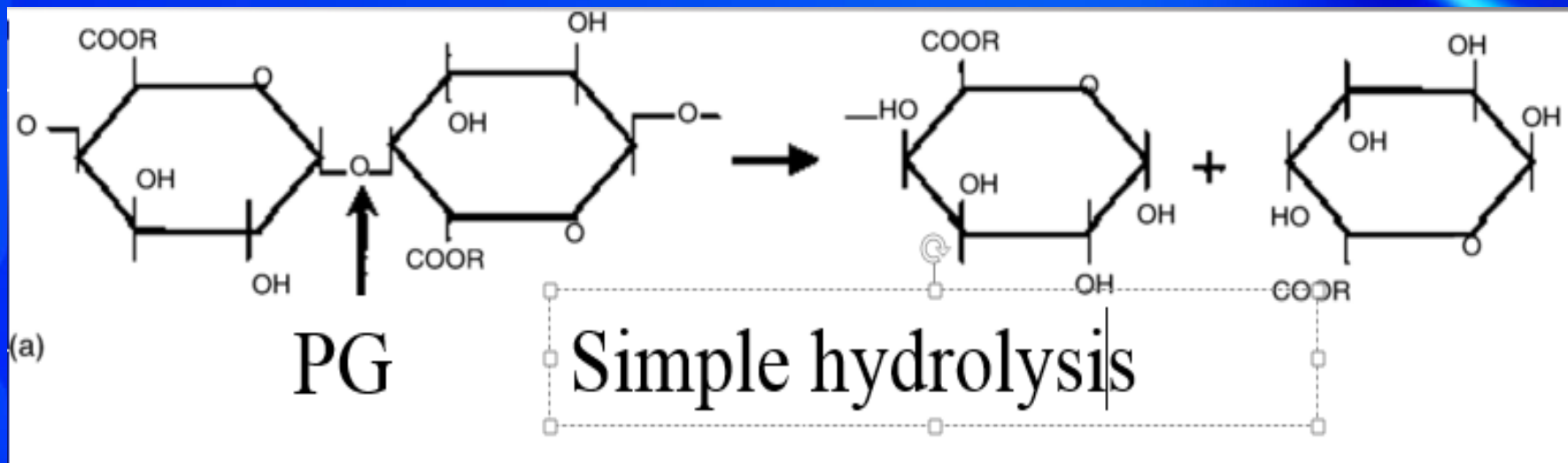
Polyglacturonase (PG)

Pectate lyase (PL)

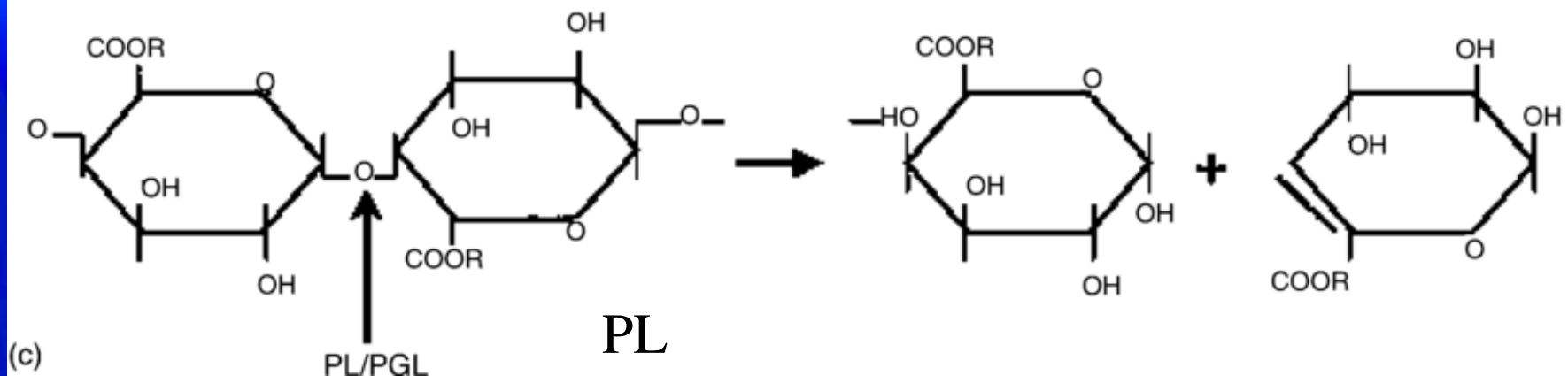
Both catalyze the reaction at the α -1,4 binding site

PG by hydrolysis

PL by trans-elimination



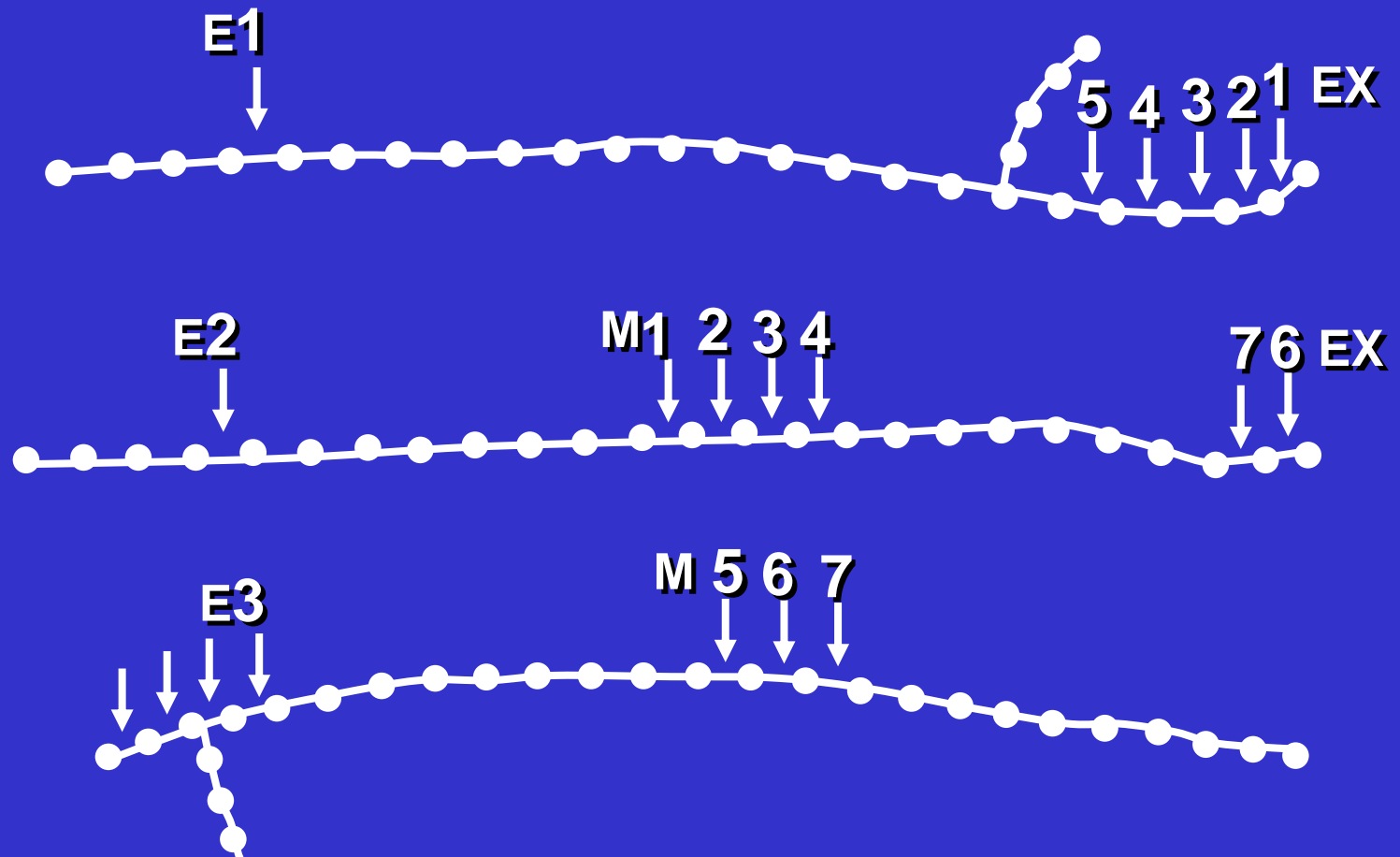
Hydrolysis by trans-elimination



Pectolytic enzymes

- **Pectolytic activity with endo and exo activity**

Endo and exo digesting enzymes



Inducible enzymes

- **Pectolytic enzymes and inducible enzymes**

In vivo and Vitro

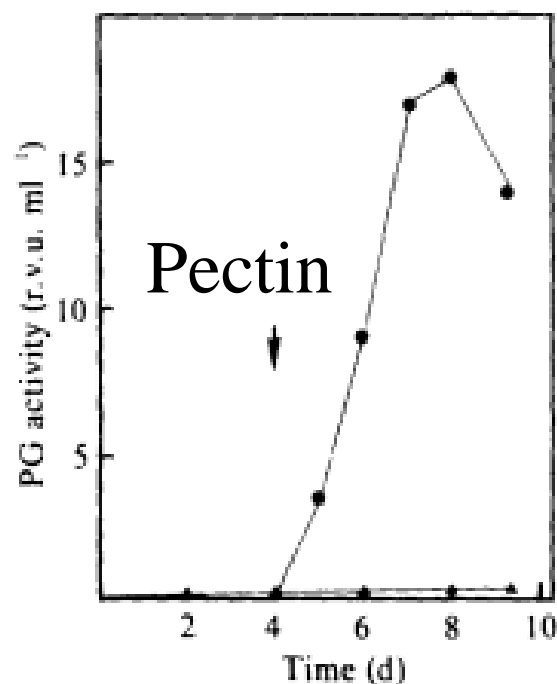
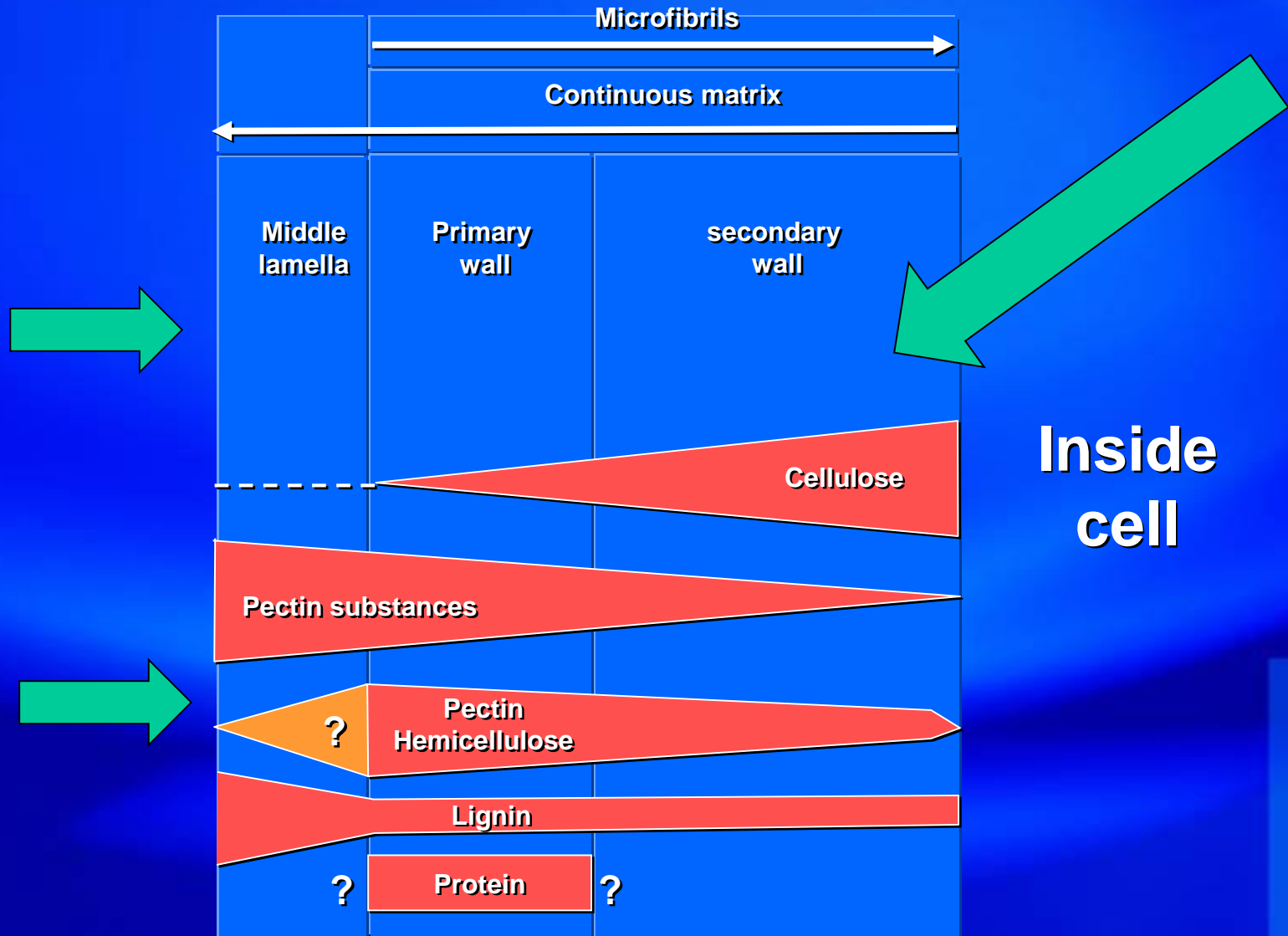


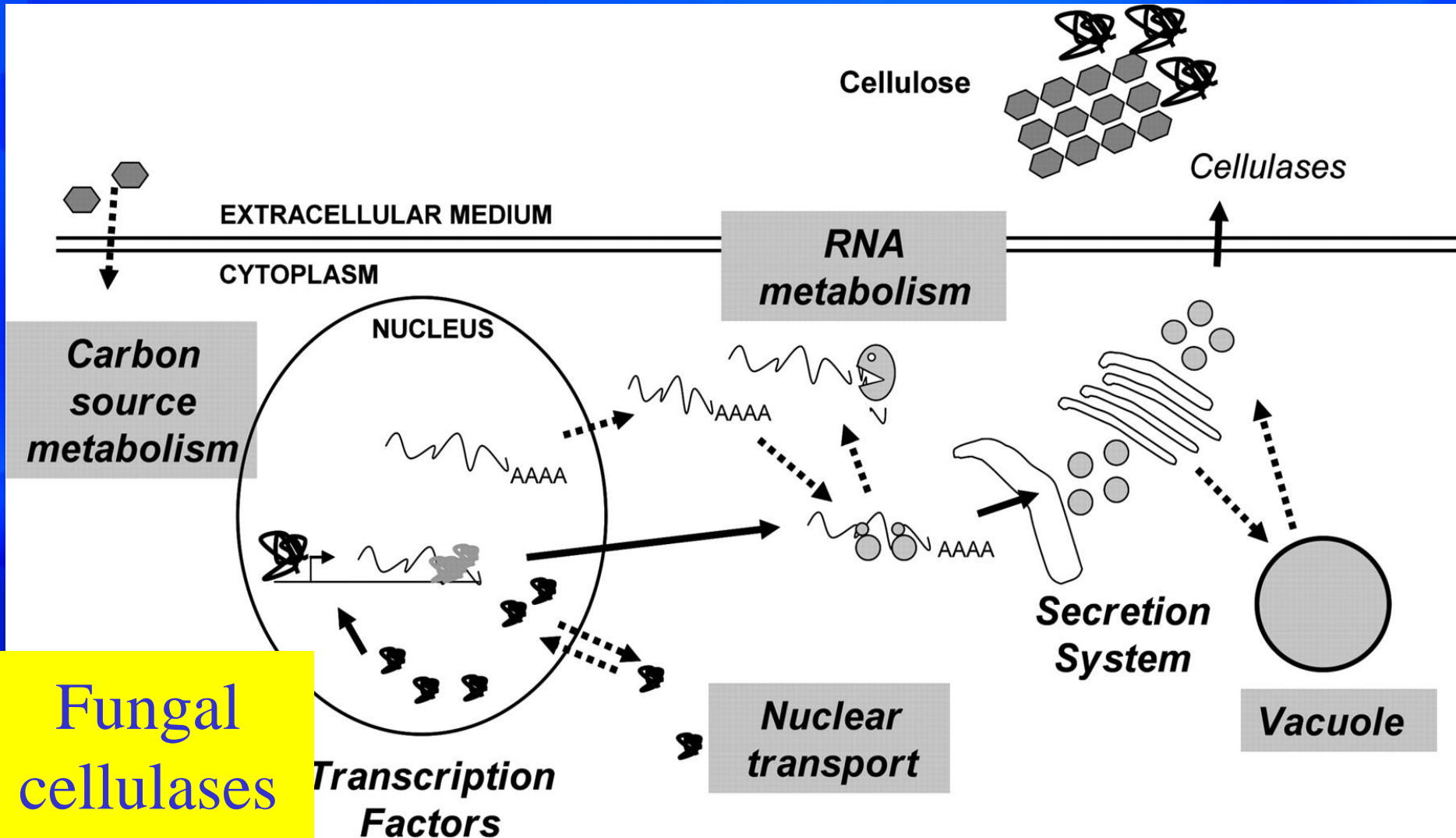
Fig. 1. Time-course of PG induction by pectin in glucose-grown *F. moniliforme*. After 4 d of growth in glucose, the mycelium was transferred to a pectin-containing medium. The mycelium was harvested 2 d later and mRNA was isolated. Other experimental conditions are described in the text. ●, Pectin-induced; ▲, non-induced.

General Cell wall structure

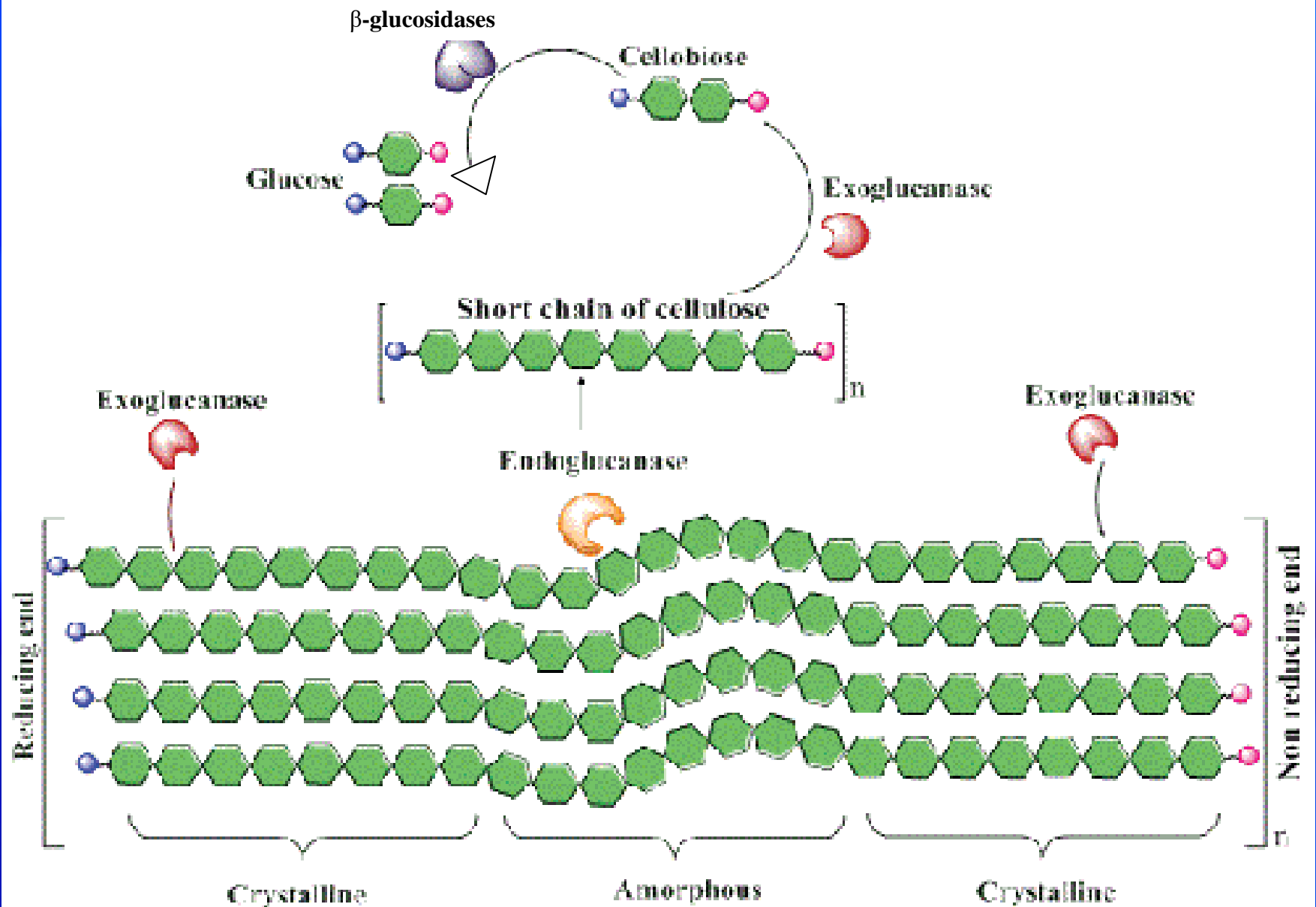
Fungal Attack



Penetration of the Host secondary wall



Fungal Cellulase



Pathogenicity of Postharvest pathogens

Pathogenicity of postharvest pathogens:

Methods of study

- **Gene expression**
- **Development of mutants with inhibited secretion**
- **Development of mutant with disrupted genes**

Pectolytic enzymes as enzymes contributing to pathogenicity: gene expression

Penicillium sp.

Sclerotinia sclerotiorum

Colletotrichum gloeosporioides



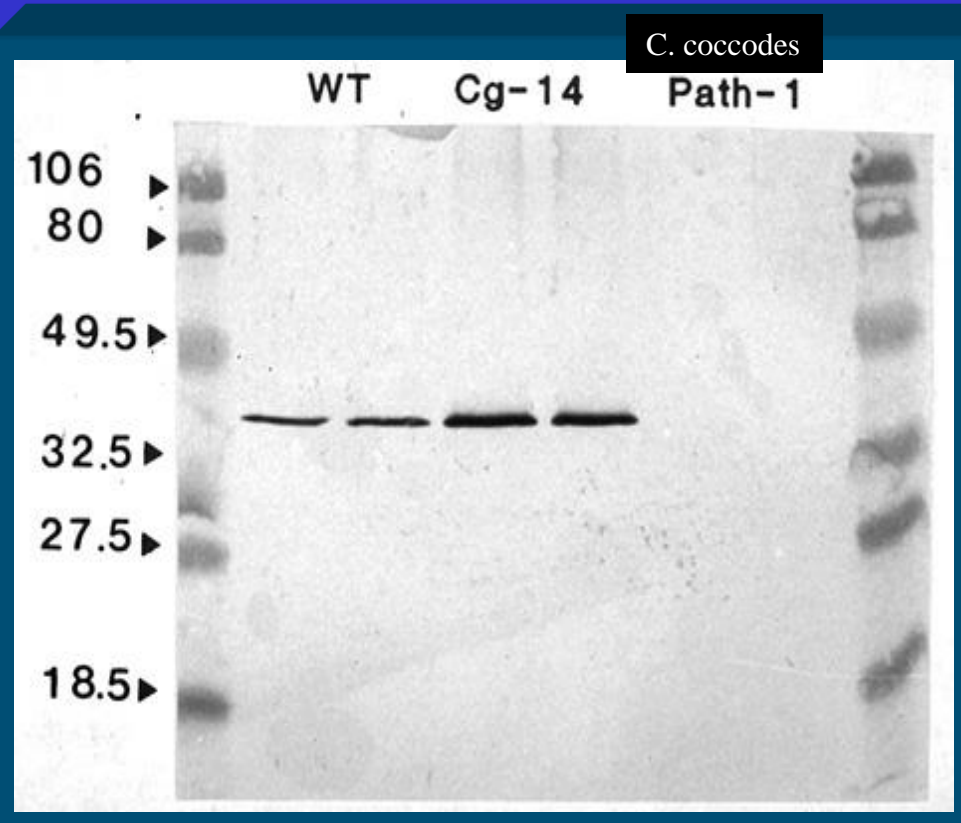
Development of an non-secreting mutant of *Colletotrichum*

Cg -14

WT

Path - 1

C. coccodes

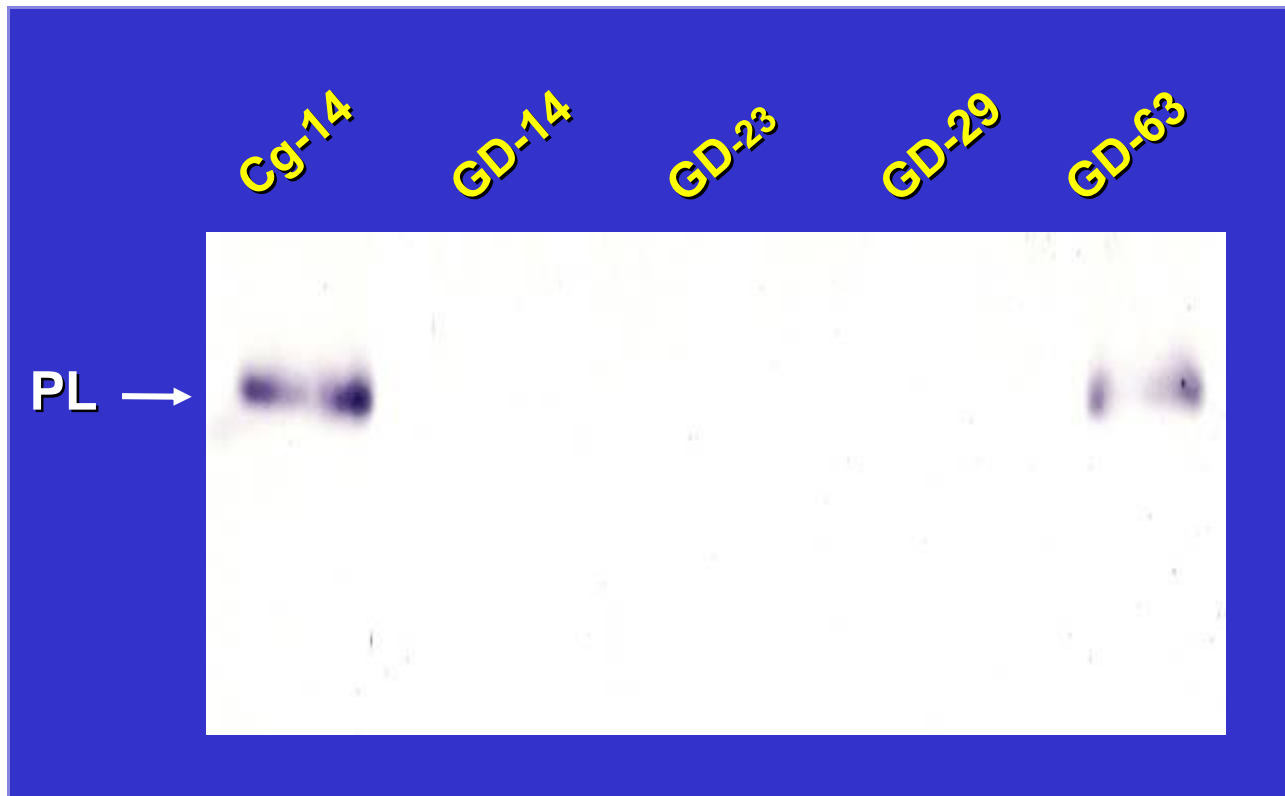


Pathogenicity of postharvest pathogens:

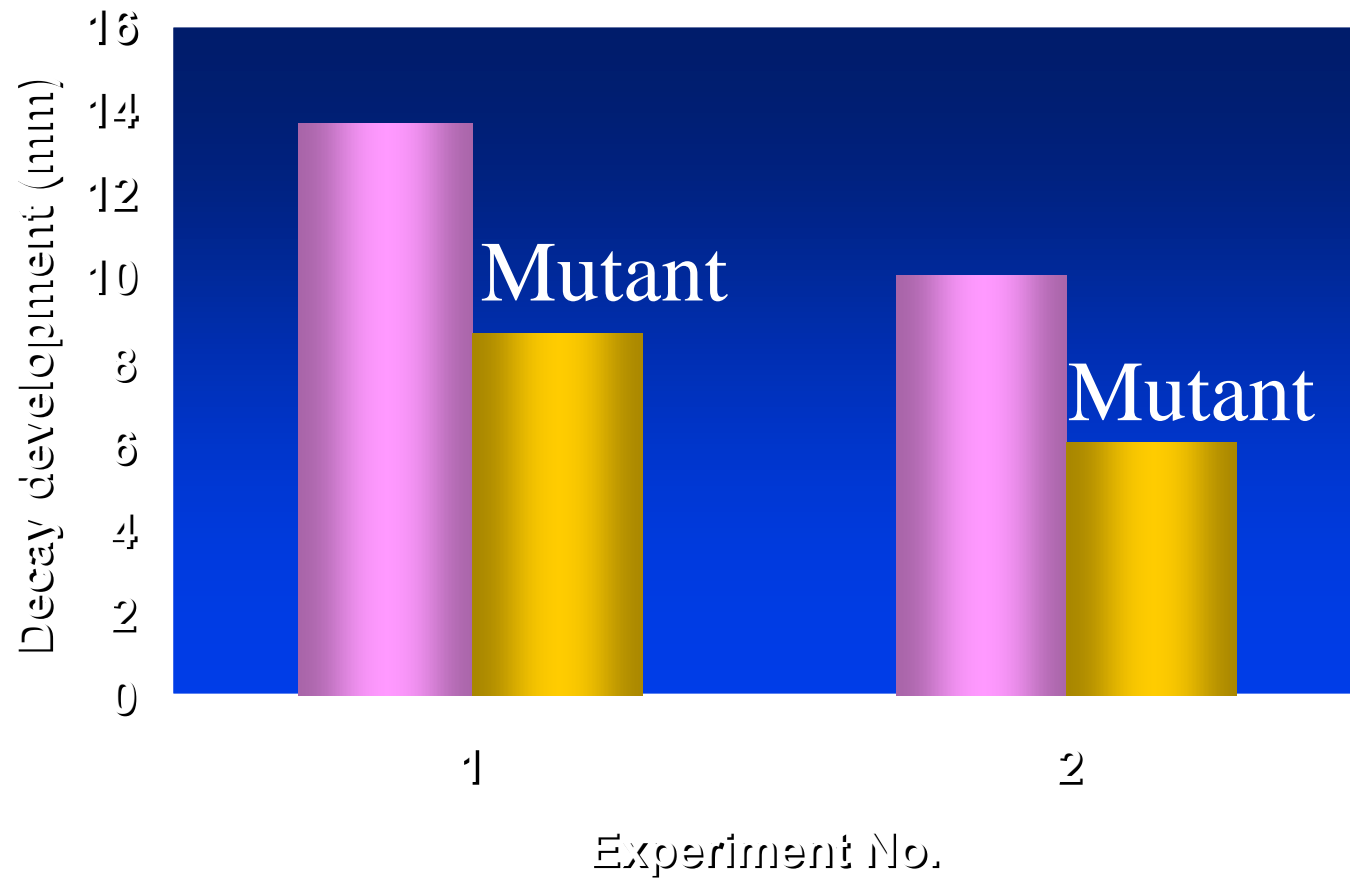
Methods of study

- **Gene expression**
- **Development of mutants with inhibited secretion**
- **Development of mutant with disrupted genes**

Western blot of Pectate Lyase secreted by *C. gloeosporioides*, the disrupted mutants and the Hyg-resistant non disrupted mutant



***Decay development affected by
mutants***
C. gloeosporioides pel/B mutant Cg-14



Pathogenicity factors expressed in postharvest pathogens

PL is a gene that contribute to 30-40% of infection capabilities of *Colletotrichum*

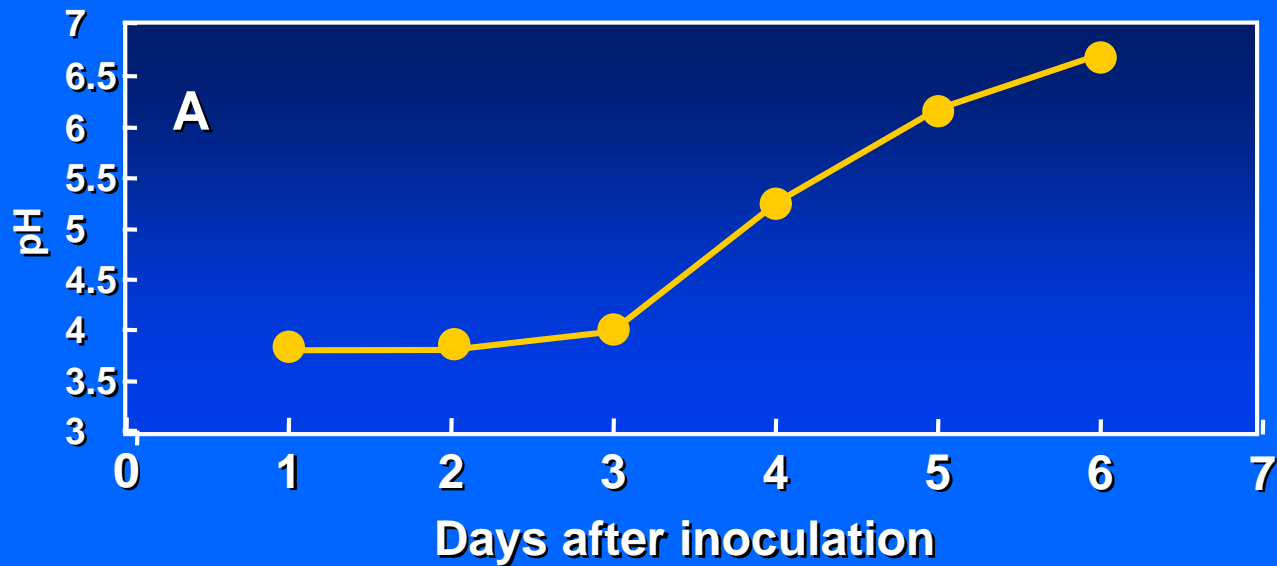
Other enzymes contribute to the other part of the maceration capabilities of the fungus

Regulation of Pathogenicity factors by postharvest pathogens

The regulation of pH

Changes in media pH during *C. gloeosporioides* growth

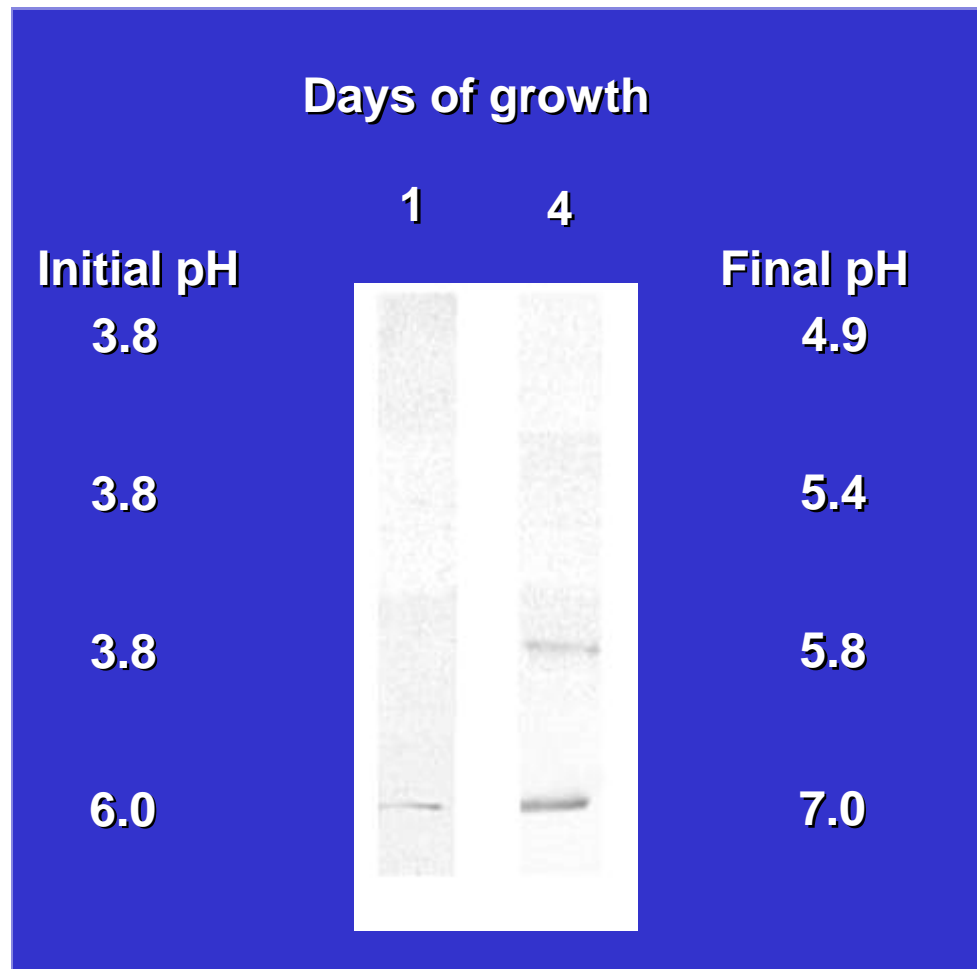
Acidic yeast extract media at pH 4.0



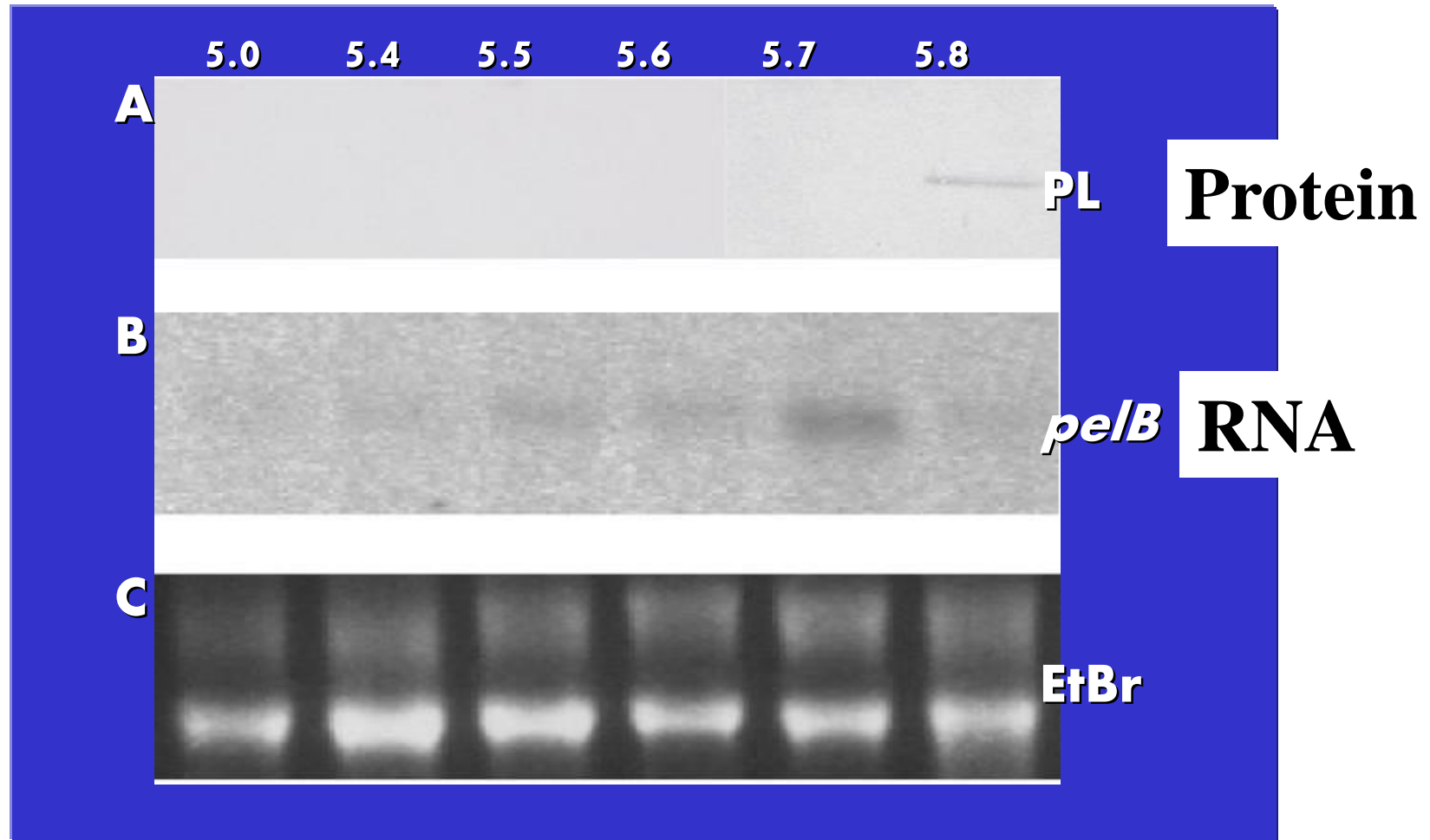
B PL →



Effect of media pH on Pectate Lyase secretion by *C. gloeosporioides*

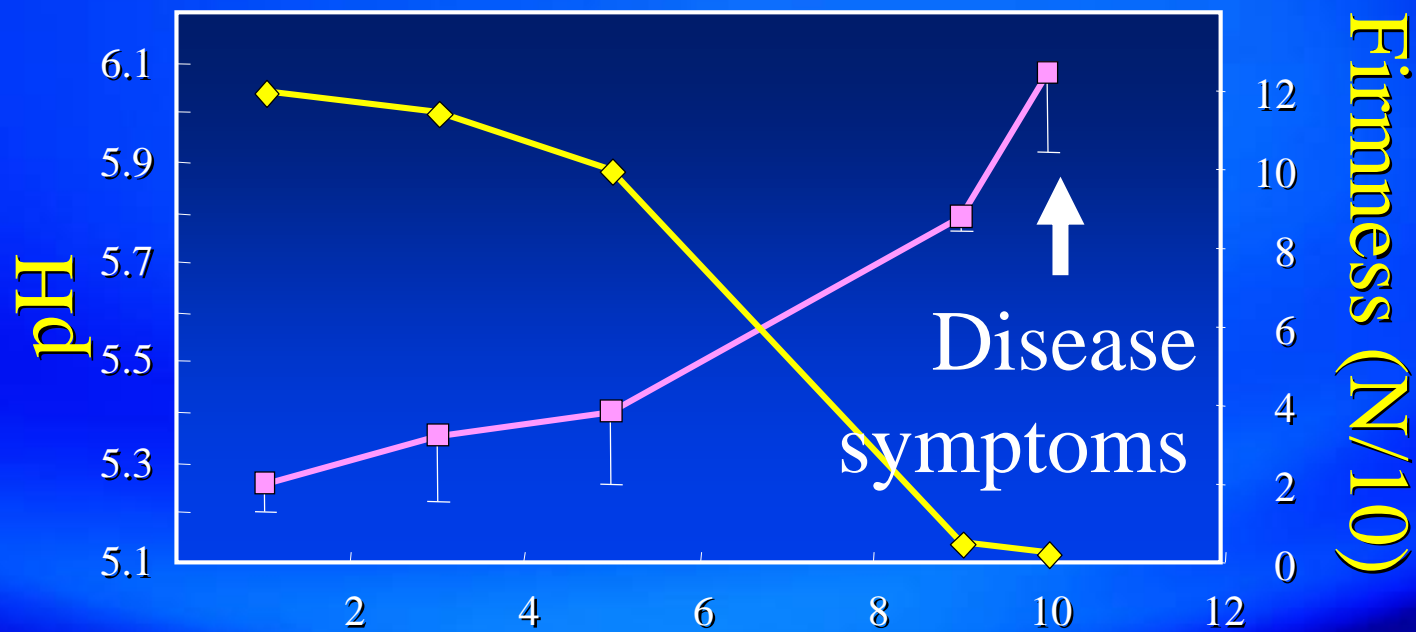


Expression of *pelB* and secretion of PL by *C. gloeosporioides* as a function of pH



**Is the host capable of modulating
the differential expression of
pectolytic enzymes so the fungus
modulate it's pathogenicity**

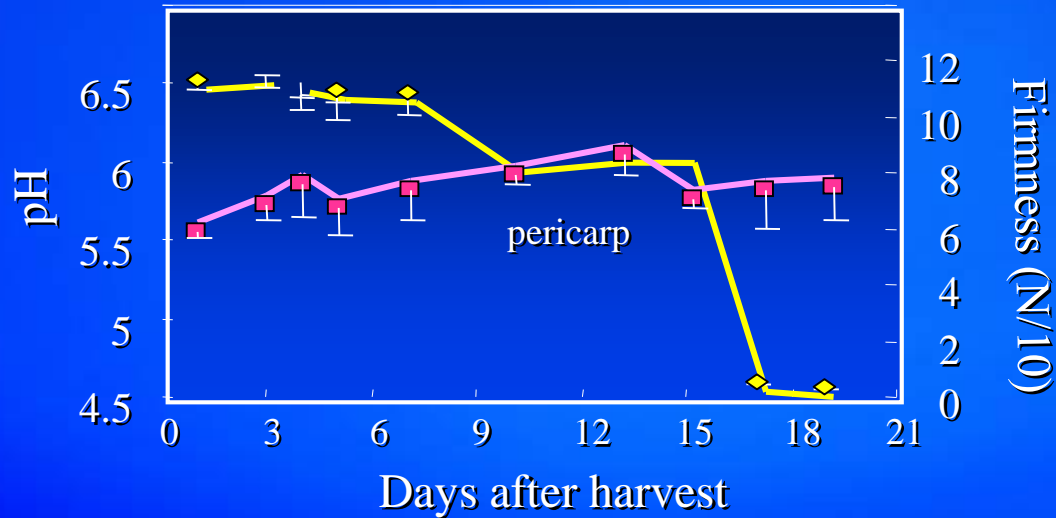
Changes in pH at the peel of the avocado fruit cv. Fuerte during fruit ripening



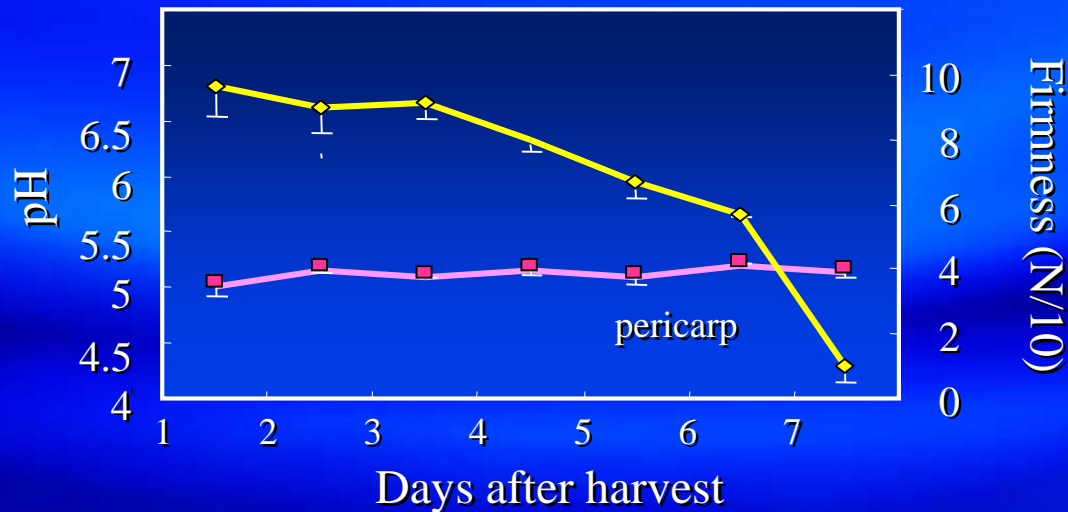
Days after harvest and inoculation

Changes in pH of two cultivars of avocado fruits

Fuerte



Ettinger



Ettinger
Lower pH

Fuerte
Higher pH

Ettinger



cv. Ettinger



Pectate lyase
inducing media at pH 3.8

cv. Ettinger



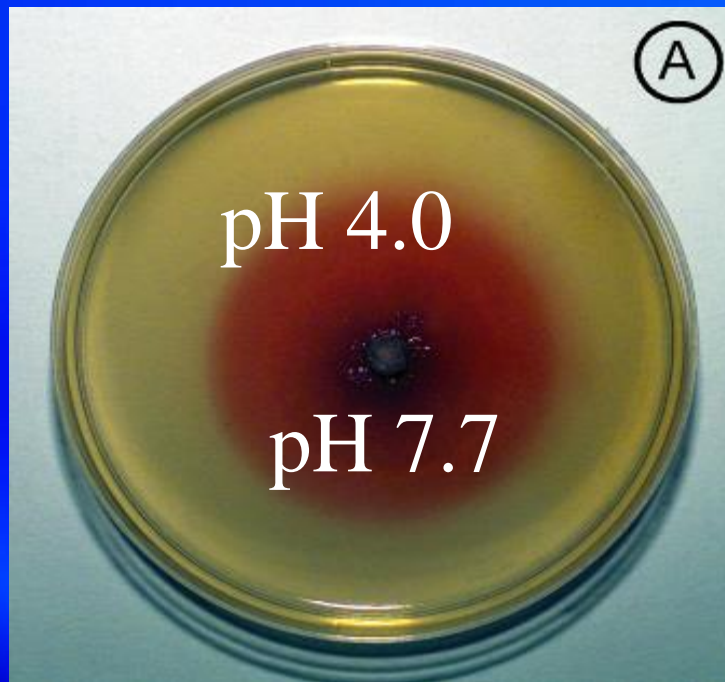
Pectate lyase
inducing media at pH 6.0

The pH of the host peel
modulates the delivery
and expression of
pectolytic enzymes for
colonization

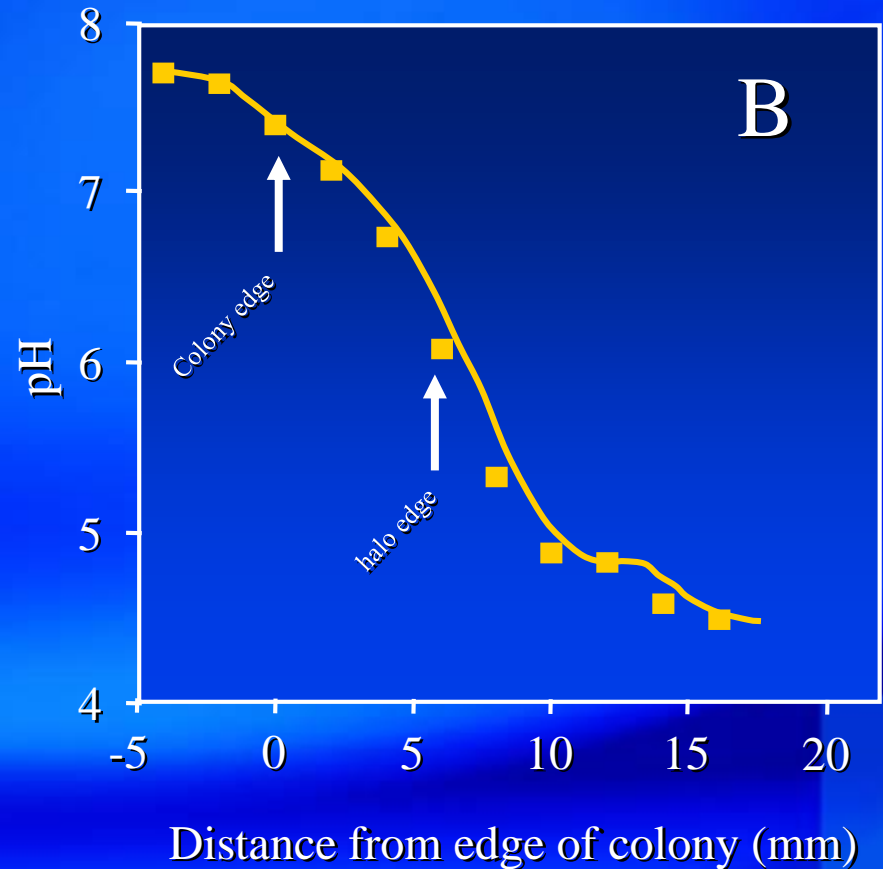
The second question:

Can the pathogen
modulate the expression
of it's pathogenicity
genes to enhance its
colonization?

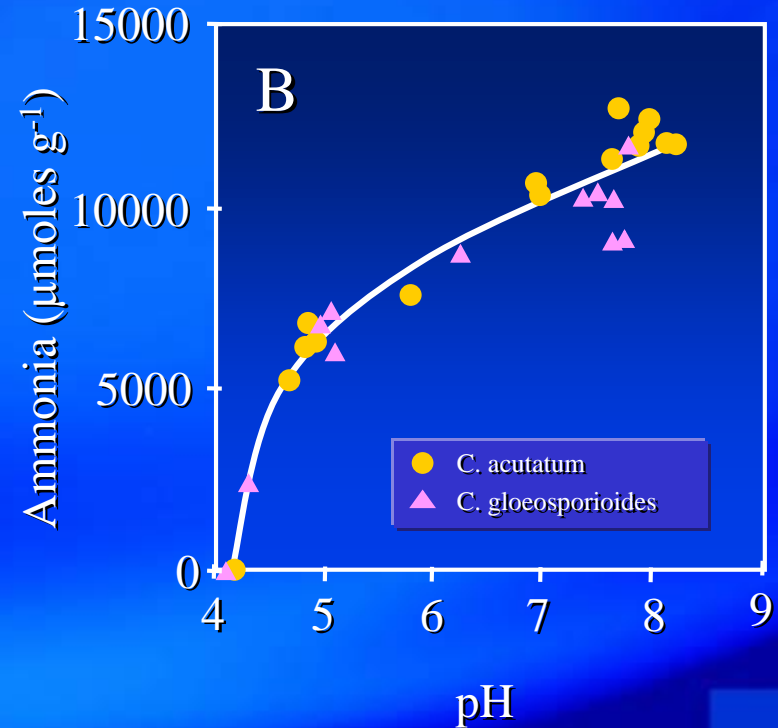
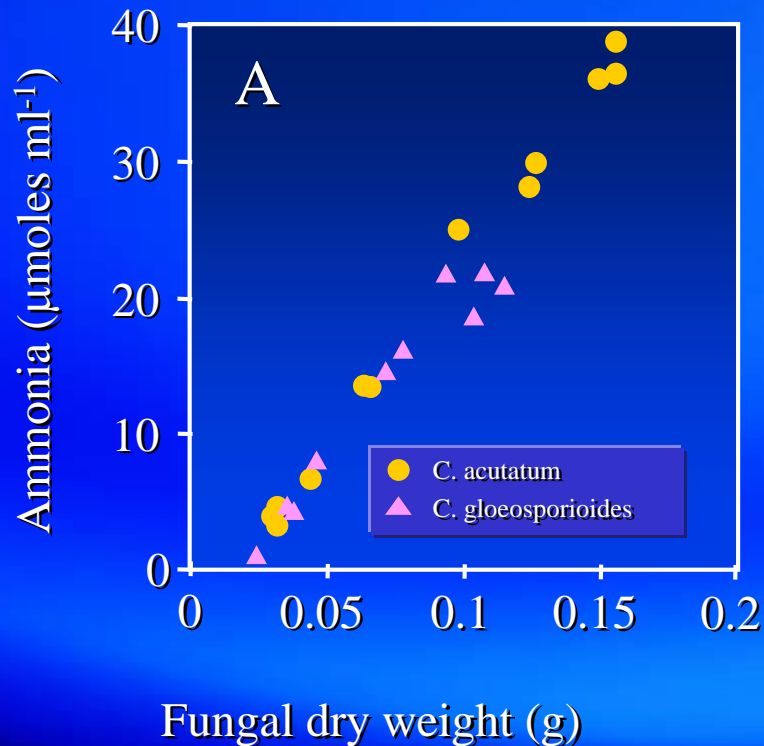
Differences in pH values induced by *C. gloeosporioides* in YE media at pH 4.0



With Alizarin Red

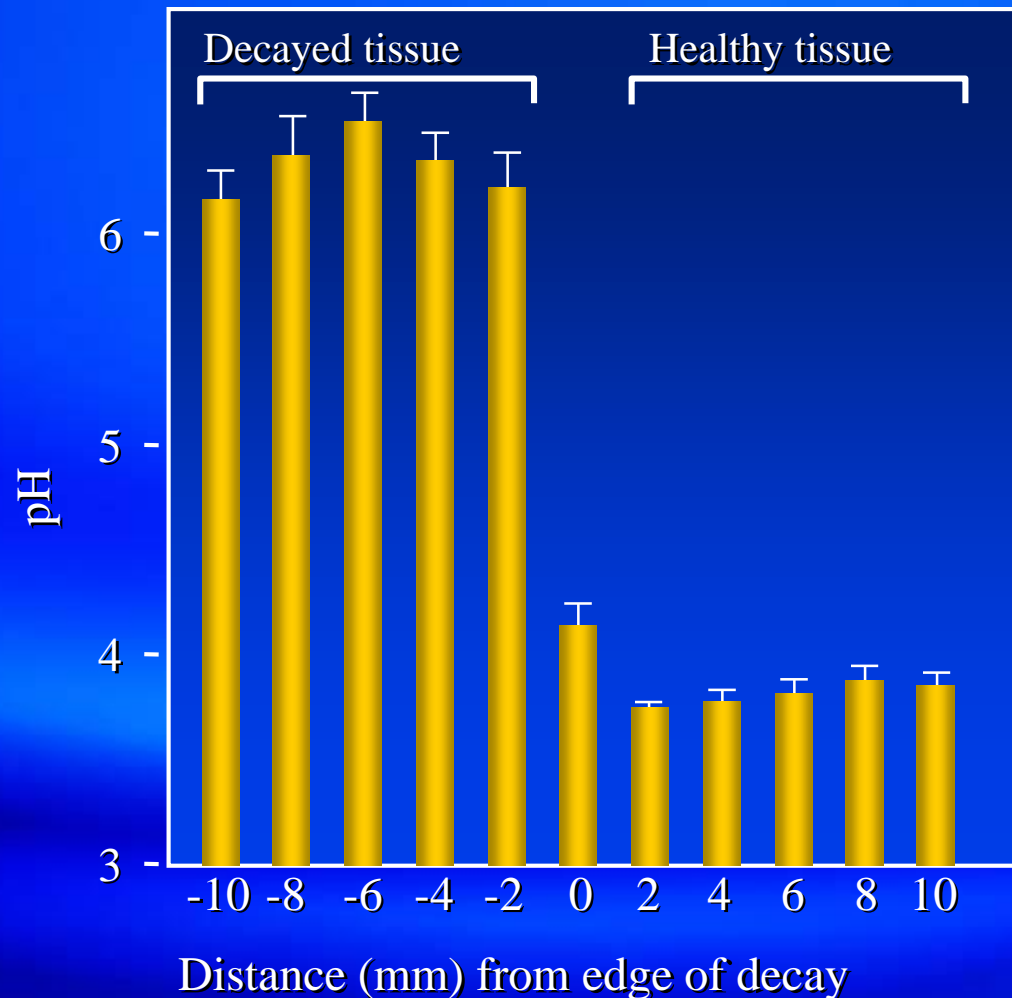


Changes in pH and ammonia accumulation induced by *C. gloeosporioides* in acidified YE media to pH 4.0

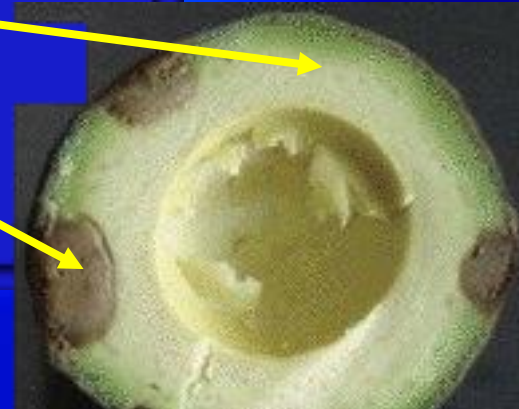
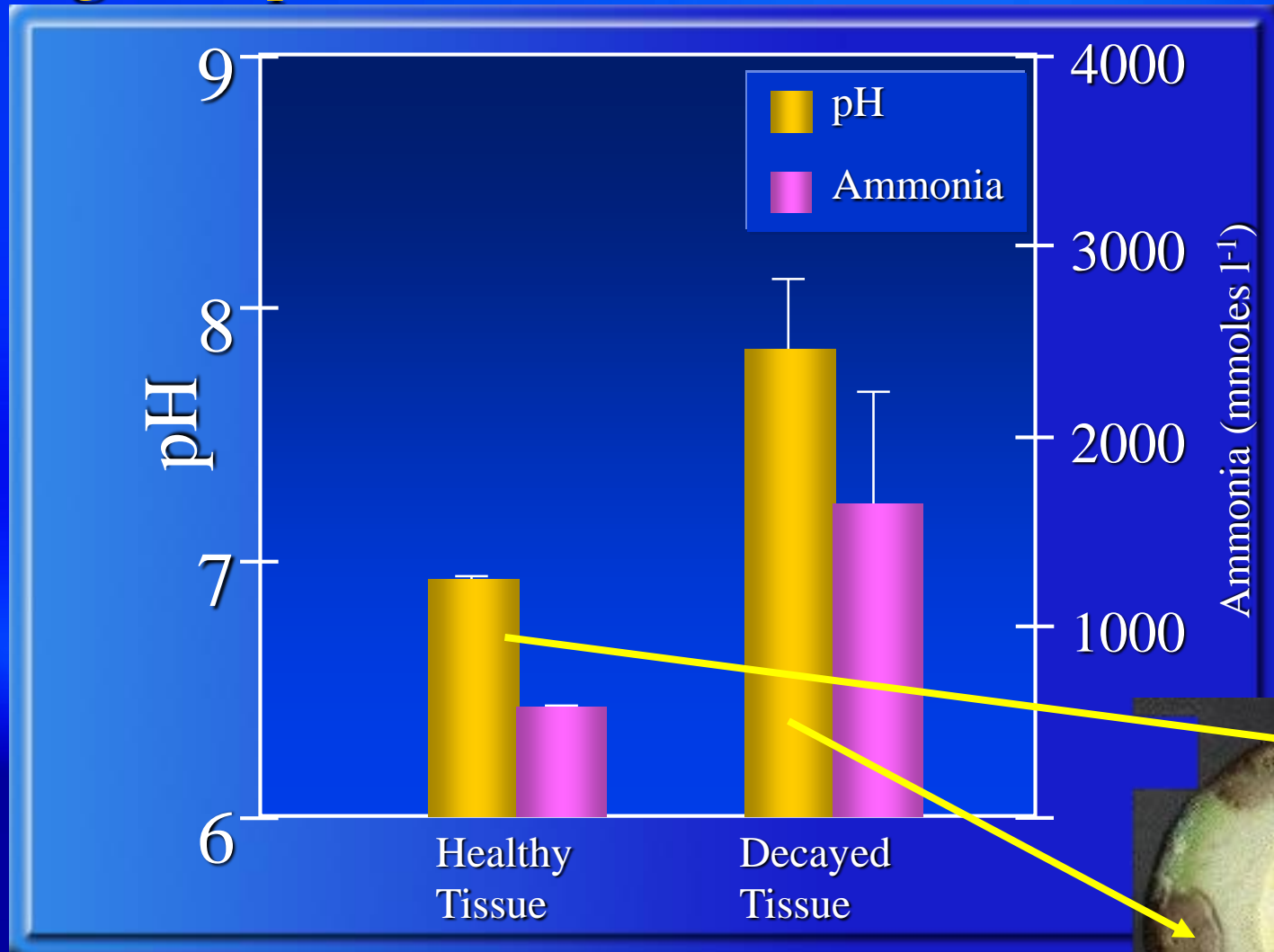


Prusky et al., 1991 MPMI

pH values at different distance of the leading edge of the decay caused by *C. acutatum* in apple fruits



pH changes and ammonia accumulation by *C. gloeosporioides* in avocado fruits cv. Hass



Transcriptional activation of *pelB* and PL secretion by *C. gloeosporioides* as a function of pH

pH	4.2	4.5	4.9	5.4	5.9	6.0	6.3
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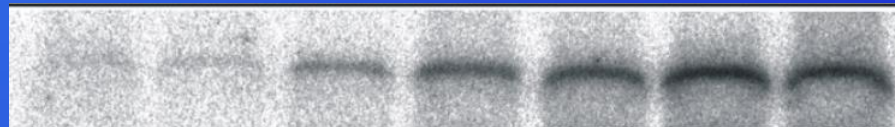
time (h)	6	8	10	12	14	16	18
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PL



Protein

pelB



RNA

rRNA



Summary of factors of pathogenicity

1. The pathogen produce pectinases (PME, PG and PelB)
2. The pathogen produces also cellulases
3. The activation of enzymes are inductive by the presence of substrates
4. The activation of genes and enzymes are pH dependent

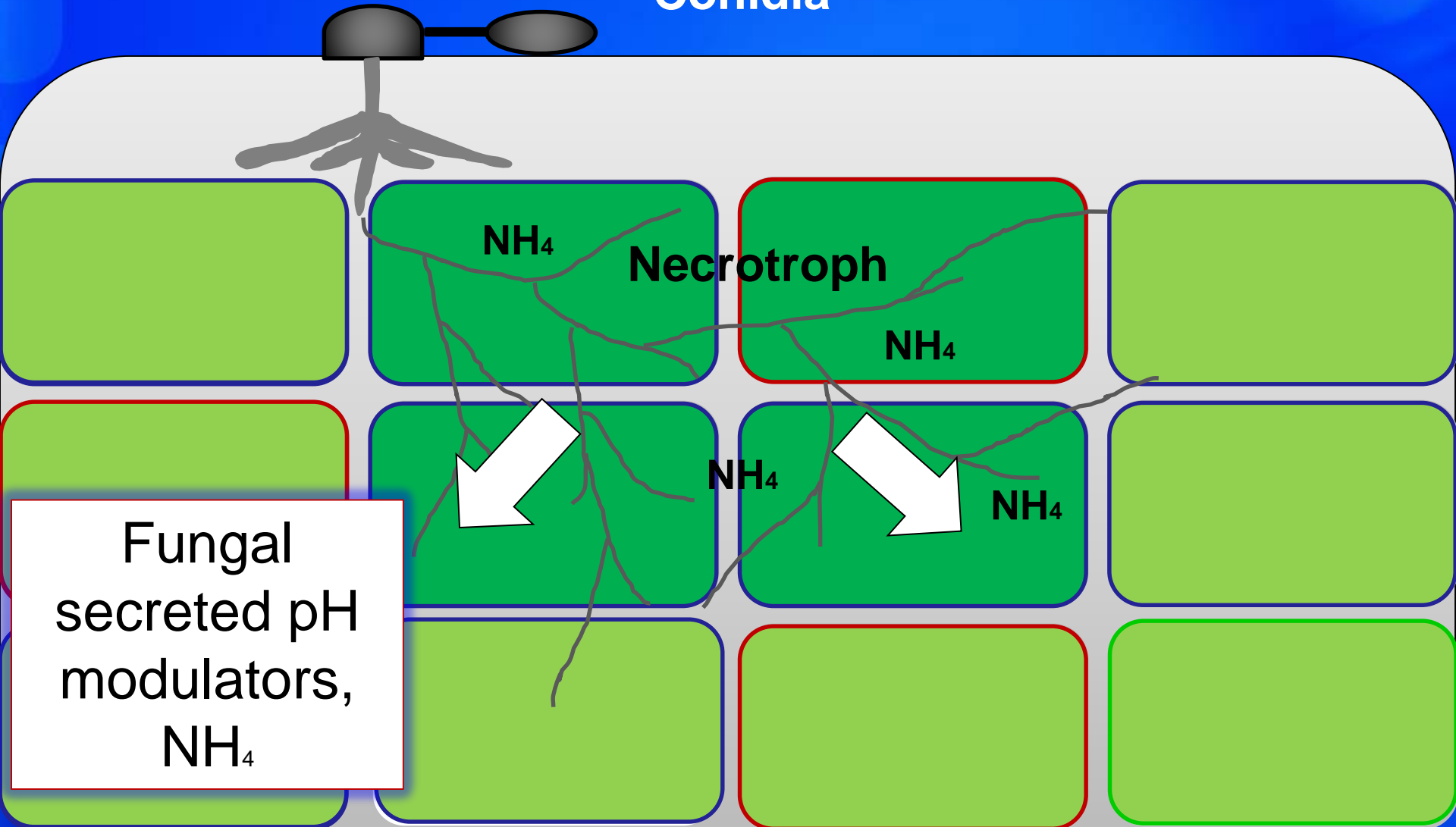
Case studies

Alkalinization of the host tissue

Alkalization by *Colletotrichum* in ripe fruits

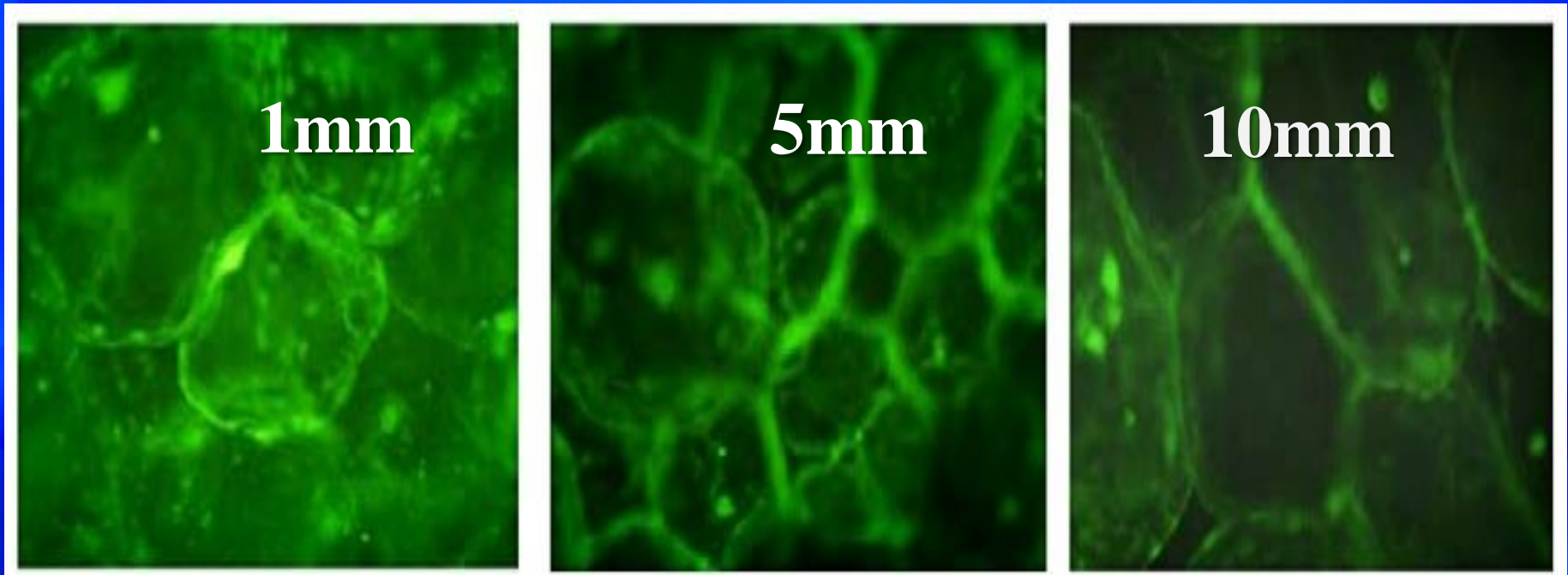
Appressoria

Conidia



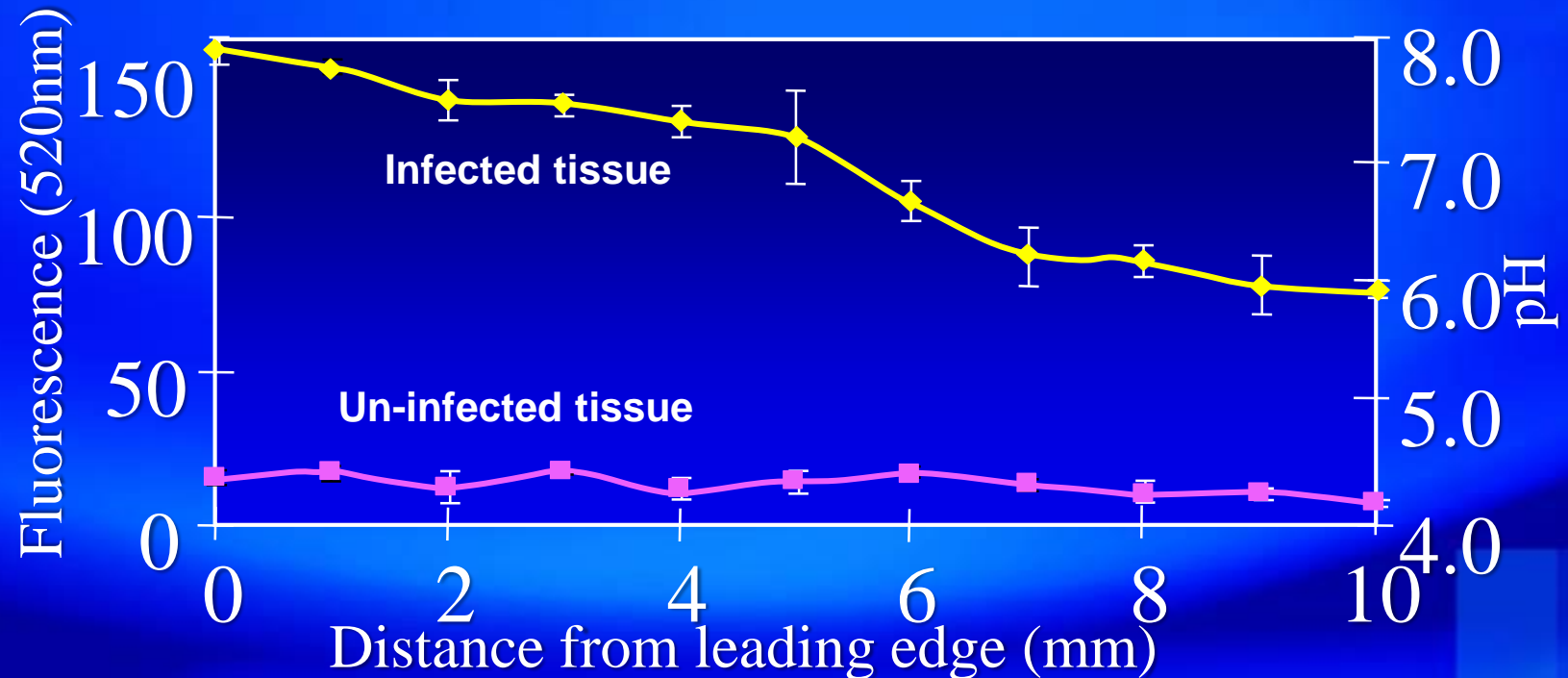
Local pH detected with BCECF in tomato fruit inoculated with *C. coccodes*

Distance from leading edge (mm)



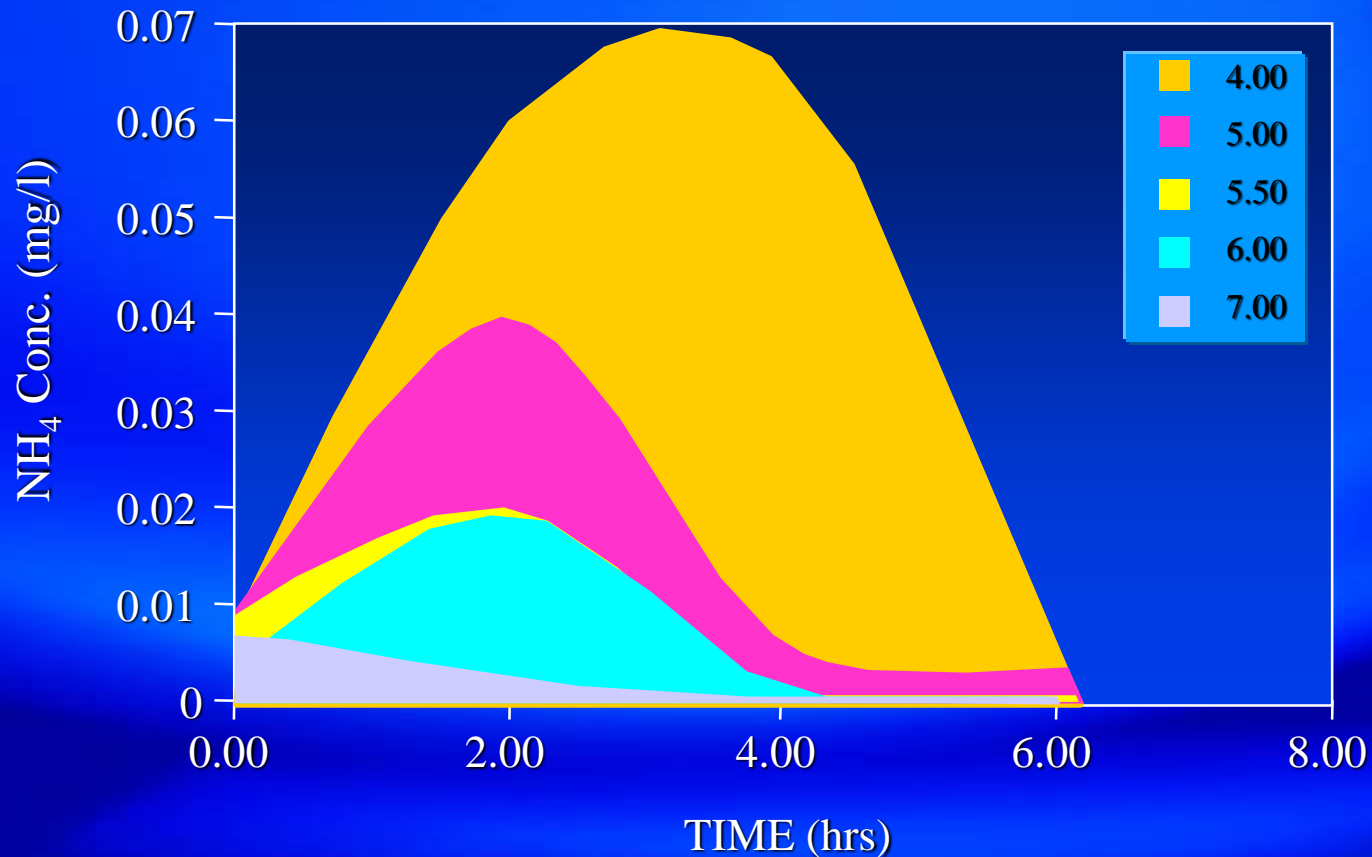
2',7'-bis(carboxyethyl)-5,6-carboxyfluorescein (BCECF),
fluorescent dye to measure intracellular pH

Local pH environment in tomato fruit at the leading edge of the decay caused by *C. coccodes*




What is the signal that
activates ammonia secretion?

Effect of pH on the inducing the secretion and accumulation of NH_4 by *C. gloeosporioides*?



What is the importance of pH modulation of the host tissue?

Secretion and accumulation of pectate lyase by *C. gloeosporioides* as a function of pH

pH	4.2 4.5 4.9 5.4 5.9 6.0 6.3						
time (h)	6	8	10	12	14	16	18
PL							

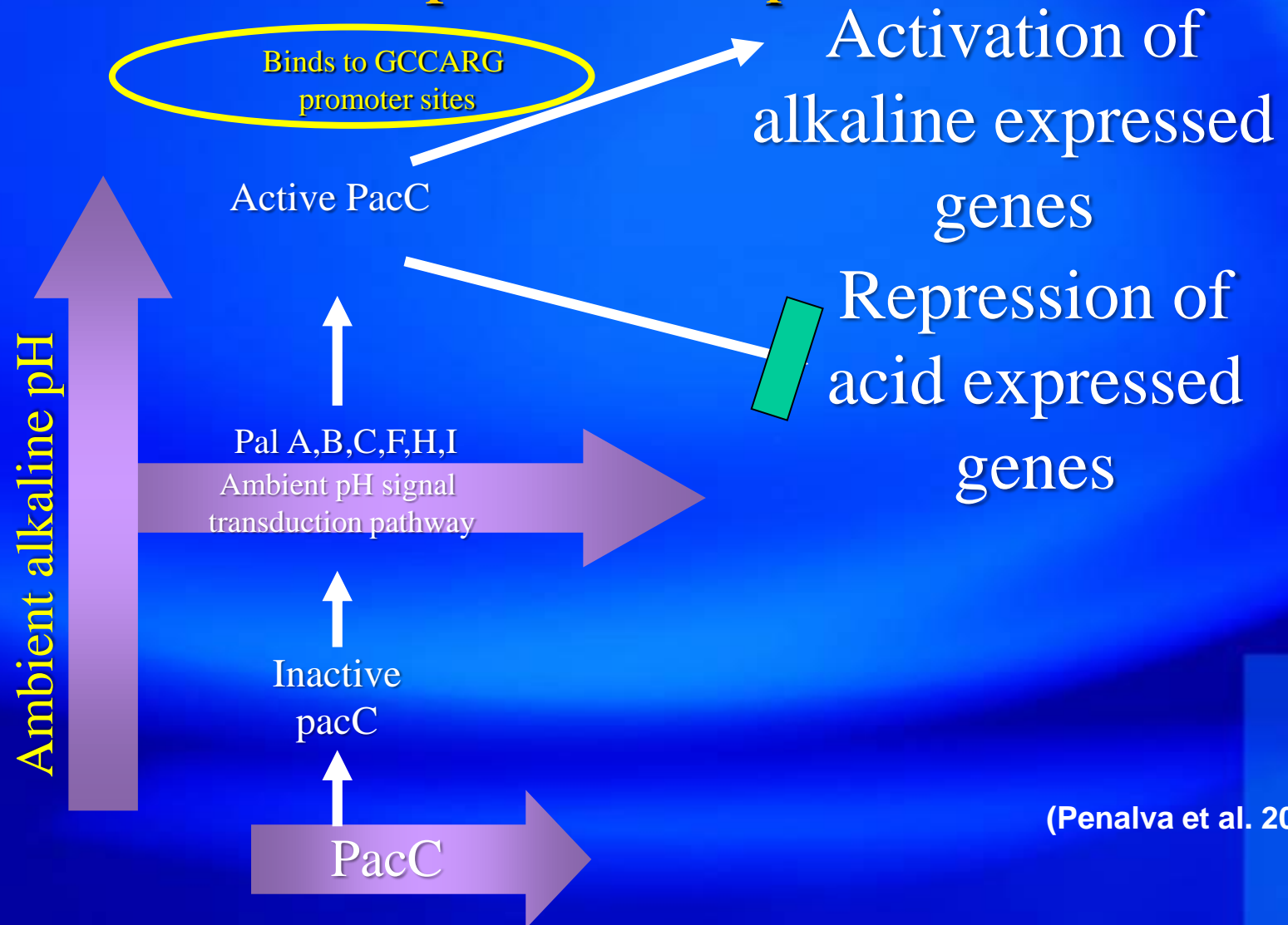
Increased maceration capabilities

pH modulation of the environment

enables the pathogen the
“selection” of specific virulence
factors needed for the particular
host

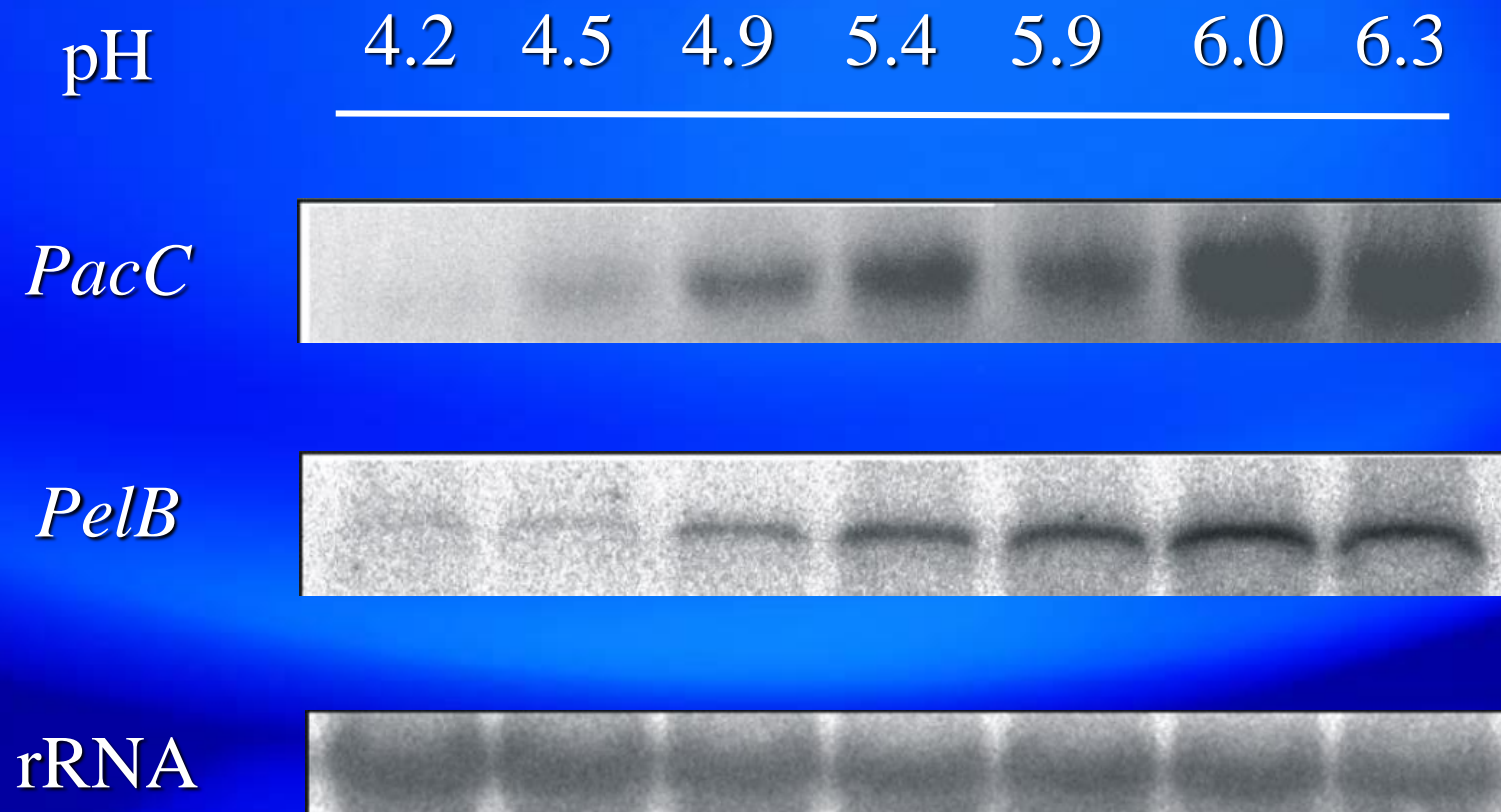
How does alkalization of the
tissue modulates gene
expression?

The modulation of genes at alkaline pH by the transcription factor *pacC*



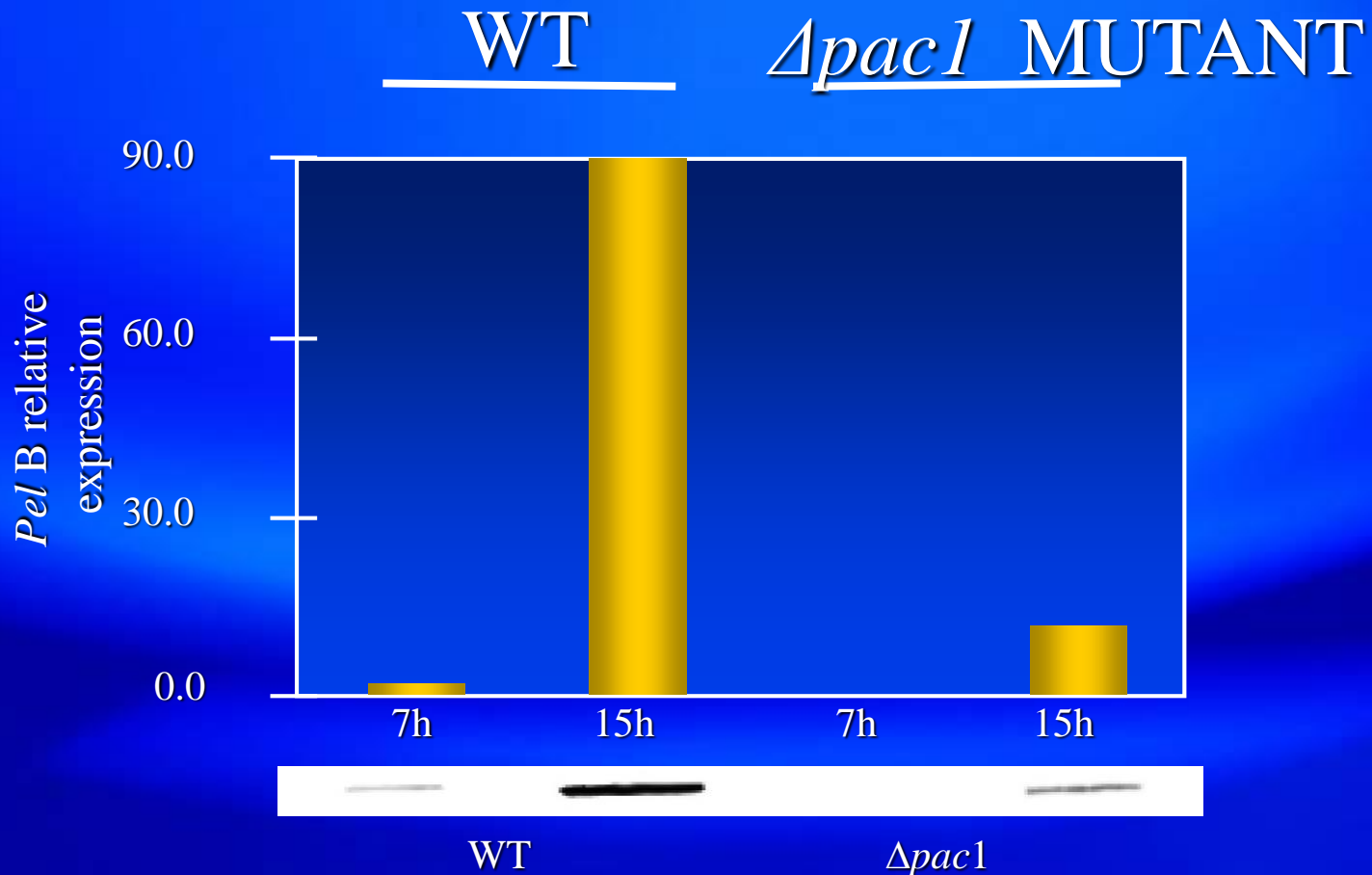
(Penalva et al. 2013)

Transcriptional activation of *PacC* and *pelB* from *C. gloeosporioides* as a function of pH



Knock out of *pacC*, the transcription factor that modulate gene expression at alkaline pH, down-regulated PL expression and reduced pathogenicity

**Transcriptional activation of *pelB* and PL secretion
detected by Western Blots
of the WT *C. gloeosporioides* and $\Delta pac1$ mutant at pH 6.0**

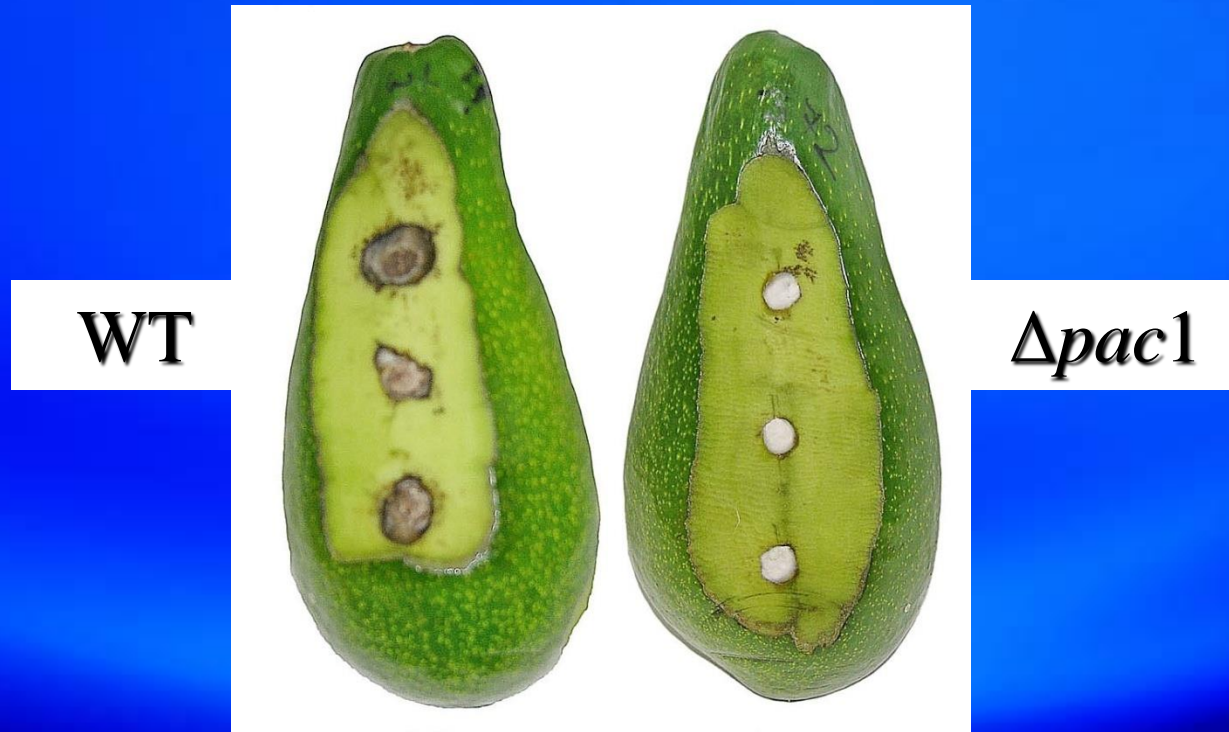


Effect of knock out of genes on reduced pathogenicity by *C. gloeosporioides*

Reduced Colonization (% control)

<i>pe/B</i>	25-30
<i>pacC</i>	70-80

Effect of *PacC* knock out on decay development



What is the purpose of the pH modulation?

- Specific activation of fungal genes
- Specific production pectolytic enzymes
- Enhance activity of the pectolytic enzymes

How is this carried out?

Activation of biochemical process regulating the production of

- ammonia

Second case studies:
pH regulators by
acidification of the tissue

The *Penicillium* case

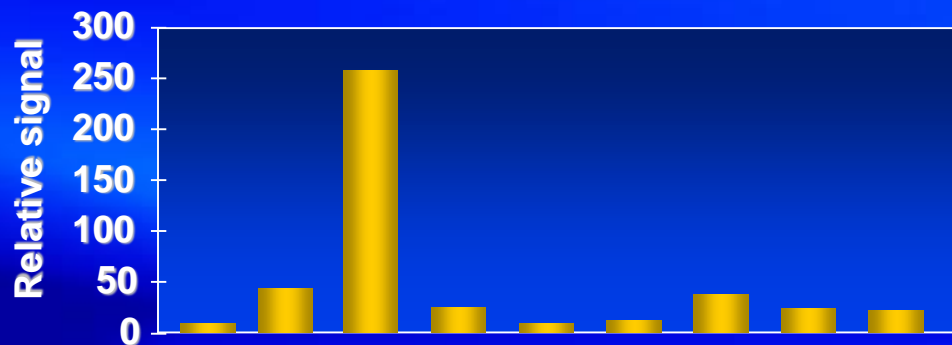
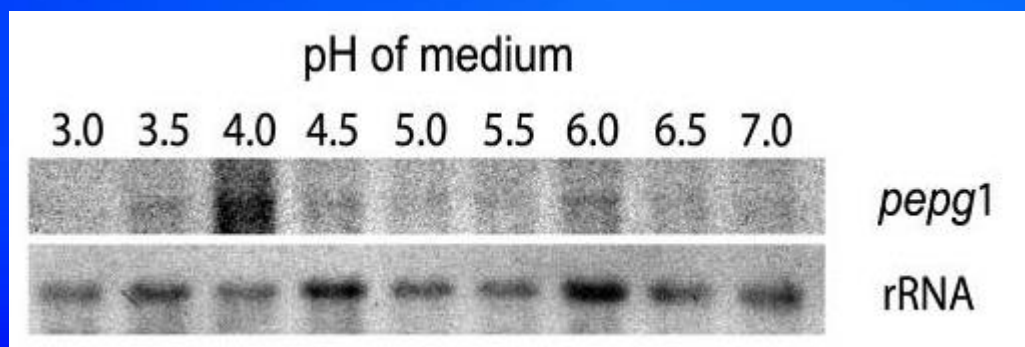
pH of healthy and decayed apple tissue inoculated by *P. expansum*

<u>Apple cultivars</u>	<u>Healthy</u>	<u>Decayed</u>
Fuji	4.57 a	3.92 b
Gala	4.35 a	3.91 b
Red Delicious	4.39 a	4.01 b
Granny Smith	4.04 a	3.65 b
Golden Delicious	4.60 a	3.80 b

**How does
acidification
modulate virulence
of *Penicillium***



Polyglacturonase, *pepg1* transcript accumulation of *P. expansum* in response to different ambient pH conditions



Corrected *pepg1* signals

**What is the
mechanism of
acidification of host
tissue**



Accumulation of organic acid in decayed apples by *P. expansum*

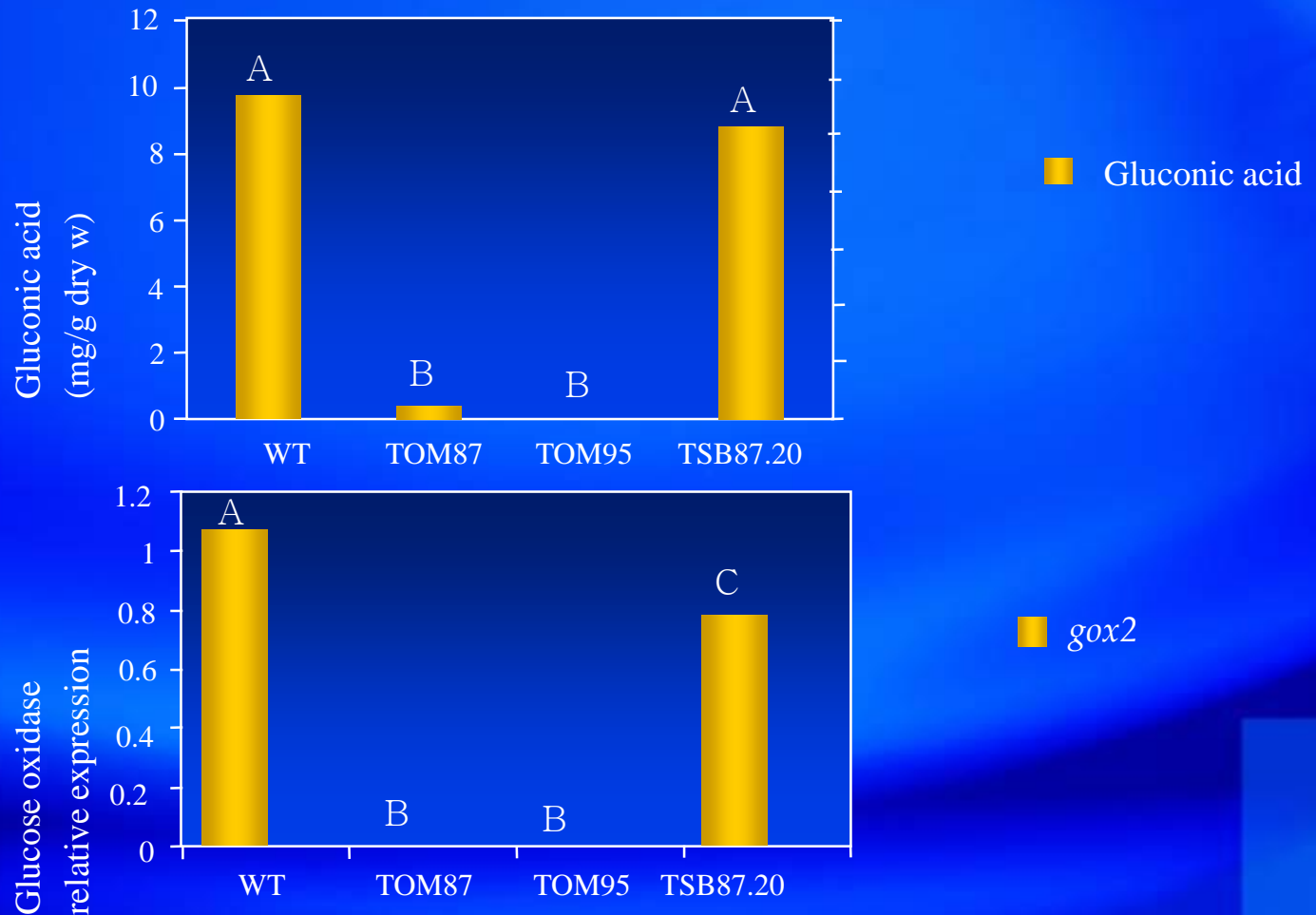
Organic acid ($\mu\text{g}/\text{gr fw}$)	Healthy	Decayed
Gluconic	182 \pm 39	2334 \pm 814
Citric	193 \pm 53	1410 \pm 211
Malic	4020 \pm 370	4380 \pm 680
Fumaric	1.4 \pm 0.3	15.8 \pm 1.8

Glucose oxidase



gox1 and gox2

Disruption of *gox2* reduced *gox2* relative expression and gluconic acid accumulation



What is the purpose of the pH modulation?

- Specific activation of fungal genes
- Specific production pectolytic enzymes
- Enhance activity of the pectolytic enzymes

How is this carried out?

Activation of biochemical process regulating the production of

- Gluconic acid and the acidification of the host tissue

**How is this mechanism
activated in vivo**

Low
TSS

% TSS

High
TSS

gdh2

NH_3

Alkalinization

?

gox2

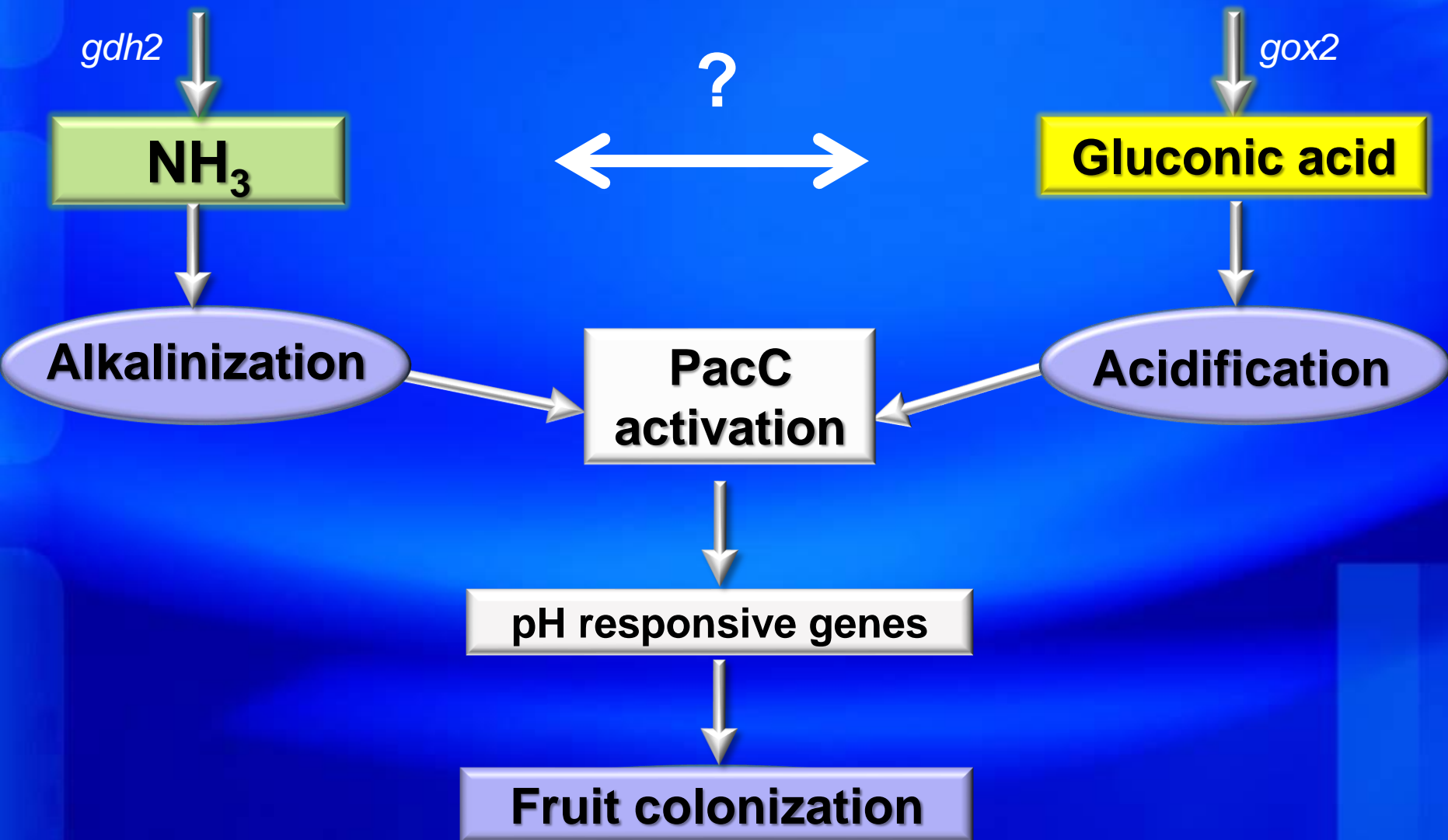
Gluconic acid

Acidification

PacC
activation

pH responsive genes

Fruit colonization



Biochemical factors regulating the synthesis of ammonia or organic acid

Carbon regulation

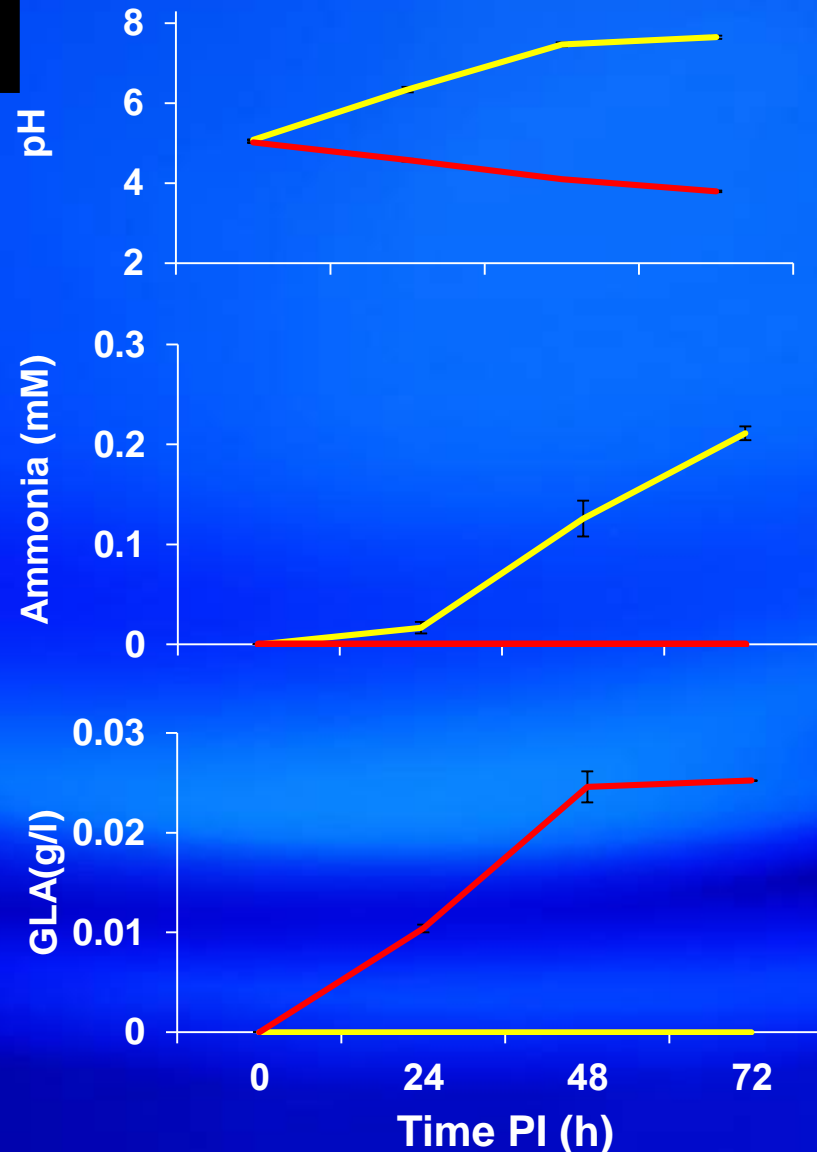
Effect sucrose concentration on pH modulation by *C. gloeosporioides*

■ 15mM Sucrose

■ 175mM Sucrose

Carbon limitation

Carbon excess



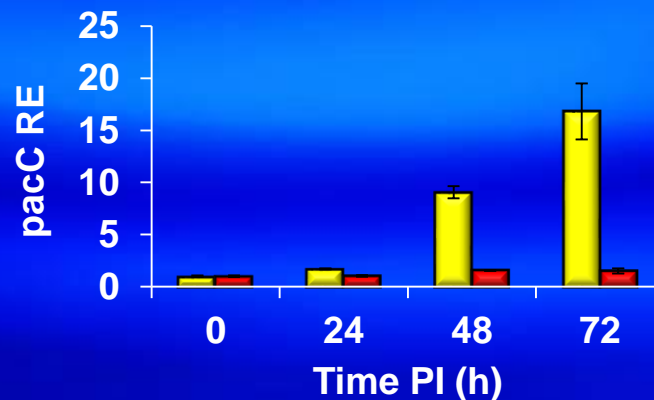
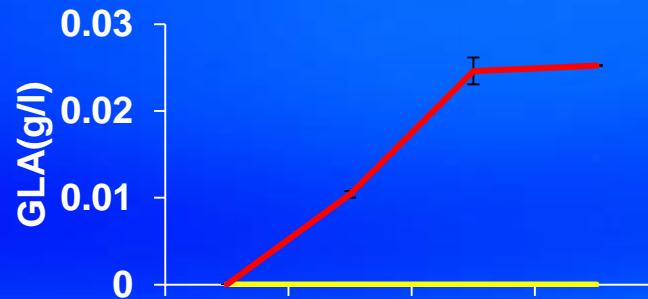
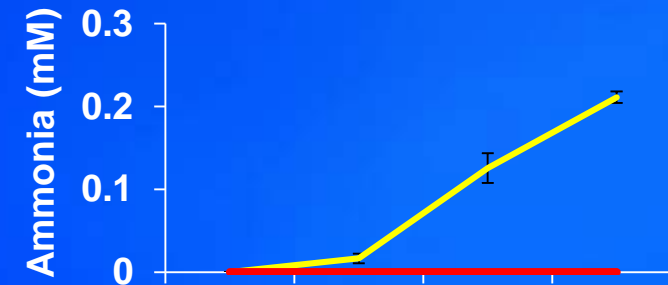
Effect sucrose concentration on pacC expression by *C. gloeosporioides*

■ 15mM Sucrose

Carbon limitation

■ 175mM Sucrose

Carbon excess



**Is this mechanism
occurring in the acidifying
postharvest pathogen
Penicillium expansum?**

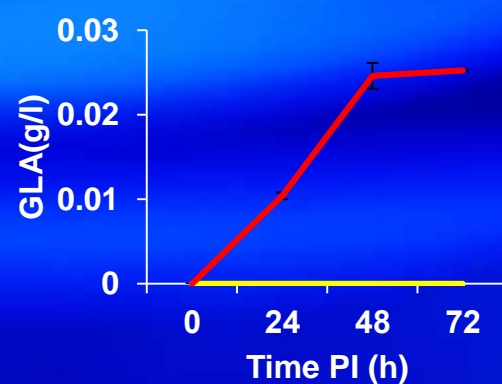
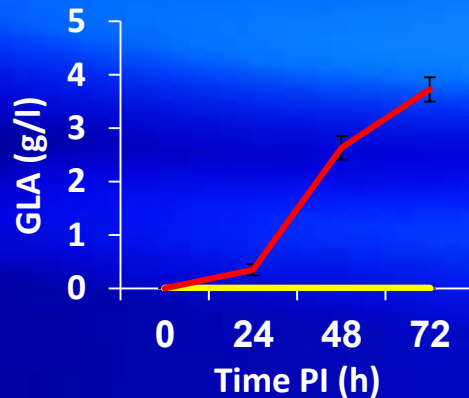
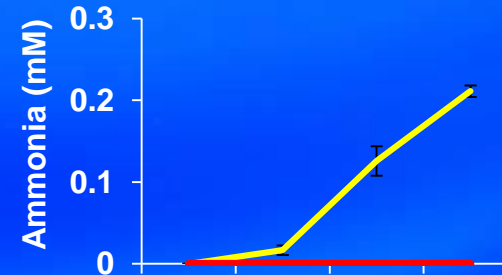
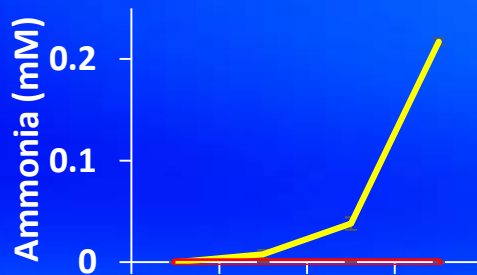
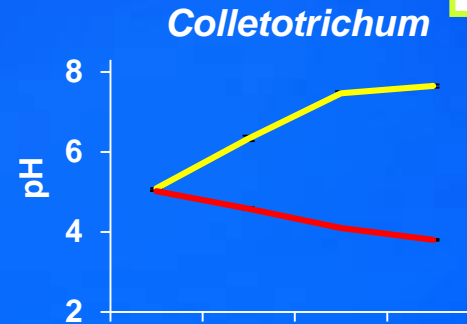
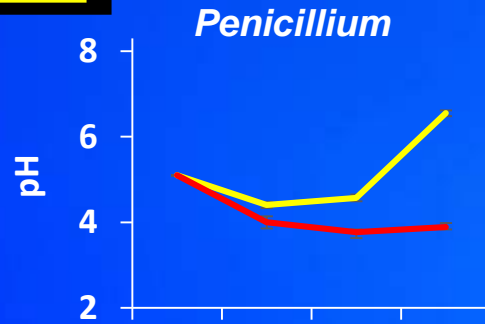
Effect of carbon on pH regulation by *Colletotrichum* and *Penicillium*

—■— 15mM Sucrose

—■— 175mM Sucrose

Carbon limitation

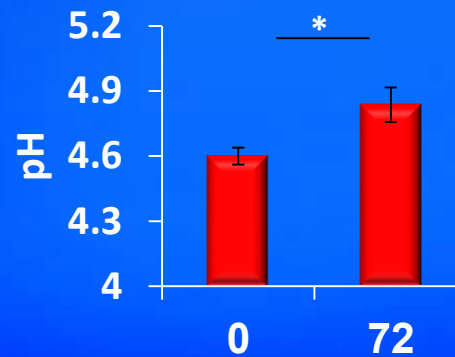
Carbon excess



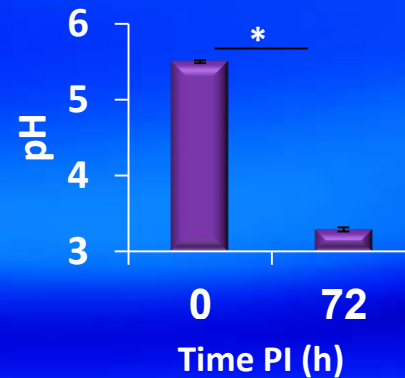
**Do we have a sugar
response present in
fruits?**

Effects of TSS on pH regulation of fruits and pathogenicity of *Colletotrichum*

Tomato
TSS 6.0

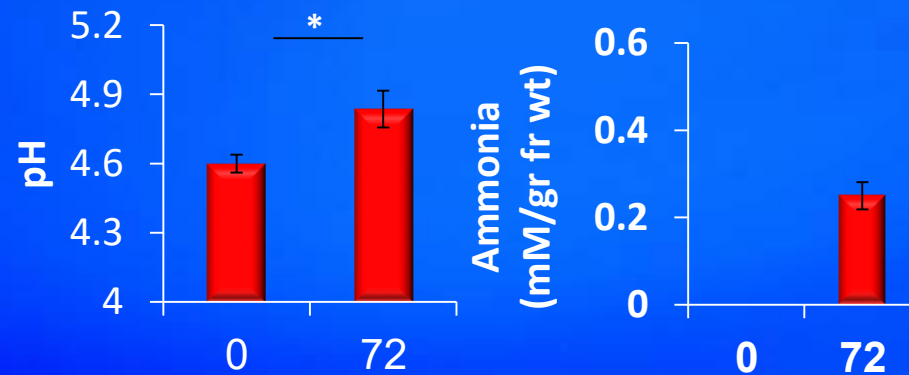


Plum
TSS 14.3

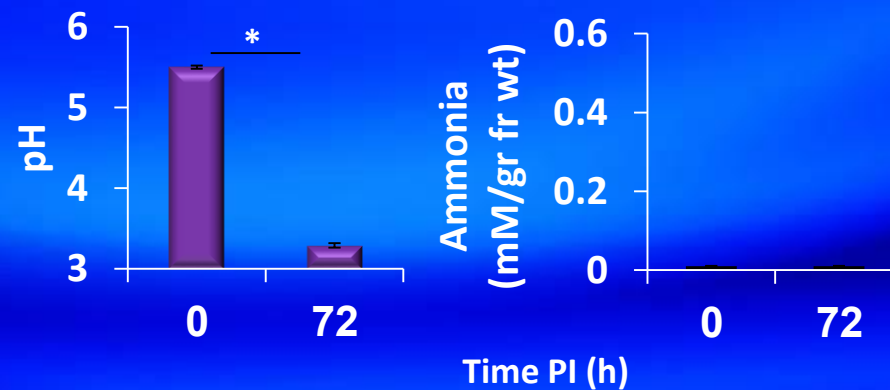


Effects of TSS on pH and ammonia regulation of fruits and pathogenicity of *Colletotrichum*

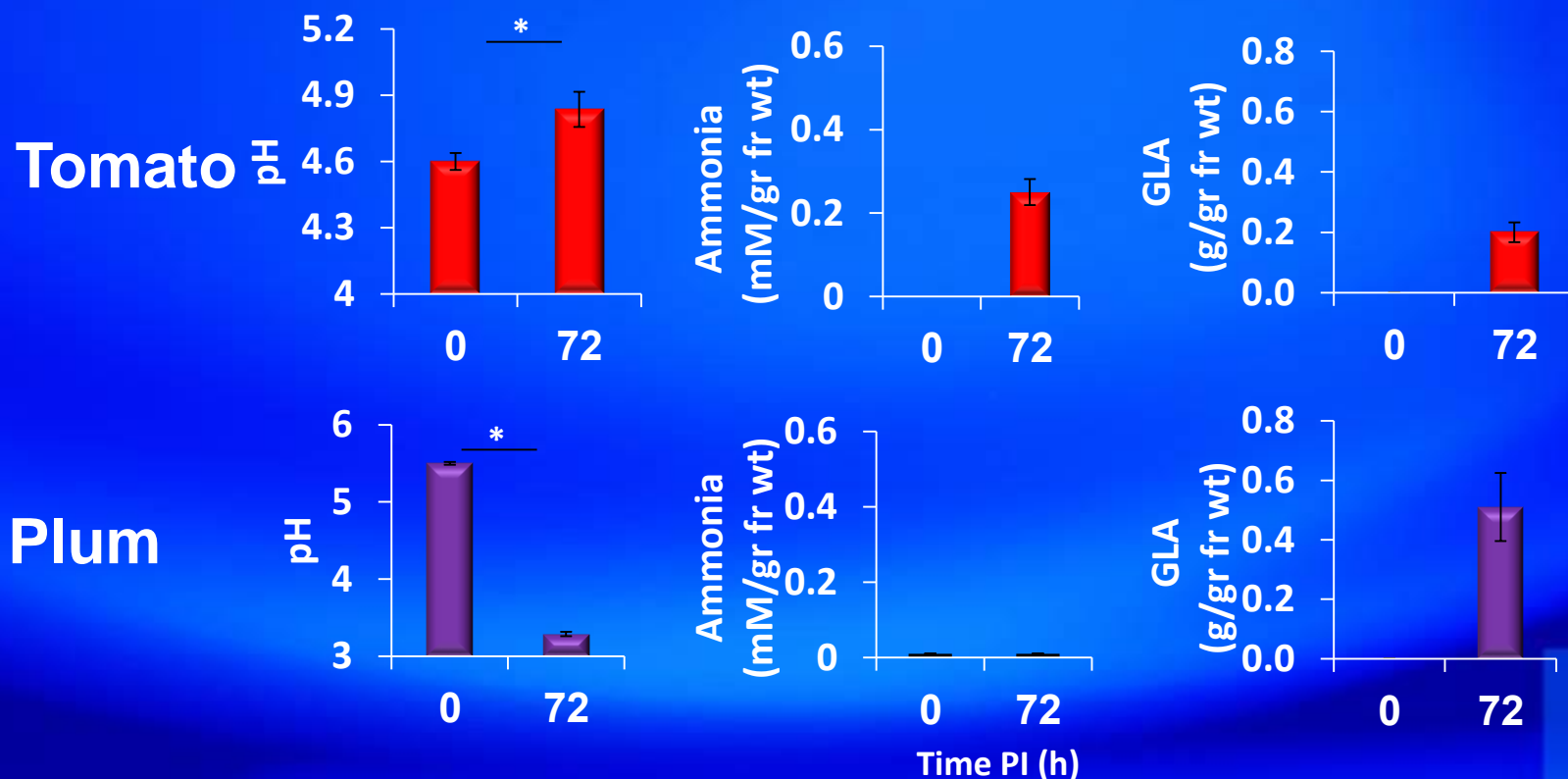
Tomato



Plum



Effects of TSS on pH, ammonia and GLA regulation of fruits and pathogenicity of *Colletotrichum*



pH modulation by pathogens (2016)

	<u>Alkalinizers</u> and <u>Acidifiers</u>
<i>Colletotrichum</i>	Ammonia and GLA
<i>Fusarium</i>	Ammonia and GLA
<i>Penicillium</i>	Ammonia and GLA
<i>Sclerotinia</i>	Ammonia and GLA

The fruit carbon content regulate the mechanism of fungal pH regulation in fruits

Fangcheng Bi, et al. 2016 Carbon regulation of environmental pH by secreted small molecule effectors modulates pathogenicity in fungi. In Press: Molecular Plant Pathology

**Low
TSS**

% TSS

**High
TSS**

Ripening process

gdh2

NH₃

Alkalinization

?

gox2

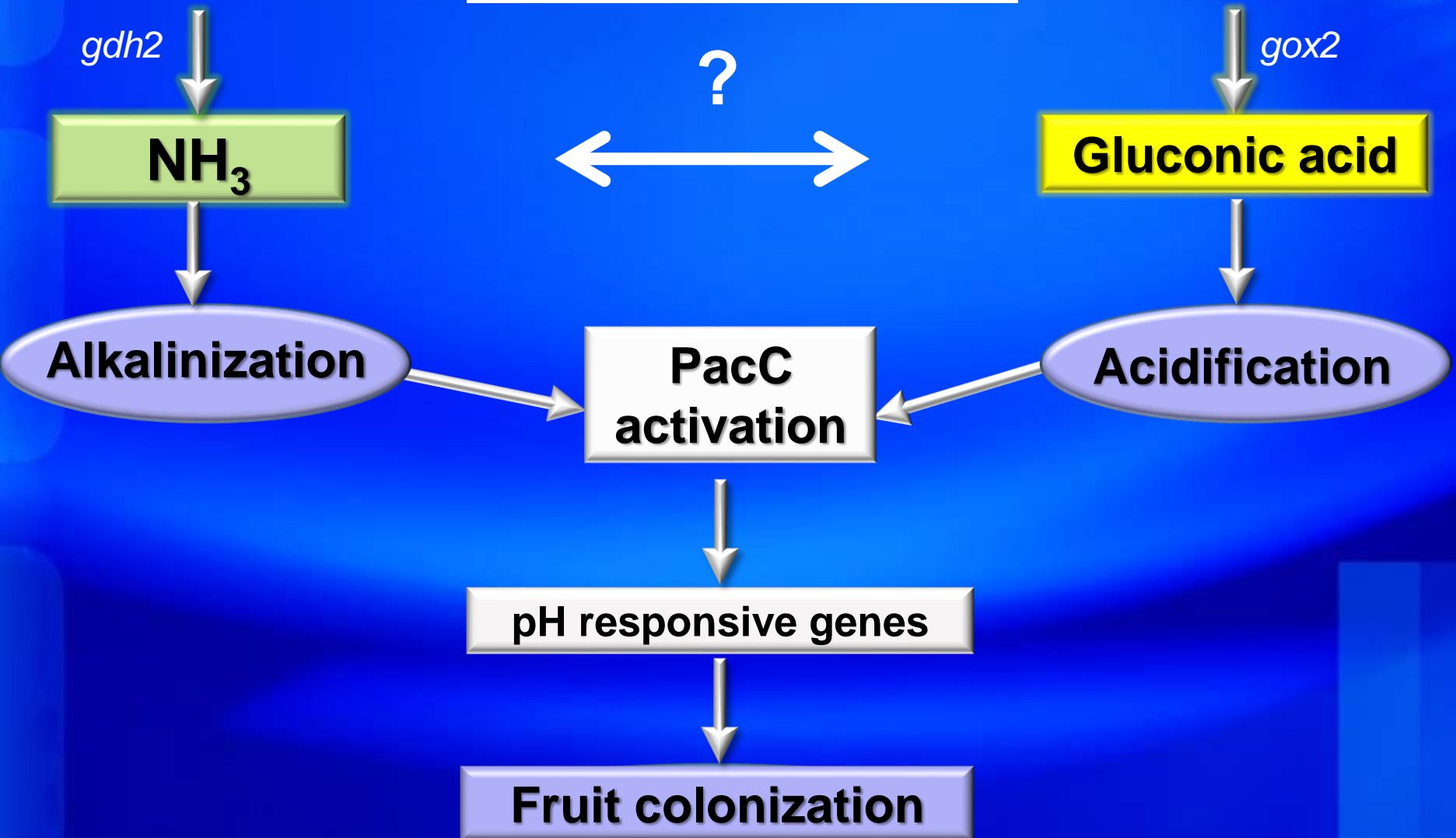
Gluconic acid

Acidification

**PacC
activation**

pH responsive genes

Fruit colonization



Summary

- Present results indicate the close relation between the accumulation of gluconic acid and pathogenicity of *P. expansum*
- Differential regulation of *gox2* might ensure tissue acidification while the contribution of *gox1* is not significant
- Initial pH is the major factor affecting glucose oxidase expression and gluconic acid production

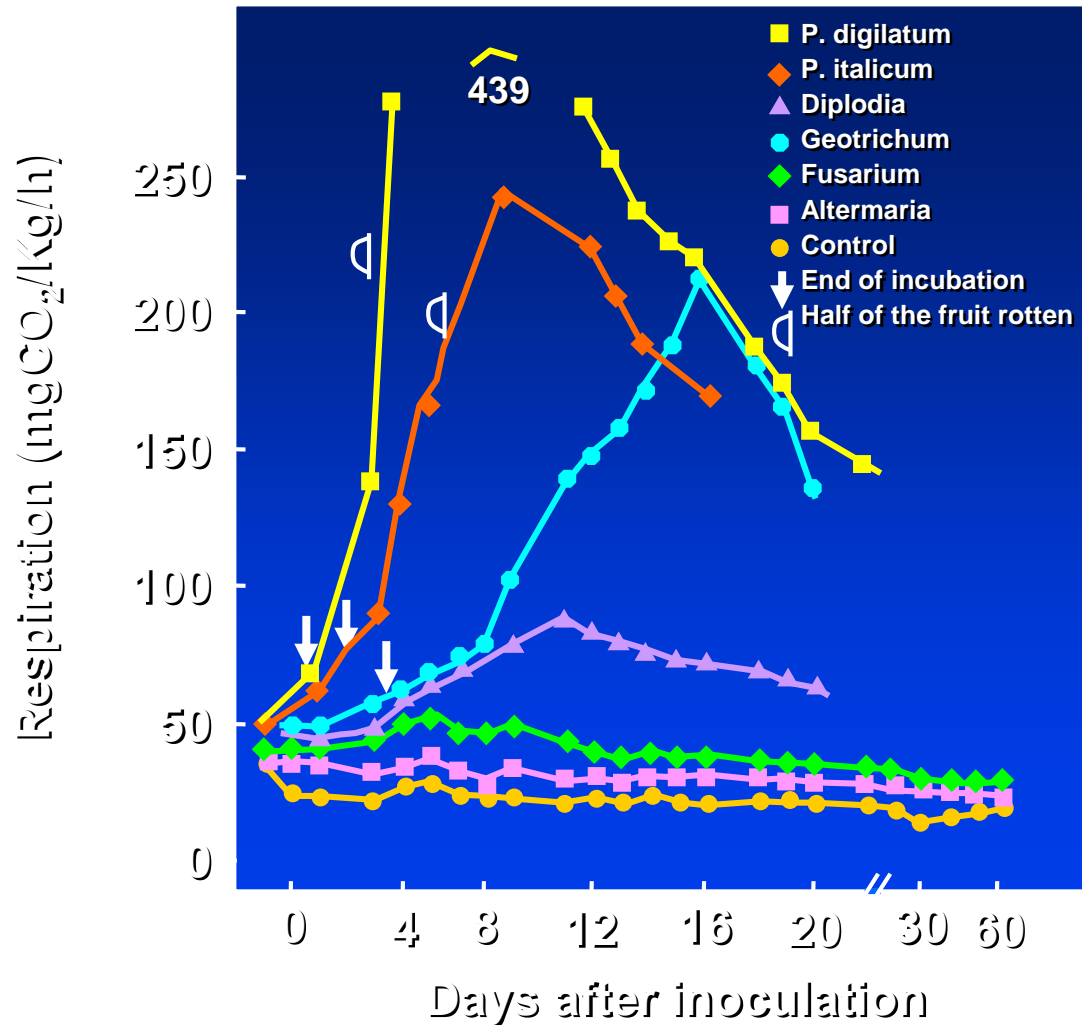
Summary of the three last lectures:

Mechanisms modulating pathogenicity in postharvest pathogens interactions

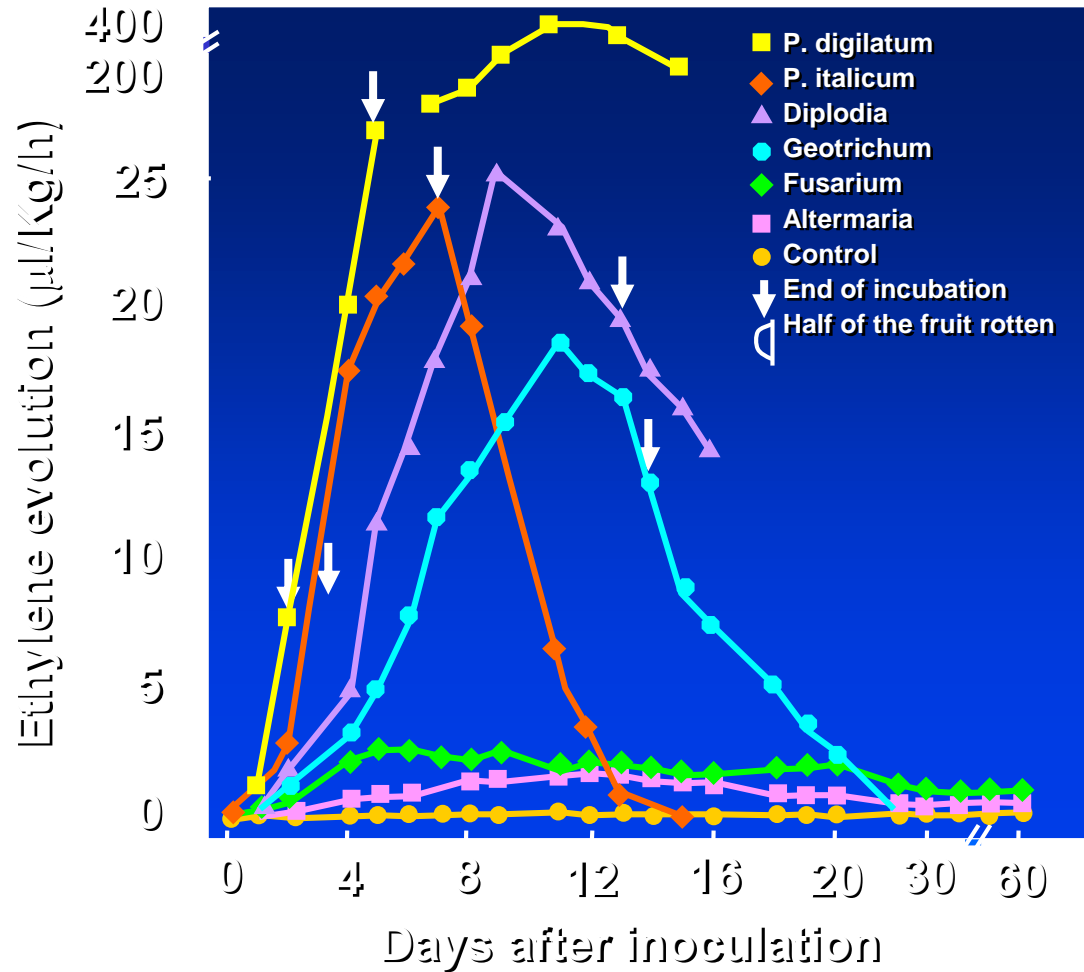
- Fungal pathogenicity is firstly determined by host resistance
- This response is mainly determined by the maturity and ripeness of the host
- The pathogen has an active function in the initiation of fungal attack by reacting to the host pH and secreting ammonia and/or organic acids leading to enhanced colonization

The physiological importance of fungal colonization on fruit physiology

Respiration rate of green lemon fruit inoculated with postharvest fungi



Ethylene evolution from green lemon fruit inoculated with postharvest fungi



Effect of fungal infection on softening and ripening

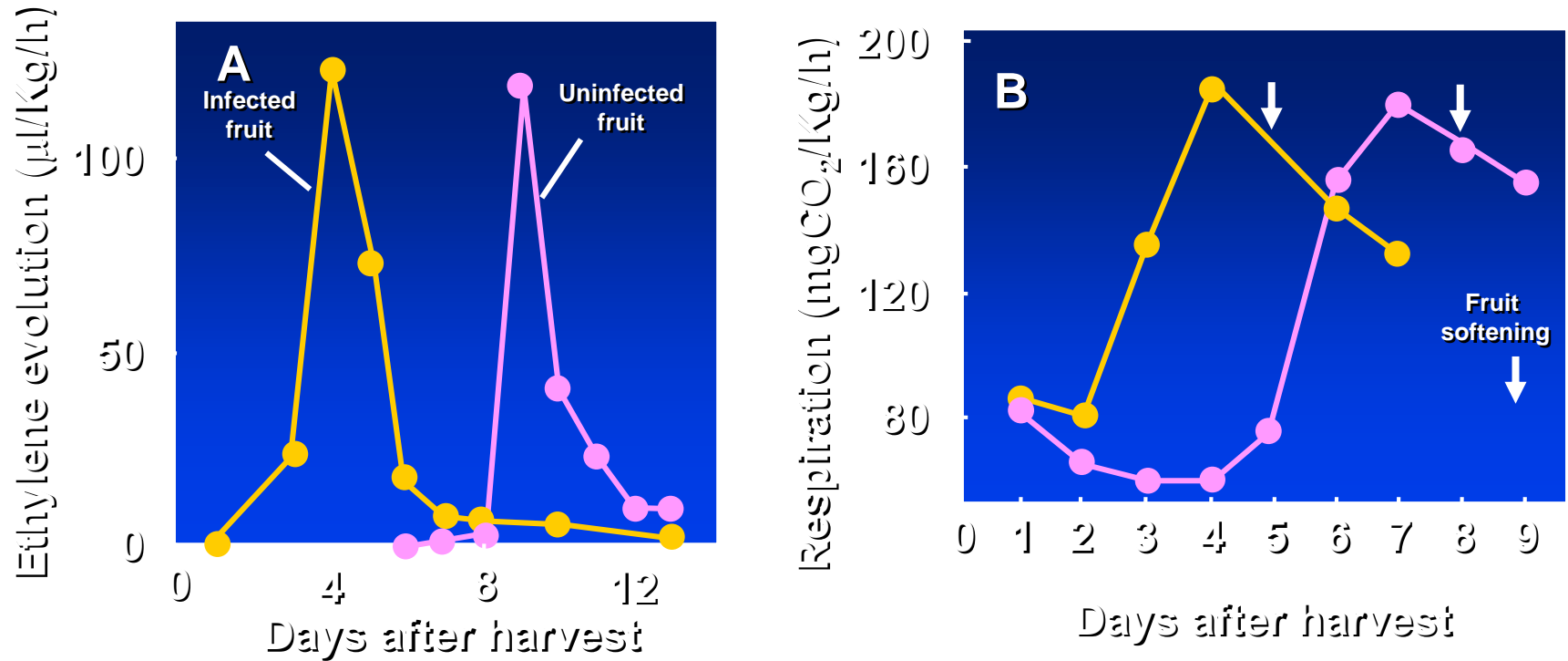


Fig. 20. Ethylene evolution (A) and respiration rates (B) of avocado fruit infected by *Fusarium solani* in comparison to uninfected fruit. (Reproduced from Zauberman and Schiffmann-Nadel, 1974 with permission of the American Phytopathological Society).